

Effects of Pollution on Marine Organisms

Alan J. Mearns^{1*}, Donald J. Reish², Philip S. Oshida³, Thomas Ginn⁴,
Mary Ann Rempel-Hester⁵, Courtney Arthur⁶, Nicolle Rutherford¹ and Rachel Pryor¹

ABSTRACT: This review covers selected 2014 articles on the biological effects of pollutants and human physical disturbances on marine and estuarine plants, animals, ecosystems and habitats. The review, based largely on journal articles, covers field and laboratory measurement activities (bioaccumulation of contaminants, field assessment surveys, toxicity testing and biomarkers) as well as pollution issues of current interest including endocrine disrupters, emerging contaminants, wastewater discharges, dredging and disposal, etc. Special emphasis is placed on effects of oil spills and marine debris due in part

to the 2010 Deepwater Horizon oil blowout in the Gulf of Mexico and the 2011 Japanese tsunami. Several topical areas reviewed in the past (ballast water and ocean acidification) were dropped this year. The focus of this review is on effects, not pollutant fate and transport. There is considerable overlap across subject areas (e.g. some bioaccumulation papers may be cited in other topical categories). Please use keyword searching of the text to locate related but distributed papers. Use this review only as a guide and please consult the original papers before citing them.

KEYWORDS: Tissue residues, toxicity, bioaccumulation, biomagnification, biomarkers, sediment quality, ecological risk assessment, endocrine disrupters, nano particles, POPs, PCBs, PAHs, PBDEs, radionuclides, pharmaceuticals, personal care products, trace metals, pesticides, biomarkers, marine biocides, oil spills, dispersants, sewage, debris, dredging, eutrophication, human disturbance, Arctic, Antarctic.

doi: 10.2175/106143015X14338845156380

^{1*} Emergency Response Division, National Oceanic and Atmospheric Administration (NOAA), 7600 Sand Point Way NE, Seattle, WA 98115; Tel.206-526-6336; email: alan.mearns@noaa.gov; ² Professor Emeritus, Department of Biological Sciences, California State University, Long Beach, CA; ³Lives in Virginia, works at the U.S. Environmental Protection Agency. ⁴Exponent, Inc, Sedona, AZ; ⁵ Partner, Aquatic Toxicology Support, Bremerton, WA.. ⁶Industrial Economics, Inc, Cambridge, MA. We thank NOAA Librarians Brian Voss and Clara Salazar for considerable help in locating papers.

Syntheses and Overviews

Understanding the effects of chronic urban contamination on the longterm productivity of anadromous fish populations has been a challenge, requiring synthesis of habitat, contaminant and fish population patterns and trends. Meador (2014) published a synthesis of information on multidecadal-long smolt-to-adult return (SAR) rates of several dozen hatchery-released Chinook salmon (*Oncorhynchus tshawytscha*) and Coho salmon (*O. kisutch*) populations, comparing return rates with physical and chemical characteristics of a wide range of urbanized to rural estuaries in southern, central and northern Puget Sound, Washington (USA). The study included existing and historical data on contaminant loadings of estuarine sediments, biota, and in salmon. Overall return rates of Chinook salmon, whose juveniles have long residence times in their estuaries, were significantly lower than for Chinook from less-contaminated rural estuaries whereas Coho salmon, whose juveniles run to sea rapidly showed no urban vs. less human-influenced differences in return rates. The author concluded that chemical contamination, marked by PCBs, has been a significant factor in the depressed Chinook return rates, additive to impacts from habitat disruption and other factors.

Michel and Rutherford (2014) conducted a detailed synthesis of historical information on the impacts and recovery rates of oiled marshes. The review covered 32 cases of oil spills field experiments finding that in most cases recovery of marsh systems occurred within two growing seasons. Recovery was slowest (many growing seasons) in marshes subject to cold climates,

heavy oiling and aggressive cleanup. Detailed response recommendations are provided.

Using worldwide tissue data collected as early as the mid-1970s, Law (2014) reviewed longterm trends of synthetic organic chemical contamination of marine mammals. Concentrations of organochlorine pesticides, PBDE's and butyltins have been slowly decreasing but also appear stalled and several are still at concentrations of considerable concern in killer whales and bottlenose dolphins. Data is insufficient to determine trends for perflorinated compounds, dioxans, furans and related compounds.

Bioaccumulation and Biomagnification

Tabular Data. Table 1 lists examples of the concentration of contaminants in tissues of marine plants and animals in 2014. Data are presented alphabetically by element then by organic compound and arranged by geographical area, concentration and author. Following are reviews of selected additional papers that report on body burdens, bioaccumulation and biomagnification.

Metals. The interspecific variations in 15 trace element accumulation in 26 species of green, brown and red seaweeds were measured in collections in the Aegean Sea (Malea et al. 2014). Brown algae showed higher concentrations of arsenic and strontium than the other elements. Finely branched seaweeds had higher concentrations of cadmium, cobalt, copper, manganese and vanadium than species with broad thalli. There were species differences, for example, the brown algal species *Cystoseira* had the highest concentration of arsenic, the red

species *Ceramium* with manganese, and the red *Polysiphonia* and green *Ulva* had the highest amounts of cadmium and selenium. It was concluded by Malea et al. (2014) that the structure of the species of algae was the determining factor in the bioaccumulation of the element. The red seaweed *Gracilaria lemaneiformis* is an important plant used for food and for production of pharmaceuticals. Laboratory experiments were conducted to measure the bioaccumulations of cadmium, copper and lead (Wang, Wang and Ke, 2014). Concentrations of copper and cadmium in tissues were higher when the two elements were together compared to the elements alone. The results indicated that bioaccumulation does occur and that cultivation of the alga should be in clean sea water.

Kalantzi and associates (2014) measured the uptake of 28 elements in polychaetes, amphipods, decapods, molluscs and echinoderms at increasing distances from fish farms in Greece. Fish were also measured for bioaccumulation of the same elements. The invertebrates accumulated lower amounts of most elements than those measured in the sediments except arsenic, phosphorus, sodium, cadmium and zinc. Tolerant species, such as most of the species of polychaetes, accumulated higher concentrations of many of the 28 elements studied than the more sensitive species in different animals groups. Biomagnification of mercury was noted in fish feeding on benthic invertebrates in the vicinity of the fish cages.

A stable nitrogen isotope was used to document the biomagnification pathway of mercury and selenium occurs in fish (Jones et al. 2014). Results indicated that mercury but not selenium was taken up by fish feeding

upon the animals from the local area. A risk assessment study was conducted in a marine bay in China to determine if butylin used in biofouling control would bioaccumulate in organisms used for food. It was found that few if any harmful compounds would occur in humans from consumptions of the cultured fish (Hu et al., 2014). Sublethal concentrations of cadmium, lead and a combination of the two elements were measured on mussel gill tissue (Poynton et al. 2014). Results of gene ontology identified several biological processes affected including nucleotide phosphate synthesis. These responses served as early indicators of stress which can lead to adverse physiological effects (Poynton et al., 2014).

Nanoparticles. The fate and effects of nanoparticles on a clam and polychaete were reviewed by Mouney et al. (2014). Data were analyzed to determine which route of exposure influenced bioaccumulation. Soluble nanoparticles attached to seawater molecules which were taken up in both species. The effect of cadmium sulfide nanoparticles on the biochemical and behavior responses to the polychaete *Hediste diversicolor* was measured by Buffet et al. (2014a). Both solutions of cadmium and cadmium nanoparticles were accumulated by the polychaete. Behavior was measured by the number of body undulations. Body movements were impaired when exposed to cadmium nanoparticles compared to control. An experimental study with mussel, *Mytilus galloprovincialis*, was conducted by Balbi et al. (2014) to determine if nanoparticles, as n-TiO₂, and cadmium together would induce measurable stress responses (biomarkers) without toxicity. The co-exposure of n-TiO₂

and cadmium resulted in effects on lysosomal biomarkers and embryonic development. However the toxicants did not result in an increase in adverse effects.

Tian et al. (2014) studied the impact of titanium dioxide nanoparticles in the presence and absence of phenanthrene under laboratory conditions. Bioaccumulations of $nTiO_2$ occurred in the presence of phenanthrene but not by itself. This difference is explained by the ingestion of the large molecule of $nTiO_2$ - phenanthrene complex by the clam. The toxicity and bioaccumulation of citrate and polyvinylpyrrolidones coated with silver nanoparticles were investigated using amphipods, mysids and polychaetes (Wang et al. 2014). These compounds were not toxic to the amphipods and mysids after a seven day exposure. However bioaccumulation occurred in the polychaete *Nereis virens*.

Synthetic Organic Chemicals. The uptake of PCBs and PBDEs in benthic fauna was measured near two marine sewage outfall, urban harbors, and a reference site (Burd et al. 2014). PBDEs were more readily taken up by biota and PCBs. However organic enrichment near waste discharges enhanced the uptake of PCBs (Burd et al. 2014). Accumulations of endocrine disrupting chemicals (EDCs) was measured in the polychaete *Paraprionospio* sp. from Osaka Bay, Japan (Nurulnadia et al., 2014). The polychaete accumulated varying concentrations of nonylphenol from 1.46 to 4.41 $\mu\text{g/g}$ wet weight. Concentrations were greater in the polychaete than sediment indicating bioaccumulation in the worms (Nurulnadia et al., 2014).

An evaluation was made of the effect of diuron, a photosynthetic inhibitor, used in sugar fields near the Great Barrier Reef, Australia (Holms et al. 2014). In a 12 year review it showed that this pesticide affected coralline algae and photosynthetic cells in corals.

Handoh and Kawai (2014) devised a model to determine mass mortality of marine mammals. The mortality data on 33 species of marine mammals were compiled and analyzed with respect to the global concentration of PCBs in fish. Based on this model, hot spots were identified for marine mammal mortality as the Mediterranean Sea and north-western Europe. See Law (2014) for additional information on this subject.

Hydrocarbons. A laboratory experiment was employed to determine if three species of fish, one decapod crustacean and a mussel ingested oil droplets (Viaene et al., 2014). Selection of the test organism was based on feeding method: carnivores or filter feeders. Only the mussel took in the droplets indicating that filter feeding was the determining factor (Viaene et al., 2014). The benthic ecosystem in the Chukchi Sea was reviewed because of the potential for oil and gas exploration. The concentration of aliphatic hydrocarbons, polycyclic hydrocarbons, and 17 metals were at background levels except where drilling was done earlier (Dunton et al. 2014). There was no evidence of bioaccumulation of these substances above background levels in biological benthic biomonitors such as crustaceans and echinoderms (Dunton et al., 2014).

Radionuclides. The uptake of polonium-210 was measured in muscle tissues and organs in predatory

fish taken off Río de Janeiro by Mársico et al. (2014). There were significant differences in the ^{210}Po in the two species of fish. There were also differences of radioactivity in the different organs with the highest concentrations in the digestive system. A radionuclide transfer model employed the known data on uptake of inorganic in marine organisms for use in a planned disposal site in the Baltic Sea (Konovalenko et al., 2014). The risk assessment utilized data known for grazers, benthos, zooplankton and fish. It showed that the data on organisms is in good agreement, but many elements (i.e. 26) need to be used in addition to just cesium and strontium (Konovalenko et al., 2014).

The bioaccumulation of eight radioactive elements was conducted under laboratory conditions using the red alga *Polysiphonia fucooides* (Zalewska, 2014) After the initial exposure, additional uptake was linear regardless of the concentration of the isotope. Once the maximum concentration was reached, a significant decline was noted in the isotopes of manganese, cobalt, cesium and americium (Zalewska, 2014).

Chemical Toxicity and Testing

Reviews below are organized by media or contaminant. Reviews of oil and dispersant toxicity follow oil spill sections at end of this paper.

Scientists continued to adapt existing methods and develop new methods to improve capabilities to quantify, analyze, and determine toxicity of various chemicals in the estuarine and marine organisms.

New Methods and Endpoints. Howe et al. (2014a) described an 8-d sublethal bioassay with sea anemone (*Aiptasia pulchella*) pedal lacerates. They exposed the pedal lacerates to Cd, Co, Cu, and Zn and observed effects on development to the juvenile stage (EC_{50} s were 55 $\mu\text{g/L}$, 262 $\mu\text{g/L}$, 5 $\mu\text{g/L}$, and 269 $\mu\text{g/L}$ for Cd, Co, Cu, and Zn, respectively). Howe et al. (2014b) used 28-d sublethal tests that observed asexual reproduction of symbiotic sea anemones (*Aiptasia pulchella*) to assess the toxicity of metals. They found 50% reductions in asexually reproduced juveniles (EC_{50}) at 14 $\mu\text{g/L}$ for Cu, 63 $\mu\text{g/L}$ for Zn, and 107 $\mu\text{g/L}$ for Ni. Faimali et al. (2014) investigated the use of ephyra of jellyfish (*Aurelia aurita*) as a model organism. They examined the influence of different culture parameters (e.g., temperature, photoperiod) on behavioral endpoints and exposed the organisms to Cd nitrate and sodium dodecyl sulfate (SDS). They thought the ephyrae of jellyfish showed promise as experimental organisms. Renzi et al. (2014) exposed diatoms (*Phaeodactylum tricorutum*) to Zn, Cu, and dodecylbenzenesulfonic acid sodium salt in accordance with the AlgalToxkit protocol to quantitatively compare responses with growth rate inhibition tests morphological (biovolume) and physiological (chlorophyll-*a*, phaeophytin ratio) endpoints. The results showed that both biovolume and the photosynthetic complex are sensitive sub-lethal endpoints of exposure.

In an effort to improve the medaka (*Oryzias latipes*) embryo-larval sediment contact assay, Le Bihanic et al. (2014) developed a reference exposure protocol with artificial sediment that was specifically designed to limit

natural sediment composition uncertainties and preparation variability. They tested and validated the artificial sediment using fluoranthene, benz[α]anthracene, and benzo[α]pyrene and observed various developmental end points.

Deep Sea Biota. Concerned with the advent of industrial activities in the deep sea, Mestre et al. (2014) described and emphasized the need for environmental risk assessments based on information generated from ecotoxicological trials that mimic, as close as possible, the deep sea environment, with emphasis to high hydrostatic pressure.

Metals. Oliveira et al. (2014) used solid phase sediment bioassays with copepods *Tisbe biminiensis* to assess the toxic effects of the sediments and trace metal analyses to measure trace metal concentrations (Cr, Zn, Mn, Fe, Cu, Pb, Co, and Ni) in the sediments from an urban mangrove patch in Brazil. They reported lethal and sublethal effect concentrations. Taylor and Maher (2014) used microcosms in a 28-d exposure of bivalves (*Tellina deltoidalis*) to selenium-spiked sediments (0, 5 $\mu\text{g/g}$, and 20 $\mu\text{g/g}$), and observed reduced antioxidant capacity, which corresponded with increased lipid peroxidation, lysosomal destabilization, and micronuclei frequency. Exposure-dose-response relationships were demonstrated for *T. deltoidalis* exposed to selenium spiked sediments. Dolores-Basallote et al. (2014) exposed amphipods (*Ampelisca abdita*) to Gulf of Cadiz sediments that had been subjected to several pH treatments to study the effects of CO₂-induced acidification on sediment toxicity. The results showed that CO₂-related acidification lead to lethal effects in amphipods and mobility of metals. Canário et al. (2014)

established baseline sediment toxicity levels by testing and analyzing Canadian Arctic and sub-Arctic sediments collected in 2005 from eight locations. Sediment elutriates were tested with ARTOXKIT M, Microtox liquid phase, and ROTOXKIT M toxicity assays, and whole sediment was tested with the Microtox solid phase assay procedure. The sediments were considered nontoxic, although some measures of toxicity were measurable. Hoang and Rand (2014) conducted short term and longer term whole sediment acute toxicity studies with amphipods (*Ampelisca abdita*) and clams (*Mercenaria mercenaria*) to examine seven field collected sediment samples from the St. Lucie River (Florida) and found no significant effect on survival and an inverse relationship between dry weight of the tested organisms and Cu and Zn concentrations in sediments and organisms.

Hariharan et al. (2014) conducted acute and chronic toxicity tests to examine the adverse effects of Pb on mussels (*Perna viridis*). In the chronic exposure, they correlated Pb concentrations with adverse effects as shown in changes in biochemical enzymes, catalase, glutathione, glutathione *S*-transferase, and lipid peroxides.

Anti Fouling Biocides. Several scientists investigated anti-fouling compounds, aquaculture pesticides, and other pesticides. Fox et al. (2014) exposed 7-d old amphipod (*Corophium volutator*) neonates to medetomidine, an anti-fouling compound, in 28-d and 76-d sediment studies. They observed mortality at 32 $\mu\text{g/kg}$. Arrhenius et al. (2014) described a bioassay that examined the effects of biocides on the initial settling and establishment of photoautotrophic biofilms, including

indigenous fouling organisms. Using this bioassay, Arrhenius et al. (2014) ranked the efficacy (based on EC₉₈ values) of select antifoulants from most to least efficacious: Cu pyrithione> TPBP (triphenylborane)> DCOIT (4,5-Dichloro-2-octyl-1,2-thiazol-3-one)> tolyfluanid> Zn pyrithione> medetomidine> Cu²⁺. Bao et al. (2014) examined the combined toxicity of zinc pyrithione and Cu on copepods (*Tigriopus japonicus*) using 96-h acute toxicity tests with adult copepods and 21-d chronic full life-cycle tests with nauplii. Zinc pyrithione has been reported to be easily trans-chelated to Cu pyrithione in the presence of Cu. Their results showed that Zn pyrithione and Cu exhibited strong synergistic toxic effects in both acute and chronic tests.

Li et al. (2014) examined the toxicity-temperature relationship using 96-h LC₅₀ tests with fish (*Oryzias melastigma*) and copepods (*Tigriopus japonicus*) and the 24-h LC₅₀ test with rotifers (*Brachionus koreanus*). The toxicants were Cu and three biocides [dichlorophenyltrichloroethane (DDT), triphenyltin chloride (TPTCl), and copper pyrithione (CuPT)]. The toxicity for fish peaked at 20°C for Cu and TPTCl and at 25°C for DDT and CuPT. The LC₅₀ values for copepods and rotifers showed an inverse relationship with temperature across all the toxicants used. Yi et al. (2014) examined the effects of triphenyltin chloride on molecular, individual, and population responses of copepods (*Tigriopus japonicus*). In a full life-cycle exposure, 1.0 µg/L elicited a delay in development and a reduction in population growth. At 0.1 µg/L, the sex ratio changed to a male-biased population. They also observed the inhibition

of transcriptional expression of glutathione S-transferase genes and inhibition of retinoid X receptor mRNA expression.

Wang, Shi, Yang, Han and Zhou (2014) used fish (*Brachydanio rerio*) and green algae (*Selenastrum capricornutum*) in acute and growth inhibition tests, respectively, to assess capsaicin (an active substance for ship anti-fouling systems). They calculated the Predicted No Effect Concentration (PNEC) to be 4.9 x 10⁻⁴ mg/L, which was one to two orders of magnitude above the average PEC for OEECD-EU commercial harbors and marinas, respectively.

Wang, Zhao, Yang, Han, Long and Zhou (2014) used available data to develop risk assessments for active anti-fouling substances used in China.

Pesticides. The toxicity of four anti-lice pesticides, used in salmon aquaculture (deltamethrin, cypermethrin, hydrogen peroxide, and azamethiphos), to copepods was examined in 1-h exposures with lethality and feeding endpoints determined 5-h post-exposure using staining techniques. EC₅₀s ranged from 30- to 117-fold, 13- to 51-fold, and 120- to 460-fold dilutions of the respective aquaculture treatments for deltamethrin, cypermethrin, and hydrogen peroxide, respectively. No effects on copepods were observed at 5-times the aquaculture treatment concentrations for azamethiphos (Van Geest et al., 2014a). Van Geest et al. (2014b) used the amphipod *Echinogammarus finmarchicus* to test the toxicity of the pyrethroid-based anti-sea lice pesticides, deltamethrin and cypermethrin in aqueous and sediment exposures. In 1- and 24-h water-only exposures, mortality

and immobility were observed at 6.7-70 ng/L and 20-220 ng/L for deltamethrin and cypermethrin, respectively, suggesting that amphipods could be exposed to concentrations of pyrethroids sufficient to cause adverse effects in effluent plumes from salmon aquaculture where treatment for ectoparasitic sea lice is occurring. Tucca et al. (2014) examined the effects of three antiparasitic pesticides (cypermethrin, deltamethrin, and emamectin benzoate) on amphipods (*Monocorophium insidiosum*) in 10-d whole sediment bioassays. They observed effects on glutathione *S*-transferase and thiobarbituric acid reactive substances. *M. insidiosum* was sensitive to antiparasitic pesticides concentrations at $\mu\text{g}/\text{kg}$ in sediments. Van Geest et al. (2014c) examined the effects of the anti-sea lice pesticide deltamethrin on *Nereis virens* (polychaete) exposed via water and sediments. Sublethal effects related to burrowing behavior and worm condition were observed at 11 $\mu\text{g}/\text{g}$. DeLorenzo et al. (2014) exposed 7-d old mysids (*Americamysis bahia*) and larval and adult shrimp (*Palaemonetes pugio*) to pyrethroid insecticides (lambda-cyhalothrin, permethrin, cypermethrin, deltamethrin, and phenothrin) in 96-h LC_{50} bioassays. Acute toxicity occurred at low nanogram per liter concentrations for some of the pyrethroids tested.

Herbicides. Yusof et al. (2014) exposed fertilized fish eggs (*Oryzias javnicus*) to high concentrations (100 to 500 ppm) of a glyphosphate-based herbicide (Roundup®). Survival and hatching percentages decreased as glyphosphate concentrations increased. Teratogenic effects were observed. Hwang et al. (2014) examined the toxicity of triclosan to gamete viability,

fertilization, and embryogenesis in sea urchins (*Strongylocentrotus nudus*). The LOEC for embryogenesis was 0.33 μM .

Personal Care Products. There is continued concern about the effects of personal care and health products on marine organisms. Several investigators examined the effects of antibiotics, antidepressants, and beta blockers. Seoane et al. (2014) investigated the toxic effects of three antibiotics (chloramphenicol, florphenicol, and oxytetracycline) on algae (*Tetraselmis suecica*) and found that photosynthetic parameters, such as chlorophyll a cellular content and auto fluorescence, were altered after 24 h of antibiotic exposure. Gomiero and Viarengo (2014) investigated the effects of Cu and oxytetracycline, separately and in combination, at different temperatures on the survival, replication, endocytosis rate, and lysosomal membrane stability in the ciliated protozoa *Euplotes crassus*. The two toxicants elicited very different and complex trends under the various thermal conditions. Johansson et al. (2014) examined the toxicity of two antibiotics, ciprofloxacin and sulfamethoxazole, to the bacterial and algal species on natural biofilms from the coast of Sweden. They used a 4-d semi-static system to expose the biofilms, and found that sulfamethoxazole exposure leads to a general decrease in carbon source utilization and ciprofloxacin exposure leads to a rearrangement of the carbon-utilization pattern in the bacteria species

Minguez et al. (2014) assessed the ecotoxicity of three antidepressants (fluoxetine, sertraline, and clomipramine) using a battery of tests/species [algal

growth inhibition (*Skeletonema marinoi* and *Pseudokirchneriella subcapitata*), crustacean immobilization (*Artemia salina* and *Daphnia magna*), development and adult survival (*Hydra attenuate*), oyster embryogenesis and metamorphosis (*Crassostrea gigas*), and *in vitro* assays on abalone hemocytes (*Haliotis tuberculata*). EC₅₀ values ranged from 43 to 15,600 µg/L for fluoxetine, 67 to 4,400 µg/L for sertraline, and 4.70 to 100,000 µg/L for clomipramine. The algae and embryolarval stages of the oyster were the most sensitive of the taxa tested. Maszkowska et al. (2014) conducted ecotoxicological evaluations on three beta-blockers (propranolol, metoprolol, and nadolol) using bacteria (*Vibrio fischeri* and *Arthrobacter globiformis*), algae (*Scenedesmus vacuolatus*), and duckweed (*Lemna minor*). They found that propranolol and metoprolol were harmful to algae.

Paredes et al. (2014) examined the toxic effects of four chemical UV filters (4-Methylbenzylidene-camphor, Benzophenone-3, Benzophenone-4, and 2-Ethylhexyl-4-methoxycinnamate) on algae (*Isochrysis galbana*), mussels (*Mytilus galloprovincialis*), sea urchins (*Paracentrotus lividus*) and mysids (*Siriella armata*). They found 2-Ethylhexyl-4-methoxycinnamate and 4-Methylbenzylidene-camphor to be the more toxic of the chemicals tested, with the algae being the most sensitive of the tested species

Multiple Products. Fabbri et al. (2014) demonstrated that acute 48-h mussel (*Mytilus galloprovincialis*) toxicity tests can be used to evaluate the effects of different model compounds representative of endocrine disrupting chemicals (nonylphenol, bisphenol

A), brominated compounds (tetrabromobisphenol A), perfluorinated compounds (perfluorooctanoic acid and perfluorosulphonate), and pharmaceuticals (ibuprofen, diclofenac, and benzafibrate) in a wide concentration range (0.01 – 1000 µg/L). Rhee, Jeog, Kim and Lee (2014) reported inhibition of growth of *Barachionus koreanus* to a suite of pharmaceuticals.

Household Cleaning Products. The 96H LC₅₀s of eight household cleaning products, ranging from baby shampoo to dishwashing detergents, to mysid shrimp (*Americamysis bahia*) and silversides fish (*Menidia beryllina*) was reported by Word et al. (2015). For the fish, 96H LC₅₀s ranged from 5.4 to 592 mg/L and for the mysid shrimp from 12.4 to 413 mg/L. Liu and Zhang (2014) experimentally compared 25 halophenolic and haloaliphatic disinfection byproducts using algae (*Tetraselmis marina*) as the test organisms. Halophenolic disinfection byproducts were generally more toxic than the commonly known haloacetic acids.

Nano Materials. Choi et al. (2014) used algae (*Pseudokirchneriella subcapitata*), daphnia (*Daphnia magna*), and bacteria (*Vibrio fischeri*) to examine the toxicity of water-solubilized aminoclay nanoparticles. The EC₅₀ for algal growth was 1.29 mg/L and 0.26 mg/L for cell yield, while the NOEC for *V. fischeri* was 25,000 mg/L and the LOEC (48-h) for *D. magna* was 100 mg/L. Castro-Bugallo et al. (2014) examined the effects of two metal oxide nanoparticles [zinc oxide (ZnO) and yttrium oxide (Y₂O₃)] on three microalgae (*Phaeodactylum tricoratum*, *Alexandrium minutum*, and *Tetraselmis suecica*). They tested the effects on growth, carbon content, carbon-to-

nitrogen ratio, chlorophyll fluorescence, and reactive oxygen species production. Pronounced differences were observed between species to the test materials highlighting the importance of analyzing diverse groups of microalgae and various physiological effects. Park et al. (2014) assessed the effects of salinity (5‰ - 35‰) on Cu, Zn, and CuO nanoparticles and ZnO nanoparticles on copepods (*Tigriopus japonicus*) in 96-h exposures. They observed no differences in acute toxicity at different salinities. Wang et al. (2014) investigated the toxicity, bioaccumulation, and biotransformation of citrate and polyvinylpyrrolidone coated Ag nanoparticles in *Nereis virens* (polychaete), *Ampelisca abdita* (amphipod), and *Americamysis bahia* (mysid) via marine sediment exposure. The results showed that the silver nanoparticle surface capping agents influenced Ag uptake, biotransformation, and/or excretion, and demonstrated the accumulation and speciation of Ag nanoparticles in *Nereis virens*. Minetto et al. (2014) discussed the complexity of experimental exposure scenarios for titanium oxide nanomaterials in the marine environment: providing statistical information about different matrices, organisms, and nanomaterials, and comparing ecotoxicity effects of titanium oxide nanomaterials.

Other Commercial Compounds. Ventura et al. (2014) provide ecotoxicological data for ten cholinium-based salts and ionic liquids (cholinium bicarbonate, cholinium bitartrate, cholinium chloride, cholinium acetate, cholinium dihydrogenophosphate, cholinium dihydrogenocitrate, cholinium salicylate, benzyltrimethyl(2-hydroxyethyl)ammonium chloride,

cholinium propanoate, and cholinium butanoate) to gain insight on the toxicity mechanism of these compounds. They found that not all the cholinium tested is harmless to bacteria (*Vibrio fischeri*), and that the cholinium exhibits a different mechanism of toxicity compared to the imidazolium ionic liquids previously described.

Sediment Quality Guidelines. Kwok et al. (2014) summarized the key findings of an expert group convened to discuss the important scientific and regulatory challenges with developing sediment quality guidelines. They also identified areas of scientific research that are needed to improve sediment quality assessment. Diepens et al. (2014) produced a review paper that describes sediment toxicity tests for microorganisms, macrophytes, benthic invertebrates, and benthic communities. They presented recommendations and identified knowledge gaps and priorities for further research. Field and Norton (2014) describe approaches to improve the performance of empirical models developed from a large nationwide data set to predict sediment toxicity from chemistry from regional applications. Their study suggests that calibrating nationwide models to a regional data set may be a more efficient and effective approach for improving model performance than developing region-specific models.

PAHs. Mu et al. (2014) exposed early life stages of fish (*Oryzias melastigma*) to phenanthrene and retene (7-isopropyl-1-methylphenanthrene) and found retene to be more toxic. Both phenanthrene and retene caused developmental malformation of embryos with phenanthrene affecting the peripheral vascular system and retene affecting cardiac tissues. Wang, Wang, Mu, Wang,

Yao and Lin (2014) used existing toxicity data for Chinese marine species to derive predicted no-effect concentrations for three polycyclic hydrocarbons (phenanthrene, pyrene, and benzo(a)pyrene). Pastore et al. (2014) examined the kind of nuclear damage found in fish (*Sparus aurata*) hepatocytes continuously exposed to sublethal doses (10 µg/mL – 1pg/mL) of benzo[α]pyrene (B[α]P) for 24 and 72 h. They observed apoptosis induction and nuclear abnormalities by immunofluorescence analysis and found no evidence of a threshold dose below which B[α]P was found not to be genotoxic in sea bream cultured hepatocytes.

Biomarkers

Biomarker Reviews. Several biomarker reviews were published in 2014. Regoli and Giuliani (2014) wrote a very useful review of oxidative stress biomarkers in marine organisms. They covered what each biomarker does, how it is regulated, and how it tends to respond to given chemical pollutants. They also discussed the reasons that researchers often find mRNA levels do not correspond to catalytic activities of antioxidant enzymes, including differences between transcriptional and post-translational regulation. Bolognesi and Cirillo (2014) reviewed the use of the comet assay and micronucleus test to measure genotoxicity in *Mytilus* spp. They found that both methods were useful, and they demonstrated differing sensitivity to contaminant classes, possibly because the comet assay provided detection of recent exposure, whereas the micronucleus test could reveal accumulated damage during the lifespan of the organism. It was important to take into account the effects

of other factors besides chemical pollution, and they recommended using large sample sizes, caged mussels, standardized deployment and sampling protocols to minimize variability in response. A review of environmental stress proteomics studies by Tomanek (2014) found that comparing responses of closely related species to population responses provided information on the cellular processes that set the environmental tolerance limits of a given species. Rodrigues and Pardal (2014) reviewed studies with the green crab *Carcinus maenas* and developed a list of the most suitable biomarkers to detect response to classes of contaminants, for example the use of the ethoxyresorufin-O-deethylase (EROD) assay with hepatopancreas tissue for polycyclic aromatic hydrocarbon (PAH) and polychlorinated biphenyl (PCB) exposure. They found that confounding factors for data interpretation included gender, size, morphotype, nutritional status, historical contaminant exposure, temperature and salinity. Lehtonen et al. (2014) presented recommended biological effects indicators for the Baltic Sea based on several region-wide studies. These included lysosomal membrane stability (LMS) in fish, bivalves or amphipods to measure general stress caused by a range of contaminants; induction of micronuclei in fish, bivalves or amphipods for effects of genotoxic contaminants; embryo aberrations in eelpout or amphipods for reproductive success impairments; fish disease index based on externally visible fish disease, macroscopic liver neoplasms and liver histopathology for general health status; imposex in gastropods for tributyltin effects; and PAH metabolites in fish for PAH exposure. Other methods were placed on the candidate list, including

intersex or vitellogenin (VTG) in male fish to measure endocrine disruption, acetylcholinesterase (AChE) activity for neurotoxicity, and EROD for biotransformation.

Field Studies: Molluscs. Tsangaris et al. (2014) placed caged mussels (*Mytilus galloprovincialis*) for one month periods both at and near a disposal site for dredged river sediment in Greece. They found that sediment contaminant loads and Microtox® toxicity were highest at a location within the dumping zone and persisted even after dumping was terminated. However, PAH and aliphatic hydrocarbon levels in mussel tissues were elevated at all sites, even one outside of the immediate dumping zone, and the elevated contaminant load was accompanied by significant decreases in condition index (CI), scope for growth (SFG), glutathione-S-transferase (GST) activity, and catalase (CAT) activity but had no effect on lipid peroxidation (LP), metallothionein (MT) levels or AChE activity in digestive glands. Contaminant load and biomarker effects decreased after termination of dumping, indicating the water column effects, though more widespread during dumping, were more transient than possible benthic effects at the dump site. Mussels (*M. galloprovincialis*) transplanted for 4 weeks in two harbors on the Atlantic coast of Spain had induced gill GST activity that correlated with the sediment chemical pollution index (CPI), mussel bioaccumulation index (MBI), benzo(a)pyrene (BaP), and Cu in mussel tissue; induced gill glutathione peroxidase (GPx) activity that correlated with MBI and BaP; and inhibited gill AChE activity that was negatively correlated with sediment CPI and mussel Cu concentration (Vidal-Liñán et al. 2014). Bellas et al.

(2014) combined effects on gill GST, GPx and AChE activities and SFG metrics into an Integrated Biomarker Response (IBR) to examine possible effects of pollutant bioaccumulation in wild *M. galloprovincialis* collected from 40 locations on the N-NW coast of Spain. The highest IBR values were correlated with Σ dichlorodiphenyl trichloroethane (DDT) and Σ hexachlorocyclohexane concentrations in mussel tissue. Lacroix et al. (2014) tested a series of potential reference genes to determine the best to use in a study looking at effects on mRNA biomarkers in caged (1 month) and wild mussels (*Mytilus* spp.) in the Bay of Brest, France. They found it was best to use an index of three of the most stable genes as a reference, which included *28s rRNA*, elongation factor (*ef*) 2 and *ef1a* in gills and α -tubulin (*atub*), ribosomal protein L7 and actin (*act*) in digestive glands. The best biomarkers for differentiating between caged, wild, and reference mussels were cytochrome P450-3-like-2 (*cyp32*), π -*gst* and Cu/Zn-superoxide dismutase (*CuZn-sod*) in gills. Brenner et al. (2014) found that mussels (*M. edulis*) grown from larvae hanging in the water column offshore matured earlier in the year, had little to no parasites, but accumulated more metabolic concentrations of lipofuscin and neutral lipids compared to intertidal mussels. They also developed a lighter shell, which significantly skewed the CI compared to intertidal mussels, leading the authors to conclude that intertidal and offshore mussels should be separated into subgroups when conducting large scale environmental programs. The gastropod *Morula granulata* was collected from 9 intertidal sites off the coast of Goa, India and analyzed for DNA integrity using the time-dependent

partial alkaline unwinding assay, and DNA damage using the comet assay (Sarkar et al. 2014). Both methods showed similar trends, with increased effects correlating with PAH concentration in sediments, and interestingly also nitrate, salinity and phosphate in the collection site water. Seabra Pereira et al. (2014) performed an extensive study with both mussels (*Perna perna*) caged for 3 months and oysters (*Crassostrea rhizophorae*) caged for 28 days in an industrialized estuary on the São Paulo Coast, Brazil. Mussels caged near two sewage outfalls had increased gill EROD, GST, GPx and dibenzylfluorescein dealkylase (DBF) activity, along with increased DNA strand breaks and gill histopathology and decreased LMS, which correlated with degraded macrobenthic community structure in the area. Similarly oysters caged at industrial areas had higher gill EROD, GST, DBF activity, DNA damage, LP and lower LMS which also correlated with degraded macrobenthic community structure. There was also an interesting association of decreased gill LMS with decreased normal development of the larvae from spawned caged mussels.

Field Studies: Crustaceans. Vasanthi et al. (2014) examined heavy metal bioaccumulation and histopathology in mud crabs (*Scylla serrata*) from brackish Pulicat Lake in India. Concentrations of Cu, Pb, Zn, Cd, Mn and Fe were all significantly elevated above those in reference crabs, and concentrations were higher in gills and hepatopancreas over muscle tissue. All three tissue types demonstrated histopathology, including lamellae abnormalities in gills, necrotic tubules and thickened basal lamina in the hepatopancreas, and atrophy and connective

tissue damage in muscles. Jebali et al. (2014) placed caged Mediterranean crabs (*Carcinus maenas*) for up to 60 days in Téboulba Harbor, Tunisia, where PAH and metal concentrations exceeded sediment quality guidelines. Proteomic analysis of hepatopancreas tissue indicated time-dependent effects on proteins including chitinase, carboxypeptidase B, cathepsin L, cryptocyanin, heat shock protein 70 (HSP70), hemocyanin and monocarboxylate transporter. Ramos et al. (2014) collected gooseneck barnacles (*Pollicipes pollicipes*) once a season from three locations in north Portugal. Within season GST activity was for the most part elevated at Lavadores (municipal contamination) and Matosinhos (oil refinery contamination) over the reference in both the cirrus and peduncle. Cirrus LP was decreased at the two contaminated sites during all seasons, but peduncle AChE activity decreased at Lavadores only in winter and spring, and at Matosinhos in spring and summer. There were large increases in cirrus LP and GST activity and to a lesser extent peduncle GST activity in spring and summer across all sites, indicating a possible confounding factor in toxicity data interpretation.

Field Studies: Fish. Pre-spawning female grass goby (*Zosterisessor ophiocephalus*) were collected from five locations in Bizerte lagoon, Tunisia (Barhoumi et al. 2014). Significant positive correlations were found between liver EROD, LP, GST and CAT activities, and muscle tissue ΣPAHs, ΣPCBs and organochlorine pesticides. Muscle AChE activity was inversely correlated with muscle hexachlorobenzene and ΣDDTs, and no correlation was found between metals in muscle and any biomarker.

Dos Santos et al. (2014) examined correlations between liver butyltin concentrations and bile PAHs and liver biomarkers in marine catfish (*Cathorops spixii*) collected from Paranaguá Bay, Brazil. Lower EROD and GST activity, lower DNA damage and higher LP and histopathology were found in fish from the location where tributyltin was highest, versus the sight where bile PAHs and dibutyltins were highest. Lyons et al. (2014) examined effects of location, liver concentrations of coplanar PCBs, sex and age on CYP1A content and EROD activity in California round stingrays (*Urobatis halleri*) off southern California, USA. Males from the mainland had significantly greater EROD activity and CYP1A content than those from the relatively cleaner Santa Catalina Island, but EROD and CYP1A values for females were similar between the two locations, even though the PCB concentrations were much higher in the mainland females. Adult males had lower EROD activity than juveniles, females had a negative correlation between disk width and EROD activity, and both CYP1A and EROD were lower in females than males of all age classes. Podolska et al. (2014) performed an interesting study in the southern Baltic Sea looking at AChE activity in both cod (*Gadus morhua*) and the parasitic acanthocephalans (*Echinorhynchus gadi*) in their gastrointestinal tract. Female cod had lower AChE activity than males, and there was an inverse relationship between the cod and the *E. gadi* AChE activity. There were no site differences in the cod AChE activity, but there were in the *E. gadi* activity, indicating that the parasite may be more sensitive to environmental changes than the host.

Field Studies: Sea Turtles and Mammals.

Guerranti et al. (2014) measured levels of porphyrins, intermediate metabolites in heme biosynthesis, in stranded loggerhead turtles (*Caretta caretta*) in Italy and tried to correlate them with bioaccumulation of several organic compounds. They found a positive correlation between liver perfluorooctane sulfonate (PFOS) concentrations and uroporphyrins. Lehnert et al. (2014) found that hematology markers in rescued abandoned grey seal pups (*Halichoerus grypus*) from the North and Baltic Seas did not vary between admission and release from a rehabilitation facility, but transcription of *hsp70*, aryl hydrocarbon receptor, aryl hydrocarbon receptor nuclear translocator, and to a lesser extent peroxisome proliferator-activated receptor alpha and cytokine interleukin-2 (*IL-2*) in the blood were all higher at admission than release, and lower in pups than adults. Polizzi et al. (2014) sampled Franciscana dolphins (*Pontoporia blainvillei*) incidentally caught by fisherman from an environmentally impacted estuary and a nearby open ocean area off Argentina, and found no correlation between renal or hepatic Cu and Zn and MT concentrations. Seven stranded sperm whales (*Physeter microcephalus*) in southern Italy were sampled by Marsili et al. (2014). Western blot revealed similar CYP1A1 and CYP2B levels in the skin, but CYP1A1 was higher than CYP2B in the liver. Cell cultures were developed from fibroblasts of the whales, and CYP1A1 was induced in the cell cultures by exposure to a mixture of Arochlor 1260, pp'DDT and pp'DDE, and both CYP1A1 and CYP2 were induced by a mixture of BaP and beta-naphthoflavone.

Laboratory Studies: Metals. Green mussels (*Perna viridis*) exposed to Pb for 30 days accumulated the metal several fold above control levels at all concentrations tested (0.008 to 0.109 mg/L) (Hariharan et al. 2014). Several biomarkers were measured in gill and whole body, but only CAT activity (decrease) and LP (increase) in gill and histology of the gill and adductor muscle showed effects at Pb concentrations below the survival LOEC of 0.026 mg/L. Lead caused microtubule cytoskeleton depolymerization and cell death in the seagrass *Cymodocea nodosa* at tissue concentrations of at least 18.33 and 20.24 µg/g dry weight, respectively (Malea et al. 2014). Toxicity appeared to correlate better with rate of Pb uptake rather than to total tissue Pb concentration. Won et al. (2014) identified genes for Cu/Zn-SOD1, Cu/Zn-SOD2, and Mn-SOD in the polychaete *Perinereis nuntia*, then exposed the polychaete to 50 µg/L As, Ni and Pb either singly or in combination for 48 hours. Lead alone and in combination with As enhanced expression of *Cu/Zn-sod1*. The mixture of As and Pb also enhanced *Mn-sod*, more so than Pb alone, As caused a gradual decrease of expression, and the combination of all three metals depressed expression. *Cu/Zn-sod2* was not as sensitive as the other two forms of SOD.

Manila clams (*Ruditapes philippinarum*) exposed to 4 µg/L Cd for 35 days then depurated for 35 days had depressed O:N ratios (a measure of energy source) from day 7 of exposure to 15 days after removal of Cd, and depressed SFG from day 21 of exposure to 7 days after removal (Zhao et al. 2014). Clams exposed to 40 µg/L Cd had depressed O:N ratio and SFG throughout the exposure

and depuration, and SFG was negative for days 21 to 35 of exposure, indicating that the clams were unable to obtain enough energy to fuel their metabolic requirements. Chalkiadaki et al. (2014) exposed the bivalves *Mytilus galloprovincialis*, *Callista chione* and *Venus verrucosa* to 0.5 mg/L Cd for 20 days followed by 10 to 30 days depuration. Cadmium accumulation and MT levels were positively correlated in gill, mantle and body for *M. galloprovincialis*, in mantle and body for *C. chione* and gills for *V. verrucosa*. Cadmium accumulation was positively correlated with LP in *M. galloprovincialis* gills and mantle, and *C. chione* gills, mantle and body, but there was no correlation in *V. verrucosa* tissues. Cadmium accumulation was negatively correlated with AChE in *M. galloprovincialis* gills, mantle and body and *C. chione* mantle and body, but again there was no correlation in *V. verrucosa* tissues. Neither Cd tissue levels nor biomarkers returned to pre-exposure levels after depuration in any species.

Kim et al. (2014) exposed the copepod *Tigriopus japonicus* to up to 100 µg/L of Cu, Zn, Ag, As and Cd for 96 hours to determine effects on reactive oxygen species (as measured by production of dichlorofluorescein from 2',7'-dichlorodihydrofluorescein diacetate) and antioxidant enzyme activities. Cadmium was the least toxic increasing only GST, GPx and SOD activities, and Cu was the most toxic increasing ROS, GSH, GST, glutathione reductase (GR), GPx and SOD, and the toxicity of the other three metals fell in order of Zn>Ag>As. Transcription patterns of 9 HSP genes were also examined, Cu affected the most genes, and *hsp20* and especially *hsp70* were found to be the

most sensitive genes to all five metals. *Heat shock protein 20* expression was also found to be sensitive to Cu as well as Ni in the diatom *Ditylum brightwellii*, but expression was not sensitive to bisphenol A, Arochlor 1016 or endosulfan (Lee, Guo and Ki 2014). Liu, Xiang and Shao (2014) characterized a gene for globular C1q-domain-containing proteins (C1qDC), which are involved in immune response, in *Mytilus coruscus*. They found the gene was temporarily up-regulated upon exposure to pathogenic bacteria, perhaps indicating an acute-phase role in immune response. Exposure for 30 days to 20 µg/L Cu caused up-regulation by day 5, but the levels dropped to background by day 20, and 0.2 mg/L Cd only significantly increased expression on day 20 of the exposure. Morris et al. (2014) characterized the gene for Vacuolar-ATPase subunit A (fVHA-A) and a truncated, potentially non-functional modified RNA transcript (tVHA-A) in the alga *Fucus vesiculosus*, and also developed antisera for the proteins. Algae collected from sites with more metal contamination had higher fVHA-A protein content and *fVHA-A* expression than sites with lower contamination, and also showed the presence of tVHA-A protein and *tVHA-A* expression, where other sites had none. The concentration of the fVHA-A protein increased in algae exposed *in vitro* to 30 µg/L Cu over 11 days. In 300 µg/L Cu fVHA-A further increased, and tVHA-A began to appear after day 6, and gene expression patterns were similar. This may be the first example of an RNA editing event induced by environmental stress.

Laboratory Studies: Nanoparticles. Hu et al. (2014) exposed *Mytilus edulis* *in vivo* and their hemolymph *in vitro* to 400 to 1000 ppb CuO nanoparticles (NP) for 1

hour. In the *in vivo* experiment, the gill tissue contained two proteins that were targets of thiol oxidation (act and triosephosphate isomerase) and three that were targets for carbonylation (α -tub, tropomyosin and Cu/Zn-SOD), indicative of oxidative damage, and the mantle, connective tissue and digestive tubules had increased deposition of pigmented brown cells at all concentrations. All concentrations also increased LMS *in vitro*. The ragworm *Hediste diversicolor* and the clam *Scrobicularia plana* were exposed in a mesocosm for 21 days to 10 µg/L of either soluble Ag (AgNO₃) or Ag NP (Buffet et al. 2014b). Both forms induced oxidative stress, detoxification, apoptosis, genotoxicity and immunomodulation in both species, but GST, SOD, LP and burrowing rate in worms and capsase 3-like protein and burrowing rate in clams were more sensitive to soluble Ag. Nanoparticle Ag caused higher lysozyme activity in worms, and higher laccase-type PO protein levels and DNA damage in clams. Balbi et al. (2014) exposed *M. galloprovincialis* for 96 hours to 100 µg/L of Cd²⁺ and TiO₂ NP both alone and in combination to examine interactive effects. Nanoparticle TiO₂ increased NO production and serum lysozyme activity in hemocytes, but not in combination with Cd. Cadmium increased expression of *mt10* and *mt20* in hemocytes, but not in combination with TiO₂ NP. In the digestive gland additive effects were observed on LMS, and the mixture increased lysozyme and decreased toll-like receptor protein expression when neither toxicant alone had an effect. A similar study was performed by Canesi et al. (2014) examining the interaction of TiO₂ NP and 2,3,7,8-tetrachlorodibenzo-*p*-dioxin (TCDD) in *M.*

galloprovincialis both *in vitro* and *in vivo*. There was an antagonistic effect between the two toxicants on phagocytic activity of hemocytes and efflux activities mediated by ABC transporters in gills both *in vitro* and *in vivo*, and synergistic effects on digestive gland lysosome/cytoplasm volume ratio and estrogen receptor transcription *in vivo*. Also, interestingly, TiO₂ NP also caused significantly higher TCDD to be accumulated in whole soft tissue than when the mussels were exposed to TCDD alone.

Laboratory Studies: Pharmaceuticals and Personal Care Products. Munari et al. (2014) exposed the clam *Venerupis philippinarum* to fluoxetine (a selective serotonin reuptake inhibitor (SSRI) and the active ingredient in the antidepressant Prozac®) for 7 days to examine effects on immune parameters and gill AChE activity. The lower concentrations tested (1 to 5 µg/L) decreased LMS and AChE activity, whereas the higher concentrations (25 to 625 µg/L) increased hemocyte proliferation, and at 25 µg/L only, total hemocyte count. The effects of 0.001 to 1 µg/L fluoxetine and another SSRI antidepressant, sertraline, on amphipod (*Echinogammarus marinus*) light-response behavior and transcription of light-response related genes were examined by Bossus et al. (2014). The amphipods were significantly more active in both light and dark phases, and recovery time from light stimulation was altered for up to 1 day of exposure to each SSRI, but by day 8 no significant differences were observed. On day 8 transcription of rhodopsin, arrestin and neurocan core protein in the head of the amphipod were all downregulated in the lower fluoxetine concentrations, but there were no significant effects in the sertraline exposure.

Ribalta and Solé (2014) exposed liver microsomes of six fish species to up to 100 µM of diclofenac (analgesic), fluoxetine, gemfibrozil (lipid regulator), galaxolide (fragrance), nonylphenol (detergent degradation product) or triclosan (antibacterial agent) to examine effects on cytochrome P450 activity. All compounds depressed EROD activity in most fish species. 7-benzyloxy-4-trifluoromethylcoumarin-*O*-debenzyloxylase activity was depressed in the most species by diclofenac, and was significantly depressed by all compounds except gemfibrozil in two of the species, *Trachyrhynchus scabrus* and *Mora moro*. 7-ethoxycoumarin-*O*-deethylase activity was depressed in *T. scabrus* by all compounds, especially diclofenac, and activity was depressed by diclofenac, galaxolide, nonylphenol and especially fluoxetine in *Alepocephalus rostratus*. Maranhão et al. (2014) exposed the polychaete *Hediste diversicolor* for 14 days to sediments spiked with 0.05 to 500 ng/g carbamazepine (anti-epileptic drug), ibuprofen (analgesic), and propranolol (β-blocker drug) and 0.01 to 100 ng/g fluoxetine and 17α-ethinylestradiol (contraceptive), separately. Carbamazepine increased LP and GST activity, and GPx and AChE activity at lower concentrations only. Ibuprofen increased AChE activity and DNA damage at higher concentrations and increased LP and decreased DBF activity at intermediate concentrations. Propranolol increased EROD activity and decreased DBF and GST activity, and increased LP at lower concentrations. Fluoxetine increased EROD activity, AChE activity and LP and decreased DNA damage. 17α-ethinylestradiol increased EROD and decreased DNA

damage in all but the highest concentration, and increased AChE activity and LP at lower concentrations.

Laboratory Studies: Halocarbons. Green mussels (*Perna viridis*) were exposed for 7 days to 0.1 to 1000 µg/L of either PFOS or perfluorooctanoic acid (PFOA) (Liu, Gin and Chang, 2014). Whole tissue CAT activity and hemolymph DNA damage (comet assay) increased, and relative condition factor decreased, in a dose-dependent manner for both PFOS and PFOA. Filtration rate and gill EROD activity were only decreased at 1000 µg/L PFOS, and hemolymph LMS was only decreased at 100-1000 µg/L PFOA. Liu, Chang, Gin and Viet Tung (2014) exposed green mussels to the same concentrations of PFOS and PFOA, plus perfluorononanoic acid (PFNA) and perfluorodecanoic acid (PFDA) for 7 days to examine DNA damage. All four compounds increased single strand breaks (comet assay) and chromosomal breaks (micronucleus test), and all but PFOA increased apoptosis (DNA diffusion assay), with an overall magnitude of toxicity of PFOA < PFNA < PFDA < PFOS. A 7 day depuration decreased single strand breaks in PFOS and PFNA-exposed mussels, but not chromosomal breaks or apoptosis. Rhee et al. (2014) developed an expressed sequence tags database for the Arctic green sea urchin (*Strongylocentrotus droebachiensis*) and used it to examine transcription effects of a 48 hour exposure to 1 ppb PCB (Aroclor 1254). They found that *gst-a*, *gst-m*, *gst-o*, *gst-p*, *gr*, *Mn-sod*, *Cu/Zn-sod*, *hsp70.5*, and *hsp90* were all upregulated. Fong et al. (2014) used isobaric tags for relative and absolute quantification (iTRAQ) and 2D gel electrophoresis to examine proteomic effects of dietary

exposure to 2,2',4,4'-tetrabromodiphenyl ether (BDE-47) in the gonads of marine medaka (*Oryzias melastigma*). A 21-day exposure to up to 1.30 µg/g daily intake caused differential expression of up to 30 proteins in testes including down regulation of histone variants, which may disrupt spermatogenesis, and HSP90. In ovaries up to 30 proteins were also differentially expressed, including upregulation of apolipoprotein A-1 and four VTGs, perhaps indicating that BDE-47 may act as an estrogen mimic. Hong, Shen, Wang and Shi (2014) exposed marine medaka embryos to 5 to 200 µg/L technical grade hexabromocyclododecane from fertilization to 17 days post fertilization (dpf). Malformation of hatched larvae including yolk sac and pericardial edema, spinal curvature and heart malformation increased in a dose-dependent manner, along with heart rate at 5 and 8 dpf. Oxidative DNA damage (8-oxo-2'-deoxyguanosine) 6 dpf, caspase-3 activity 5 and 8 dpf, caspase-8 and -9 activity 8 dpf, *p53* and *IL-1β* expression 5 and 8 dpf and 1st fry stage, tumor necrosis factor expression 5 dpf and 1st fry stage, and apoptosis in the heart region at 1st fry stage all increased in a dose-dependent manner.

Laboratory Studies: Biocides. Chen et al. (2014) exposed marine medaka to the antifouling compounds butenolide (2.31 µg/L) and 4,5-dichloro-2-n-octyl-4-isothiazolin-3-one (DCOIT, 2.55 µg/L) for 28 days to examine proteomic effects. In male brains butenolide changed expression of 26 proteins primarily involved in DNA and RNA processing and cytoskeletal assembly, whereas DCOIT changed expression of 18 proteins primarily involved in signal transduction and cellular

communication. In female brains 27 and 23 protein expressions were changed by butenolide and DCOIT, respectively, and both affected DNA and RNA processing, metabolic processes and cytoskeletal assembly similarly, with the exception of the proteins collagen alpha-1(I) chain and intermediate filament ON3-like isoform 1, which were upregulated by butenolide and downregulated by DCOIT. Gomes et al. (2014) determined that the primary form of cholinesterase in the polychaete *Capitella teleta* is AChE, and its activity had an *in vitro* 30-minute IC₅₀ of 60.7 nM chlorpyrifos-oxon (active metabolite of insecticide chlorpyrifos), similar to that found in fish and molluscs. Lozano et al. (2014) exposed 3 microalgae (*Phaeodactylum tricoratum*, *Rhodomonas salina* and *Cylindrotheca closterium*) for 24 hours to Cu (5 and 10 µg/L), atrazine (herbicide, 25 and 50 µg/L) and Irgarol (antifoulant, 0.5 and 1.0 µg/L) to examine effects on antioxidant enzymes. In general, SOD and CAT activity increased and GPx and ascorbate peroxidase activity decreased, with *P. tricoratum* appearing to be the most sensitive species, and atrazine having the strongest effect. The diatom *Ceratoneis closterium* was exposed for 96 hours to nominal concentrations of Cu (2.5 µg/L), ammonia (4 mg-N/L), water-accommodated fraction of oil (1 g/L) and the herbicide simazine (300 µg/L) to examine general effects on transcription (Hook et al. 2014). The number of differentially expressed transcripts was 319, 325, 210 and 151 for Cu, ammonia, oil and simazine, respectively. Ammonia, Cu and oil downregulated most genes, whereas simazine caused a downregulation of respiration-associated

transcripts, but increased the abundance of photosynthesis, purine metabolism, and stress response transcripts.

Laboratory Studies: Polycyclic Aromatic Hydrocarbons. Deng et al. (2014) developed a digital gene expression library for the testes of scallops (*Chlamys farreri*) exposed to 0.5 µg/L BaP for 10 days. Two hundred and twenty-three genes were upregulated and 828 were downregulated, including genes involved in aromatic compound catabolism, endocrine disruption, energy synthesis and immune response. European flounder (*Platichthys flesus*) exposed to contaminated sediment for 7 months showed little evidence of bioaccumulation, with only elevated muscle Pb and biliary 1-hydroxypyrene, a metabolite of pyrenes (Williams et al. 2014). There were, however, 259 transcripts and metabolites with altered abundance, which when analyzed had a similar profile to that seen in BaP exposure, and erythrocyte DNA damage (comet assay) was significantly higher than in fish exposed to reference sediment. Clams (*Venerupis philippinarum*) exposed to 2 to 50 µg/L phenanthrene for up to 15 days increased gill and digestive gland SOD activity and glutathione (GSH) content in a time and dose-dependent manner, but EROD and GST activity tended to peak at day 6 and 10 µg/L (Zhang et al. 2014). DNA damage (alkaline unwinding assay) was highest on day 3, and LP on day 6. Xia et al. (2014) exposed clams (*Ruditapes philippinarum*) to the water-soluble fraction of Shengli oil field (China) crude oil at nominal concentrations of 0.1 to 3.2 mg/L petroleum hydrocarbon for up to 15 days. Concentrations of 0.8 to 3.2 mg/L tended to cause an increase in hepatopancreas SOD, CAT, GST and peroxidase activity

during the first two days of exposure, followed by a decrease back to control level (CAT) or below control levels (SOD, GST, peroxidase). Concentrations of 0.1 to 0.2 mg/L caused an increase in CAT and GST activity peaking at days 4 and 8, respectively, with no effect on SOD or peroxidase activity. GPx activity was suppressed at 0.1 to 0.2 mg/L and increased at 1.6 to 3.2 mg/L at most time points, with little effect at 0.4 to 0.8 mg/L. *Tigriopus japonicas* copepods exposed to the water-accommodated fraction of Iranian heavy crude oil for 24 hours had dose-dependent decreases in development and hatching rate and increases in GST, GR and CAT activity, with GSH content peaking at 40% and no effect on GPx activity (Han et al. 2014). Three P450 CYP3 genes were transiently upregulated: *CYP3024A2* peaking at 3 hours, *CYP3024A3* peaking at 6 hours, and *CYP3027C2* peaking at 12 hours.

Laboratory Studies: Radiation. Two bivalve species, *Pahia malbarica* and *Meretrix casta*, were exposed to a single dose of 2 to 10 Gy of γ radiation, the amount expected to be released in a nuclear accident, to examine hemocyte DNA damage via comet assay (Kumar et al. 2014). Damage increased in a dose dependent manner with little difference between species. The damage decreased over time up to 72 hours, perhaps indicating repair, but still remained significantly elevated above control.

Biomarkers of Climate Change Effects. De Souza et al. (2014) exposed Atlantic halibut (*Hippoglossus hippoglossus*) for 14 weeks to pCO₂ of 1000 μ atm, equivalent to a pH drop of 0.4 from control, at both optimal temperature (12°C) and upper tolerance limit (18°C) to simulate the potential levels in 2100 in a fossil fuel-

intensive scenario. Proteomic analysis indicated that at both temperatures CO₂ affected regulation of immune function proteins in the plasma, including complement component C3, fibrinogen β -chain precursor and IgM heavy chain constant region. In the gills proteins involved in energy production (e.g. ATP synthase, enolase- α) and cellular turnover/apoptosis (e.g. annexin 5 and receptor for activated protein kinase C) were affected at both temperatures, and cytoskeleton proteins (tropomyosins) were downregulated at 12°C only. Carregosa et al. (2014) exposed 2 native (*Venerupis corrugata* and *V. decussata*) and one invasive (*V. philippinarum*) clam to both low (0, 7, 14 ppt) and high (35, 42 ppt) salinities for 6 days (control levels were 21, 28 ppt). Mortality was highest for *V. corrugata*, with 100% mortality at 0, 35 and 42 ppt. In general, LP, total GSH content (all species) and GST activity (*V. corrugata*) increased at both low and high salinities, reduced GSH content and reduced/oxidized GSH ratio increased at 14 ppt only (all species), SOD activity was highest at 14 ppt and decreased at higher salinities (all species), CAT activity decreased at higher salinities (all species), and GST activity decreased at both low and high salinities (*V. decussata*). Madeira et al. (2014a) increased the water temperature from 20°C, 1°C/hour, to determine the critical thermal maximum (CTMax), the point at which loss of equilibrium occurs, for the intertidal rock goby (*Gobius paganellus*), and also sampled muscle, liver and gills every 2°C to examine effects on HSP70 and total ubiquitin levels. The CTMax was 33.1°C. Gills contained the highest levels of HSP70 and ubiquitin, and temperature increase only caused a transient increase ubiquitin at 30°C

in liver. Madeira et al. (2014b) performed a similar study with juvenile gilthead seabream (*Sparus aurata*), but included intestine, hepatopancreas, muscle, gills and brain for HSP70 and ubiquitin measurements, and also measured histopathological alterations. The CTMax was 35.5°C. Transient increases in HSP70 were measured in brain (30°C), muscle and gills (34°C), and intestine (CTMax). Transient increases in ubiquitin were measured in muscle (32°C) and gills (34°C). Histological alterations from control (18°C) began to occur at 24°C in the liver and pancreatic acini, 28°C in the gills and intestine, and 34°C in muscle.

Field Methods and Pollution Indicators

Rubal et al. (2014) reported that species-rich assemblages in rocky pools (Portugal) that are subject to relatively minor pollution levels displayed taxa richness and diversity values that were not significantly different from reference pools. However, multivariate analyses revealed changes in the structure and dispersion of these assemblages when the influences of dominant taxa were down-weighted by square root transformations prior to analysis. Based on a comparison of macroalgal community composition at different sites on the Mar del Plata coast (Argentina), Becherucci et al. (2014) concluded that the order-level pattern showed a high similarity and significant correlation to the species-level pattern. Del-Pilar-Ruso et al. (2014) analyzed the responses of benthic macroinvertebrate assemblages along a pollution gradient in the western Mediterranean Sea (Spain) according to three levels of taxonomic identification: phylum, class, or

order; polychaetes at family level; and syllidae (polychaetes) at genus or species level. Multivariate statistical results demonstrated the occurrence of similar distribution patterns in relation to environmental factors for the different taxonomic levels. Based on a review of five different trawl surveys from very different habitats, Brind'Amour et al. (2014) found that identification of benthic megafauna at the genus, family, and morphospecies levels provides acceptable alternatives to species-level identifications for use in community assessments. The use of morphospecies (i.e., taxa easily identified based on obvious morphological characteristics) may be especially useful for comparing community data from disparate studies using different strategies, sampling gear, and taxonomic levels.

A study of rocky shorelines located near sewage discharges and control areas on both the Atlantic and Mediterranean coasts indicated that the responses of intertidal assemblages were location dependent and that these assemblages may not serve as generally reliable indicators of any pollution effects (Cabral-Oliveira et al. 2014). In this study, the responses of macroalgae and limpets to sewage discharges were different between the Mediterranean and Atlantic coasts. Riera and de-la-Ossa-Carretero (2014) concluded that the BOPA (benthic opportunistic polychaetes and amphipods) index was generally unreliable for detecting pollution impacts in a variety of habitats in the Canary Islands. Significant differences between impact and control areas were found only in an area of harbor expansion, while weak or no effects were observed using the index in the following

areas: fish cages, brine and sewage discharges, and thermal discharges. Daief et al. (2014) reported on the first application of the Multivariate AZTI Marine Biotic Index (M-AMBI) to assess the ecological status of urban sandy beaches. Use of M-AMBI enabled the identification and prioritization of urban beaches affected by sewage discharges near Casablanca, Morocco. Comparative assessments of benthic macroinvertebrate assemblages in coastal areas off the Yangtze River Estuary using M-AMBI indicated that the ecological status was seriously degraded, while use of an alternative index (AMBI) indicated only a slight degradation (Liu, Li, Lin, Cai and Wang 2014). M-AMBI was recommended for use in Chinese coastal areas because of the integration of diversity and species richness into the index. A study of benthic assemblages in Victoria Harbor (Hong Kong) demonstrated that benthic macrofauna and free-living nematodes respond differently to a gradient in organic enrichment of the sediments (Xu, Cheung and Shin 2014). As a group, the macrofauna appeared to be more tolerant of the pollutant stress, and the use of both biotic groups was recommended for understanding the overall response of the benthic ecosystem. Niquil et al. (2014) concluded that none of the classical trophic-dynamic concepts tested in a model application (food-chain length, detritivore/herbivore ratio, and trophic efficiency) are reliable as operational metrics to evaluate the impacts on the trophic state of an ecosystem.

Pawlowski et al. (2014) identified gradients of foraminiferal assemblages near two salmon net cage facilities on the west coast of Scotland and tested the applicability of next-generation sequencing (NGS) of DNA

and RNA extracted from sediment samples. The results indicated that NGS metabarcoding of foraminiferal species has the potential to be a valuable tool for assessing impacts to sediment assemblages of forams in the marine environment. A measured series of differentially expressed genes were recommended as potential biomarkers of polycyclic aromatic hydrocarbon (PAH) exposure in the commercially important clam *Venerupis philippinarum* (Liu, Pan, Gong, Tao, Hu and Miao, 2014). The modulation of the expression of these genes from PAH exposure may affect immune defense of the clam and may have a causal relationship with observed mass mortalities of the species. Analyses of stable nitrogen isotopes in the limpet *Patella caerulea* in the northern Adriatic Sea indicated that this information provides a useful indicator of organic matter uptake from anthropogenic sources (Rožič et al. 2014). The $\delta^{15}\text{N}$ in limpets displayed only small differences according to the size of sampled organisms, and variations between tissues for individual organisms were less than 1 percent. Evaluation of $\delta^{15}\text{N}$ for various marine biota of the Glician Coast (Spain) showed that conclusions concerning the contribution of anthropogenic sources were highly dependent on assumptions used to model contributions of alternative nitrogen sources (Bode et al. 2014). Based on these results, the authors concluded that $\delta^{15}\text{N}$ levels alone do not provide a reliable assessment of the influence of anthropogenic nitrogen in coastal waters. Martins et al. (2014) used a variety of fecal sterol measurements and associated molecular marker indices to describe the extent of sewage pollution in Babitonga Bay (Brazil). Comparisons of the

sediment sterol concentrations with sterol index limits in the current literature suggested that such limits may underestimate the magnitude of sewage contamination in the subtropical Bay. Analytical techniques were described for the quantification of five pharmaceutical residues in wastewater effluents, receiving waters, and caged mussels off the Irish coast (McEneff et al. 2014). Three of the five pharmaceuticals were detected in mussel tissue at concentrations of 4–29 ng g⁻¹ dry weight. The results indicated the value of *Mytilus* spp. as bioindicators for this class of substances in the marine environment. A survey of urea levels in the Gulf of Trieste demonstrated that urea is a reliable indicator of the extent of sewage discharges in the coastal zone (Cozzi et al. 2014). Urea was found to be more sensitive and specific than other nutrients, and it is important in biogeochemical nitrogen cycles in the Gulf.

The Catlin Seaview Survey is a new framework for monitoring of coral reefs using underwater imagery (González-Rivero et al. 2014). The computerized framework maps a variety of coral reef characteristics with a spatial resolution of 2 to 6 m². Rengstorf et al. (2014) reviewed the methods for predicting the distribution of benthic biota in the deep sea and evaluated the use of several kinds of information for predicting the occurrence of the deep-water coral *Lophelia pertusa*. The results demonstrated that predictive models incorporating vertical and horizontal demersal flow patterns are substantially better than models using only substrate variables in predicting *Lophelia* distribution. In the Learmonth Bank area of northern British Columbia, Neves et al. (2014) used multibeam sonar (backscatter, bathymetry, and slope) to

characterize bottom substrates. Subsequent analyses and comparisons with ROV surveys demonstrated significant correspondence between substrates and coral/sponge biotopes, indicating that substrate characterizations are useful surrogates in predicting biotope distributions for this area. High-resolution sidescan sonar (410 kHz) and multibeam echosounder data were used to successfully produce high-precision maps of reefs of the tubiculous polychaete *Sabellaria spinulosa* near an offshore windfarm in the North Sea (Pearce et al. 2014). Post-construction maps of the site revealed that the wind farm was having a positive influence on the overall biodiversity of nearby areas. Reshitnyk et al. (2014) compared the applicability of high-resolution satellite imagery and a 200-kHz acoustic system for mapping the distribution of submerged macrophytes in British Columbia (Canada). The satellite imagery was accurate at depths less than 3 m and was able to distinguish various macrophyte types and unvegetated substrate. The acoustic technique was more accurate for the distribution of eelgrass but was unable to distinguish between some algal types. Beeden et al. (2014) described a participatory monitoring program for the Great Barrier Reef (GBR) that provides current information to resource managers on benthic community composition, reef condition and impacts, coral diseases, and presence of debris. The program has been refined over a 10-year period and accumulates observational data from rangers, tourism operators, and the general public. Although specific to the GBR, the protocol could be modified to meet the needs of resource managers at other reef areas.

Based on an assessment in Rijeka Harbor (Croatia), Frančišković-Billinski and Cukrov (2014) concluded that analysis of bulk sediments (<2 mm) is a better approach to assess contamination of marine sediments than analysis of the fine fraction (<63 μm). No significant statistical differences between the sediment fractions were detected for 19 elements using multivariate analyses and comparisons of element concentration ratios. A group of abiotic variables was found to explain about 42 % of the variability in rocky reef community characteristics on the Queensland coast (Australia) (Richmond and Stevens 2014). The correlation with purely physical characteristics was relatively poor (explaining 22 % of variation), but the predictive ability of the statistical model increased dramatically when fishing pressure was included in the combination of explanatory factors. Foster et al. (2014) presented guidelines for designing surveys of benthic habitats using autonomous underwater vehicles (AUVs). In cases where there is no *a priori* information on the distribution of environmental covariates, structured 2-dimensional designs (grid, stratified, or GRTS) are recommended. Kernel density estimation surfaces were demonstrated as a novel and reliable technique for interpretation of trawl data in the Grand Bank/Flemish Cap area of the northwest Atlantic (Kenchington et al. 2014). When used in conjunction with literature data, this approach can be useful for defining vulnerable marine ecosystems, such as assemblages of large sponges, sea pens, and gorgonian corals in the study area.

Effects of Wastewater Discharges

Diez et al. (2014) described a long-term (1984–2012) monitoring program of rocky subtidal macrophytes along the Basque coast (Spain) during a period of sewage treatment upgrades for a large metropolitan area. Although some recovery of macrophyte assemblages was observed, the development of canopy-forming macrophytes that are characteristic of reference areas had not occurred during a period up to 11 years after implementation of secondary sewage treatment. Rocky shore macroinvertebrate assemblages near a sewage discharge on the Portuguese coast had higher secondary production levels than reference areas (Cabral-Oliveira, Dolberth and Pardal (2014). The increase in production resulted from higher abundances of tolerant suspension-feed biota near the discharge. The long-term decline of marine algal taxa near Igidea (Korea) was attributed to a nearby sewage discharge (Shin et al. 2014). Ganesh et al. (2014) characterized a 3- to 6-km gradient of benthic macroinvertebrate assemblages offshore of a sewage discharge in the surf zone of the Bay of Bengal (India). Although the outfall is located in a high-energy environment, the nearby benthic communities were dominated by opportunistic spionid polychaetes. A before-and-after study of a new offshore sewage discharge in the Brač Channel (Adriatic Sea) revealed an increase in phytoplankton biomass in the deep zone after initiation of the discharge, but there was no associated phytoplankton bloom or oxygen depletion (Skejić et al. 2014). In the post-discharge period, there was an increase in phytoplankton species diversity, indicating that moderate nutrient inputs do not necessarily result in adverse effects. Bănaru et al. (2014) described the variability of $\delta^{15}\text{N}$ values according to

zooplankton size and seasons in the Bay of Marseille. The study indicated that contaminated particulates from local sewage discharges are being mixed with marine phytoplankton (mainly pico- and nanoplankton) and are transferred up the zooplankton trophic links.

Berreta et al. (2014) described the levels of various pharmaceutical and personal care products (PPCPs) in sediments in a bay in Brazil that has received domestic and hospital sewage for extended periods. The highest measured concentrations were associated with the fragrances galaxolide (52.5 ng g^{-1}) and tonalide (27.9 ng g^{-1}), with pharmaceuticals being measured at lower concentrations of 0.7 to 14.3 ng g^{-1} . The endocrine-disrupting chemicals 4-nonylphenol (NP) and bisphenol A (BPA) were detected in seawater samples from the Cape D'Aguilar Marine Reserve (Hong Kong) at mean concentrations ranging from 62.5 to 392.5 ng L^{-1} (Xu et al. 2014). Based on a risk quotient approach, the ambient concentrations of NP were predicted to pose a potential hazard to marine organisms. A comprehensive survey of polar organic micropollutants in nearshore seawater samples from a variety of global locations revealed that caffeine was the most frequently detected compound, at concentrations up to 3068 ng L^{-1} (Nödler et al. 2014). The high detection frequency of caffeine in these samples (95.4 percent) indicates the widespread occurrence of untreated sewage discharges in coastal areas.

Effects of Dredging and Extraction

Hwang, Lee, Choi et al. (2014) documented negative effects on fish assemblages of sand extraction

activities in Gyeonggi Bay (Korea). When compared with two non-mining areas, the sand mining area had significantly lower values for total fish abundance, species richness (R1), and species diversity (H'). Based on a review of marine surveys at seven sand extraction sites off Korea, Kim et al. (2014) concluded that turbidity plumes may be transported considerable distances from the extraction site and have adverse effects on the marine environment. Recommendations are presented for monitoring strategies and for minimizing adverse effects of sand extraction operations. Monitoring of a dredged material disposal site in the Saronikos Gulf (Greece) using sediment toxicity tests (Microtox[®]) and various biomarkers in caged mussels indicated contaminant effects primarily associated with high levels of PAH and AH (Tsangaris et al. 2014). Elevated hydrocarbons and associated sublethal effects in caged mussels were not persistent after cessation of disposal activities, indicating that effects on water column organisms are transient. Alternatively, sediment contamination and toxicity persisted after termination of disposal. De Backer et al. (2014) observed similar changes in benthic macroinvertebrate assemblages in the North Sea as the result of three different kinds of human activities: sand extraction, dredged material disposal, and construction of offshore wind facilities. All three activities resulted in a change in sediments toward finer particle sizes, which led to increases in macrobenthic species typical of muddy sands. The overall effect on community structure was an increase in biodiversity for all three sites.

Field Survey Assessments

Sediment concentrations of various organic substances near the port areas of Athens (Greece) indicate that this region is one of the most contaminated sites in the Mediterranean (Kapsimalis et al. 2014). Maximum aromatic hydrocarbon concentrations were dominated by high-molecular-weight PAHs and ranged up to 4,457 $\mu\text{g g}^{-1}$. An analysis of the chemical inputs and impacts of a submarine groundwater discharge (SGD) in the Ionian Sea indicated that the water column at the site had moderate to bad quality, based on a eutrophication index (Pavlidou et al. 2014). However, the SGD has insignificant adverse effects on local benthic macroinvertebrate assemblages. Stewart et al. (2014) reported that the concentrations of a wide variety of emerging contaminants (e.g., flame retardants, pharmaceuticals, and alkylphenols) in sediments near Auckland (New Zealand) were similar to those reported for worldwide studies. Multifactor models were reasonably accurate in predicting uptake of PCBs and polybrominated diphenyl ethers (PBDEs) in benthic macrofauna in the Strait of Georgia (Canada) (Burd et al. 2014). Overall, PBDEs were accumulated more extensively than PCBs, indicating that PBDE transfer to benthos is more dependent on recent organic detritus and associated contaminant levels from ongoing discharges. Slijkerman et al. (2014) documented some evidence of eutrophication based on nitrogen concentrations recorded near the island of Bonaire, which has some of the highest quality coral reef habitats in the Caribbean. A group of liver and gonadal abnormalities were identified in large-toothed flounder (*Pseudorhombus arsius*) and oriental sole (*Synaptura orientalis*) that may provide useful biomarkers for assessment of contaminant

effects in Kuwait Bay (Stentiford et al. 2014). The highest prevalences were measured for foci of cellular alteration (FCA) in the liver and putative intersex (ovotestis) in the gonads of both species.

Hard-bottom assemblages near a discharge of iron process mine tailings in northern Chile were dominated by encrusting species that had replaced the normal macroalgae that inhabit nearby control areas (González et al. 2014). Overall species diversity near the discharge was similar to diversity at control locations, indicating that the observed effects were caused by species replacement, rather than a decrease in overall species richness. Intertidal areas of Baynes Sound (British Columbia, Canada) used for intensive aquaculture of clams were found to have higher abundances of the invasive species *Batillaria* sp. and *Hemigrapsus oregonensis* (Bendell 2014). The aquaculture areas are seeded with juvenile clams and appear to attract the important predatory crab *H. oregonensis*, with unknown effects on overall ecosystem functioning. Mirto et al. (2014) found that nematode species composition in areas near fish farms in the Mediterranean Sea were significantly different from reference areas, but abundance, biomass, and biodiversity did not change near the aquaculture facilities. The nematode genera *Richtersia*, *Desmoscolex*, and *Halalaimus* were very sensitive to discharges from fish farms and almost completely disappeared from the nearby sediments. Benthic macrofaunal assemblages at two Antarctic stations (McMurdo and Casey) had similar relationships with sediment contamination for community composition and biodiversity (Stark et al. 2014). However, very different

responses to contaminated sediments were observed for crustacea (primarily amphipods), which decreased at McMurdo and increased at Casey.

The first scientific survey of fish and invertebrate communities on oil platforms off West Africa documented the presence of diverse platform assemblages that are very different from the soft-bottom shelf communities that are predominant in these waters (Friedlander et al. 2014). The total estimated fish biomass for some individual platforms exceeded 1 ton and was dominated by barracuda (*Sphyraena* spp.), jacks (carangidae), and rainbow runners (*Elagatis bipinnulata*). Following a period in 2010 of warm water and increased storms and runoff, the shallow-water reefs in St. John (U.S. Virgin Islands) showed relatively minor adverse effects in relation to the magnitude of potential stresses (Edmunds and Gray 2014). The detected effects of the unusual year were detected in 2011, in the form of decreases in coral recruitment and higher densities of serpulid polychaetes on settlement tiles placed near the reefs.

Resident male and female bottlenose dolphins in Doubtful Sound (New Zealand) displayed different foraging responses to human disturbance by tour vessels (Symons et al. 2014). When boats were present, male dolphins increased bottom time and conducted fewer dives, while females decreased bottom time and performed more dives. Based on comparisons with an energetic model, it was concluded that both sexes experienced decreased net energy gains because of vessel interactions. Smith et al. (2014) analyzed the impacts of large, seasonal hypoxia events in the Gulf of Mexico on the fishery for brown

shrimp (*Farfantepenaeus aztecus*). Empirical fishery data for the Gulf were generally consistent with the model predictions. A review of environmental considerations for offshore geological storage of CO₂ indicated that leakage from such facilities can have substantial effects on sediments, biogeochemical cycles, and marine biota (Carroll et al. 2014). Recommendations are presented for development of baseline surveys at candidate sites. Lange and Griffiths (2014) presented the results of a large-scale survey of epibenthic megainvertebrates off the west coast of South Africa that may be useful in the future designation of marine protected areas.

Effects of Fishing Activities

Grabowski et al. (2014) reviewed available literature to develop a framework to assess benthic impacts of the various fixed and mobile fishing gear common to New England waters. In general, both susceptibility and recovery scores were highest for hydraulic dredges, intermediate for otter trawls and scallop dredges, and lowest for fixed gear. When compared with a fishery closure area, a demersal trawl fishing area on the continental shelf in the Tyrrhenian Sea (Sicily and Calabria, Italy) displayed a lower density index (DI) and number of species (S) for infaunal communities (Mangano et al. 2014). The fished sites also had an absence of characteristic burrowing decapod species, and the assemblages were dominated by subsurface deposit feeders and opportunistic polychaete taxa. Muntadas et al. (2014) described the changes in benthic community structure and productivity at a trawl site in the northwestern

Mediterranean Sea when compared with a non-fished area. The community changes in the trawled area may be beneficial to adults of the commercially important red mullet, but may cause adverse effects for red mullet recruits. This study demonstrated that permanent fishing closures that allow recovery of benthic communities may be more beneficial to demersal fish species than temporary closures. Van Denderen et al. (2014) found a negative relationship between trawling intensity and species richness of benthic communities in the North Sea. The results of this study emphasized the importance of the spatial scale used to assess trawl impacts because of complex relationships among factors associated with habitat heterogeneity, trawl intensity, benthic community structure, and community productivity. Studies of trophic position ($\delta^{15}\text{N}$) and energy flow in benthic communities in the North Sea demonstrated that either species- or community-level energy flows are good indicators to detect changes in benthic community function after cessation of trawling for 14 months (Dannheim et al. 2014). Because of the cessation of trawling activities in areas developed as future wind farms in the North Sea, there will be an excellent opportunity to monitor the recovery of structural and functional characteristics of benthic communities.

Comparisons of a trawling and dredge fishing area to a nearby fishery exclusion zone in New Zealand indicated that the substrate conditions had been highly modified by the fishing activities (Handley et al. 2014). The baseline demersal habitat was characterized by a shell-gravel surficial layer and the presence of large molluscs, while the fished areas had a homogeneous fine mud bottom

with reductions of species richness and large-bodied taxa. Hydraulic dredging for cockles in Dundalk Bay (Ireland) was found to have no significant effect on sediment characteristics or benthic community structure (Clarke and Tully 2014). The dredging operations resulted in a relatively short-term decrease in densities of the abundant bivalve *Angulus tenuis*. Meseck et al. (2014) reported that a one-time dredging for northern quahog clams (*Mercenaria mercenaria*) in Long Island Sound had only minor effects on a wide variety of sediment chemical characteristics. Short-term differences in organic nitrogen and hydrogen flux were measured following dredging, but these effects disappeared within weeks. Experimental scallop dredging in the Firth of Lorn (Scotland) resulted in significant changes in epifaunal community composition at two of the three study sites (Boulcott et al. 2014). Based on analyses of the effects of dredging over different substrates, it was concluded that pebble and cobble substrates could be especially susceptible to dredging effects on epifauna. A series of photographic surveys of deep-water benthic habitat and megafauna at three potential marine protected areas (MPAs) on the Hebridean Slope (Scotland) revealed the presence of trawl marks in all areas (Hughes 2014). Trawl marks were observed over the complete depth range, but peak abundances were observed in the north area at depths of 1,300 to 1,400 m.

A survey of bait collectors on the sandflats of Langebaan Lagoon (South Africa) revealed that large amounts of sediments were disturbed (6,189 tons), and large areas were trampled ($>500,000\text{ m}^2$) (Nel and Branch 2014). Although the harvesting of sandprawns for bait did

not constitute a threat to the stock, the sediment disturbances resulting from these activities were predicted to have adverse effects on the lagoon ecosystem. Long-term harvesting of intertidal clams was found to have measureable effects on the small-scale distribution patterns of meiofaunal nematodes (Boldina et al. 2014). Clam digging resulted in an attenuation of the normal aggregation patterns of nematodes and a lack of a cyclical pattern in the spatial variogram model. Purroy et al. (2014) developed an interactive map of a large marine protective area in northeast Spain that displayed cumulative impacts of four fishing-gear types: bottom and surface longlines, trammel nets, and gill nets. The greatest potential impacts were associated with areas near the coast and along canyon margins. Surveys of four deep rocky habitats in the Tyrrhenian Sea documented extensive damage to coral colonies from lost fishing gear, consisting primarily of long lines (Bo et al. 2014). Clear injuries to hard-bottom communities were observed in 19 to 62 % of the video frames analyzed from the survey. In a mesocosm experiment, Ingels et al. (2014) assessed the potential indirect effects of trawling disturbances by exposing natural nematode communities in soft sediments to varying densities of large macrofauna that may be reduced by trawling. Removal of large bioturbating macrofauna (e.g., ophiuroids, polychaetes, and bivalves) resulted in indirect effects on nematode community structure. Based on an individual-based model of harbor porpoises in Danish waters, Nabe-Nielsen et al. (2014) estimated that annual by-catch rates from fisheries ≥ 10 % would result in a monotonic population decline that ultimately leads to

extinction of the local population. For the 1990s, the estimated by-catch in the Danish North Sea was approximately 4.1 % of the porpoise population, resulting in a predicted substantial population reduction for the species.

Effects of Marine Debris

Marine debris studies continue to address gaps in understanding about the impacts of all types of debris, especially derelict fishing gear and consumer plastics.

Reviews and Overviews Ivar do Sul and Costa (2014) analyzed 101 publications on microplastic debris and found 60% were written since 2009. Lavender Law and Thompson (2014) provides a brief overview of the state of microplastic research and a perspective on future needs including a better understanding of how often marine organisms encounter microplastic fragments in the natural environment and the interactive risks posed by such encounters. A three-pronged summary of the issues posed by microplastics in the marine environment was published in Koelmans et al. (2014b), with separate and intersecting perspectives from the pharmaceutical industry, academia, and government which all discussed potential solutions for improved education, recycling, and waste management. In agreement with the concept of multi-stakeholder engagement, Lee et al. (2014) found success in addressing the impacts caused by polystyrene buoys in South Korea by implementing a series of participatory workshops to develop policy solutions, such as subsidizing removal of polystyrene buoys after their effective lifecycles to minimize polystyrene fragmentation in marine systems.

Effects on Tourism. Jang et al. (2014), estimated the economic impacts to tourism caused by a marine debris event driven by excessive rainfall on Geoje Island, South Korea. Authors considered the decreased number of visitors and typical expenditures when developing an estimate of lost tourism in the amount of \$29-37 million U.S. dollars (Jang et al. 2014), and suggest countermeasures such as cost sharing by local governments as implementation of the “polluter pays” principle and management of river dams that could be employed to trap debris before it flows downstream. An approach to surveying recreational users of rocky shores in the UK and internationally, in order to determine perceptions of recreational uses and their effects on the environment, spontaneously identified littering as a concern of coastal users as it was a commonly mentioned theme in open-ended questions about recreator insights about factors that affect personal enjoyment of coastal areas (Wyles et al. 2014). This finding suggests marine litter impacts both the ecological environment, as noted in the remainder of this literature review, as well as the social environment through negative impacts to user experiences during recreational trips to coasts (Wyles et al. 2014).

Global Scale Impacts and Directives. A literature review examined cetacean interactions with marine debris on a global scale, determining that the aggregated published literature reports 48 cetacean species (56%) have ingested marine debris (Baulch and Perry 2014). In Europe, scientists supporting the Marine Strategy Framework Directive reviewed potential suitability of marine vertebrates as species to include in their long-term

debris monitoring studies (Galvani et al. 2014) and recommended sea turtles as useful species given known ingestion rates and population status. This recommendation will likely drive future research and monitoring of marine litter in European waters. For instance, sea turtle exposure to marine debris in the Atlantic is analyzed by Gonzalez Carman et al. (2014).

Entanglement in Marine Debris. Given the difficulties in surveying animals that have become entangled in marine debris in the wild, it is not a simple task to determine the prevalence of entanglement for marine species and the effect of such entanglements on marine populations. Generally, the only records come from organisms with severe morbidity or mortality that strand on coasts or are pulled to the surface when derelict fishing nets are retrieved. Two reviews were published in 2014 that assess the state of knowledge on entanglement of cetaceans, pinnipeds, sea turtles, seabirds, and other marine species (NOAA MDP 2014a, WAP 2014). Reviews aggregated records showing that in the United States, 44 seabird species, 9 cetacean species, 11 pinnipeds, 31 invertebrates, and all 6 sea turtle species present in the U.S. have been reported entangled in various types of marine debris. Rates of entanglement appear greater where human populations overlap with convergence zones of high fishing pressure or debris accumulation (NOAA MDP 2014a). In WAP (2014), authors documented country-specific case studies in Australia, the United Kingdom, and the United States and Canada, and used these as a springboard to suggest a global ghost gear initiative to drive solutions in ghost gear hotspots, promote learning from valuable case

studies, and enable global monitoring to better understand the issues and solutions.

Interactions with Derelict Fishing Gear (DFG). Ghost fishing is the indiscriminate take of marine organisms in derelict fishing gear, and may affect target or non-target species of the fishery. A rarely discussed impact of derelict fishing gear is smothering of important marine and coastal habitat. For example, research in the Florida Keys showed submerged marine debris is mainly sourced from the commercial spiny lobster fishery, and was not aggregated in areas of high fishing effort but instead tended to aggregate in sensitive habitat such as coral reefs due to wind and storm action. This over-represented the damaging impacts of derelict traps in coral habitat, with each trap causing damage over a 0.6 m² footprint from movement, breakage, and smothering (Uhrin et al. 2014). Through the use of remotely operated vehicles to survey rocky banks in the Mediterranean between 70-280 m deep, Bo et al. (2014) demonstrate fishing gear impacts such as broken coral colonies and scattered habitat-forming species. This study demonstrated widespread presence of various forms of marine debris, especially long lines, trawl nets, other fishing-associated gear, and litter, and establishes a relationship between presence of debris and decreased coral community health (Bo et al. 2014).

In Anderson and Alford (2014), ghost fishing in Louisiana was examined through citizen science efforts to remove derelict blue crab traps. Of the 3,607 traps collected during removal events, more than 65% were actively ghost fishing and a total of nineteen different species were identified as either captured or killed within traps

(Anderson and Alford 2014). In the Chesapeake Bay, Bilkovic et al. (2014) continued a research program investigating the spatial pattern of derelict blue crab traps in aggregating areas or hotspots. Bilkovic et al. (2014) documented through field research 31,546 marine organisms and 40 species captured in retrieved traps, and used these data to estimate over 900,000 crabs are killed in derelict blue crab traps each year. The majority of non-target species trapped and killed in blue crab traps each year are demersal species (>95%) including Atlantic croaker (*Micropogonias undulates*), black sea bass (*Centropristis striata*), white perch (*Morone americana*), and catfish species (*Ictaluridae sp.*). A total of 46 diamondback terrapins (*Malaclemys terrapin*) were captured and subsequently drowned in derelict crab traps (Bilkovic et al. 2014). In contrast to the Chesapeake Bay, fish traps in the Caribbean are used to catch a variety of species. Field experiments investigated simulated derelict traps and tracked associated fish assemblages, fish behavior, and mortality over six months (Renchen et al. 2014). Authors determined that escape panels did not function as intended, but compared to other locations such as the Chesapeake Bay and Gulf of Mexico, a small number of fish mortalities were reported (2% trapped fish) (Renchen et al. 2014). In Alaskan waters near Womens Bay, Kodiak, 192 red king crabs (*Paralithodes camtschaticus*) were tracked from 1991 to 2008 (Long et al. 2014). Thirteen tagged crabs were killed in derelict fishing gear, while another twenty were captured in ghost fishing traps and released by divers. Using these parameters and life history, population, and fishery harvest data, authors

estimate 16-37% of the population with carapace length greater than 60 mm were killed during the study (Long et al. 2014). This represents a significant portion of the fishery and may indicate a situation in which incidental mortality from ghost fishing should be considered in stock assessment models.

Arthur and Sutton-Grier et al. (2014) published a synthesis of several derelict fishing trap studies across the United States, highlighting some of the similarities among various trap fisheries in terms of the number of derelict traps present, the percentage of derelict traps that ghost fish, and the overall impact of traps on the number of target organisms captured or killed per year. Regional differences were important lines of evidence in showing the dynamics of derelict fishing gear interactions are driven by local factors. Authors present options for a derelict fishing trap management strategy that targets policy options to educate the public and fishermen about ghost fishing, and to develop more effective gear options to minimize loss and ghost fishing given the persistence of traps and the fact that traps did not perform as regulated.

Unintended consequences between marine organisms and fishing gear that has been lost or discarded are difficult to document given the wide expanse of marine waters affected by derelict fishing gear. Documenting interactions between fishing gear and marine species is important, however, when considering how to reduce such occurrences and maintain healthy populations of fishery species, seabirds, turtles, and marine mammals. Yorio et al. (2014) examine a kelp gull (*Larus dominicanus*) colony in which 27 adults were entangled by monofilament fishing

lines used by recreational fishers. Authors suggest that foraging behavior along the coastline may lead to entanglement in lines, which are then brought to the colony and cause severe morbidity and mortality when the line becomes entangled in vegetation (Yorio et al. 2014). A study by Adimey et al. (2014) summarize derelict fishing gear interactions with bottlenose dolphins (*Tursiops truncatus*), Florida manatees (*Trichechus manatus*), and loggerhead (*Caretta caretta*), green (*Chelonia mydas*), leatherback (*Dermochelys coriacea*), hawksbill (*Eretmochelys imbricata*), Kemp's ridley (*Lepidochelys kempii*), and olive ridley (*Lepidochelys olivacea*) sea turtles that stranded in Florida between 1997 and 2009. Overwhelmingly, cases were dominated by hook and line interactions with monofilament fishing line (75.3%), and regression analysis showed interactions with marine debris increased over time for dolphins, manatees, loggerhead sea turtles, and green sea turtles (Adimey et al. 2014). Barreiros and Raykov (2014) report three recent cases of loggerhead sea turtle interactions with fishing gear, in which one turtle swallowed long line and several hooks and was subsequently euthanized due to the damage this caused; one turtle had a limb amputated due to a fragment of tangled nylon long line; and one turtle was enmeshed in a tangle of fishing line such that it caused a limb to atrophy and amputate in the wild. It is suggested that occurrences of fishing gear interactions with marine organisms are more common than scientists are aware.

The literature also contains a number of reports and short communications such as the one by Barreiros and Guerreiro (2014) which provides notes on a bream

(*Pagellus acarne*) entangled in a hollow plastic bottle lid. The fish was caught by hand near the Azores and rehabilitated after removing the lid, which was acting as a collar. Aggregating these reports to understand the frequency and nature of species interactions with marine debris will be important to focusing future solutions-based research efforts.

Debris Ingestion-Invertebrates. Direct ingestion of debris has a suite of potential harmful effects, including dietary dilution, gut blockage, and starvation; laceration and secondary infection; and potentially more subtle effects on hormone function, reproduction, and the immune system that are compounded by the suite of chemicals that may associate with plastic debris as it is exposed to waters and sediments contaminated with substances such as PCBs, PAHs, trace metals, and chemicals integral to the plastic resin itself (NOAA MDP 2014b). In a review of the incidences and health effects of ingesting marine debris, authors at NOAA discuss that marine debris may be ingested directly or consumed indirectly through contaminated prey (NOAA MDP 2014b). The distinction is not novel, but indirect consumption has not been an active line of research to date and is one of many potential ways to consider the “source” of debris in terms of the pathway from point of loss to environmental effect. This section on ingestion of marine debris shows the overwhelming focus of microplastic ingestion studies on invertebrate organisms that form the base of many food webs. These studies lay groundwork for future research evaluating indirect ingestion of debris and the issues that may arise from higher trophic-level organisms eating

contaminated prey. For a detailed review of the species-specific impacts of debris ingestion, see NOAA MDP 2014b.

Studies examining the ingestion of microplastics by marine invertebrates cover zooplankton (Frias et al. 2014; Setälä et al. 2014), amphipods (Chua et al. 2014), isopods (Haemer et al. 2014), sea urchins (Della Torre et al. 2014; Kaposi et al. 2014), bivalves (Van Cauwenberghe and Janssen 2014), and crabs (Watts et al. 2014). Frias et al. (2014) conducted a study sampling Portuguese waters for microplastics associated in surface waters with plankton, in which 61% of samples (n=93) contained microplastic particles. This study is mentioned here as a placeholder for the ongoing monitoring campaigns and studies that are determining environmental concentrations of marine debris and in particular microplastic debris in the oceans. Concentration and density of particles are important variables in risk assessment approaches and fluxes should be carefully considered.

Chua et al. (2014) examine marine amphipod (*Allorchestes compressa*) assimilation of microplastic particles that were isolated from facial soap, and added complexity to the experiment by additionally exposing the amphipods to PBDE congeners. Results were somewhat inconsistent, showing reduced PBDE uptake in the presence of microplastic, but greater (proportional) uptake of the higher brominated congeners (e.g., BDE-153 and BDE-154) (Chua et al., 2014). The marine isopod (*Idotea emarginata*), when offered a food source embedded with fluorescent microplastic particles, consumed the food source without microplastic content (Haemer et al. 2014).

Fecal analysis coupled with fluorescent tracing through the digestive tract confirmed the same concentration of particles were ingested and egested, and accumulation or translocation to tissues did not occur (Haemer et al. 2014).

Sea urchin (*Paracentrotus lividus*) embryotoxicity was considered in Della Torre et al. (2014) through laboratory experiments with nano-sized polystyrene of two surface charges determined by functional groups (carboxylated and amine). The amine-group polystyrene nanoparticles caused developmental effects at 3.85 ug/mL (EC50 at 24 hours post-fertilization) and 2.61 up/mL (EC50 at 48 hours post-fertilization), which may be in part due to the greater dispersion of these particles in the seawater medium. However, in Kaposi et al. 2014, sea urchins (*Tripneustes gratilla*) at the larval stage ingested polyethylene spheres at rates proportional to the treatment concentration, though environmentally relevant concentrations did not have a determinable effect on larval growth. Watts et al. (2014) determined in a laboratory experiment that the shore crab (*Carcinus maenas*) uptake of microplastic particles may occur through inspiration in the gills or through uptake in exposed food. Ingested plastic was retained in tissues for 21 days following inspiration and 14 days following ingestion, indicating the exposure route may influence the physiology involved with trapping and excreting foreign microplastic particles (Watts et al. 2014).

In an effort to focus research on organisms often consumed by humans, Van Cauwenberghe and Janssen (2014) purchase oysters (*Crassostrea gigas*) and blue mussels (*Mytilus edulis*) intended for consumption and

analyzed edible tissues for microplastic concentration. Methods allowed for the full depuration of the organism before tissue analysis, which combined showed an average of 0.36 +/- 0.07 particles per gram (wet weight) in mussels and 0.47 +/- 0.16 particles per gram in oysters. Authors translate this to human ingestion of approximately 1,800-11,000 microplastic particles per year based on a range of mussel and oyster dietary exposures, but caution extrapolation to human health risk assessment (Van Cauwenberghe and Janssen 2014). The implications of marine debris as a source and pathway of chemicals into marine food webs, in terms of human consumption of marine fish and invertebrates, are yet to be determined but will need to be understood within the context of accepted routes of chemical exposure.

Debris Ingestion: Birds. Avian species were perhaps some of the first to be studied for marine debris ingestion, and numerous species have been documented to ingest plastic fragments as well as fishing gear and line. In Acampora et al. (2014), scientists compare juvenile and adult ingestion rates in short-tailed shearwater (*Puffinus tenuirostris*) carcasses obtained through stranding events. No significant relationship was determined between incidence of ingested debris and overall body condition, though perhaps concerning was the finding that juvenile birds had a higher frequency of ingestion and a higher ingestion concentration than adults. Bond et al. (2014) investigated stranded seabird species near Sable Island, Nova Scotia, for ingested debris and noted >72% occurrence in northern fulmars, sooty shearwaters, and great shearwaters. Lavers et al. (2014) sampled feathers and

stomach contents from flesh-footed shearwaters in Australia, and determined a significant relationship between increasing debris body burdens and reduced body condition and contaminant load. Many shearwater chicks obtained debris during parental feeding, and fledglings could attain significant amounts (ranging from 0.13-3.21 grams) of plastic during a single feeding which insinuates this as a major acute route of exposure (Lavers et al. 2014). Within seabird communities, it has been documented that certain species will use debris items within their nest structures. Verlis et al. (2014) document this behavior in brown booby (*Sula leucogaster*) nesting near the Great Barrier Reef, Australia. Authors surveyed 96 nests and found 58.3% contained marine debris, with an average of four pieces per nest structure and a predominance of hard plastic items versus other forms of debris such as fishing line.

Donnelly-Greenan et al. (2014) investigated prey abundance in relation to plastic ingestion in northern fulmars (*Fulmarus glacialis*) near Monterey Bay, California, and noted a significant increase in the plastic categories found in birds stranded from 2003-2007 though no direct relationship with body condition was noted. A broad study of the incidence of plastic accumulation in North Atlantic marine birds determined the highest prevalence of ingested plastic in great shearwaters (71%) and northern fulmars (51%), and in the case of the great shearwater, the highest concentration of plastic fragments in the digestive tract. These broader baseline studies are useful in evaluating trends over time and point to the potential utility of large-scale data aggregation systems and

databases. In a review focused on ingestion of marine debris by seabirds in Canada, Provencher et al. (2014b) found debris ingestion data for only 6 of 91 total seabird species inhabiting Canadian waters, and with additional reports without data for 33 species. Authors propose a focus on characterizing the risk of plastic fragment ingestion and continued monitoring of potential indicator species (Provencher et al. 2014b).

Debris Ingestion: Turtles. Several studies documented the incidence, frequency, and type of plastic debris ingested by sea turtles, including Camedda et al. (2014) which focused on loggerhead sea turtles in Sardinia and determined higher prevalence of user plastics, sheets, and fragments in this population. Hoarau et al. (2014) provided one of the first records of debris ingestion in Indian Ocean loggerhead sea turtles, noting debris in 51% of gut or fecal samples but not finding a relationship between the gut contents of dead individuals and the fecal contents of live turtles. In two peer-reviewed publications and a dissertation published in 2014, Schuyler analyzed global patterns of anthropogenic debris ingestion by sea turtles (Schuyler et al. 2014a, 2014b; Schuyler 2014). This work is expansive and builds on previous reports of debris ingestion to analyze geographic patterns and the similarity of certain debris items to the natural prey of sea turtles. Authors determine a significant increase in the probability that green and leatherback turtles will ingest debris over time, and oceanic species are at the greatest risk of lethal and sub-lethal effects caused by ingested debris (Schuyler et al. 2014a).

Debris Ingestion: Mammals. In two papers published in 2014, Madeira Di Benedetto and Arruda Ramos (2014) and Madeira Di Benedetto and Rodrigues Awabdi (2014) provide information about debris ingestion in large marine vertebrate species in coastal southeastern Brazil. They noted higher concentrations of debris in benthic environments and suggest benthic feeders may be at higher risk for ingesting debris (Madeira Di Benedetto and Awabdi (2014). Similarly, differences in feeding habits were noted to drive variances in debris ingestion between coastal dolphin populations of *Pontoporia blannvillei* and *Sotalia guianensis*, with the ingestion rate of the benthic feeder *P. blannvillei* was estimated at 15.7% of individuals examined, while only 1.3% of *S. guianensis* individuals had debris items within stomach contents. Kaladharan et al. (2014) describe the necropsy of a Longman's beaked whale (*Indopacetus pacificus*) that stranded near Sutrapada, India and upon examination died after the ingestion of four thick plastic shopping bags (190 g) that blocked the intestinal tract. Authors link this to the locally high incidences of anthropogenic litter. Continuing work on marine vertebrates in the Mediterranean, Fossi et al. (2014) measure higher concentration of phthalates in the muscle of basking sharks (*Cetorhinus maximus*) than in blubber of the Mediterranean fin whale (*Balaenoptera physalus*) and discuss possible connections with leaching of these chemicals from fragments of plastic debris.

Alien and Invasive Species Transport. Marine debris has been documented as a vector for the transport of various organisms that attach to debris as a hard substrate. There is much unknown about the frequency of organisms

rafting on marine debris and the incidences of aquatic invasions by non-indigenous species. Building on the current knowledge base, rafting organisms are documented in Calder et al. (2014). Scientists note the presence of fourteen species of hydroids attached to debris that originated in Japan after the Tohoku tsunami in 2011 and stranded on coastlines in the northwestern United States. Three species of hydroids were noted as cryptogenic – it is unknown if they are native or exotic to the Pacific coast of North America (Calder et al. 2014). The authors conclude by underscoring the potential importance of large debris items, such as the docks that floated from Japan to the Pacific coast of North America, in providing sufficient substrate to host communities of organisms in global dispersal. Also in the North Pacific, Goldstein et al. (2014) demonstrate a positive relationship between the size of a debris “raft” and the diversity of taxa associated and supported. Potential causes for this relationship include stochastic settlement of biological taxa randomly on all debris items present within the system, as well as increased migration onto debris items that outpaces the rate of extinction. Larger debris objects may have an advantage in greater stability at the water's surface, in addition to presenting a greater surface area on which biological settlement may occur (Goldstein et al. 2014).

Oberbeckmann et al. (2014) exposed polyethylene terephthalate bottles to conditions in the North Sea for six weeks, and determined diverse microbial communities within the resulting biofilms that varied by season and specific location. Harrison et al. (2014) investigated the colonization of low-density polyethylene in

benthic habitats through the use of mesocosm experiments. Their study showed significant increases in bacterial concentration after seven days and shifts in bacterial community composition with sediment type that did not persist over the experiment, given the dominance of *Arcobacter sp.* and *Colwellia sp.* after fourteen days (Harrison et al. 2014). This study's focus on benthic colonization stands in contrast to most of the literature, which has focused on floating pelagic plastic. Another example of such a study highlighting debris as a new type of pelagic habitat is Reisser et al. (2014), which characterizes biological diversity on millimeter-sized floating plastic debris from coastal and oceanic Australian waters. Importantly, scientists investigated the physical appearance of the debris particles through scanning electron microscopy and observed surface textures including pits and grooves similar to the shape of organisms such as diatoms and coccolithophores that colonized the debris (Reisser et al., 2014). One potential conclusion is that some organisms colonizing the plastic are able to degrade it.

In a study to document and monitor potential habitat changes caused by marine debris, Taylor et al. (2014) evaluated the deep-sea communities that associate with a lost shipping container abandoned to the seafloor, noting the faunal assemblages associated with the container and benthos within 10 meters of the container were significantly different than the surrounding area, possibly resulting from the slow successional rate for crinoids, sponges, and soft corals at that depth. Interestingly, though the container is a disturbance to the seabed which alters

local flow, it also increases habitat heterogeneity and hard substrate on which megafauna may associate.

Chemical Uptake. Scientists investigating marine debris and its role in the transfer of chemicals to the environment and marine organisms are increasingly focusing on smaller particle sizes that may provide greater surface area and greater likelihood of ingestion to a wide range of organisms. In Rossi and Monticelli (2014), the direct ingestion of plastic nano-particles is considered in the context of understanding the interaction of these particles with lipid membranes. Such studies are important within the medical field, and Rossi and Monticelli (2014) focus research on simulating molecular interactions between medical applications (e.g., dendrimers and linear charged polymers) and potential polymers of debris such as polystyrene. Characterizing such interactions will improve understanding of how chemicals may leach from plastic debris particles.

Bakir et al. published two papers in 2014 to further investigate desorption kinetics of persistent organic pollutants (POPs) under laboratory conditions that simulate the natural physiology of marine organisms, as well as the sorptive behaviors of POPs to synthetic polymers and how these behaviors are affected by varied salinity regimes (Bakir et al. 2014a, 2014b). Authors concluded that salinity (tested at 0, 8.8, 17.5, 26.3, and 35 parts per thousand) did not have an effect on the distribution coefficients governing sorption of phenanthrene (Phe) and DDT to the synthetic polymers polyvinyl chloride (PVC) and polyethylene (PE) (Bakir et al. 2014b). Salinity did not affect time to reach equilibrium, desorption rates, or capacity for accumulation

of Phe onto the polymers; however, there was a slight decrease in sorption capacity for DDT onto the polymers that was influenced by increasing salinity. Bakir et al. (2014b) determined that sorptive dynamics are governed more by the combination of contaminant and polymer type than by salinity, and developed a model that estimates between 5 and 15 days for contaminants sorbed to plastics in estuaries to leach from the polymer such that the polymer attains lower contaminant concentrations once it reaches marine waters. A second study noted enhanced desorption of POPs to synthetic polymers under simulated gut conditions than compared to sorption of contaminants in water alone (Bakir et al. 2014a). This study has implications for risk exposure and assessment models that address the impacts of plastic particle ingestion to marine organisms. Simulated gut conditions for warm-blooded organisms provided the most favorable environment for desorption kinetics (Bakir et al. 2014a). Addressing similar knowledge gaps, Lee, Shin and Kwon (2014) used laboratory experiments to measure partition coefficients between microplastic particles and seawater for 8 polycyclic aromatic hydrocarbons (PAHs), 4 hexachlorocyclohexanes (HCHs), and 2 chlorinated benzenes (CBs), using three polymers that represent a large percentage of debris found in the marine environment (PE, PP, PS). Partition coefficients were derived using a third phase (in this case, polydimethylsiloxane) to speed the process, ranged from 2.04-7.87 for polyethylene and a similar range for other polymers, and showed general agreement with log Kow (octanol-water partition coefficient) values.

Polychaete worms (*Arenicola marina*) were exposed to microplastics coated with nonylphenol and phenylrene and the added chemicals Triclosan and PBDE (Browne et al. 2014). Nonylphenol was taken up by the worms, reducing the ability of the coelomocytes to remove pathogenic bacteria. Uptake of Triclosan affected the worm's ability to burrow in the sand and death. The results indicate that micoplastics can affect the physiology of benthic organisms.

Utilizing field collections, Kwon et al. (2014) analyzed the regional distribution of styrene constituents generated from polystyrene (PS; foamed plastic) degradation in the North Pacific, and found breakdown products were more likely to exist along the North American west coast than in Alaska or Hawaii though drivers of this trend are unknown. Llorca et al. (2014) detected perfluoroalkyl substances associated with a limited number of plastic pellets and sediment samples collected along coastlines in Greece. While these studies do not explicitly address the effects of the plastics and/or the associated contaminants, they assist the scientific community in more fully understanding the inherent risks posed by plastic particles as transport mechanisms for chemical contaminants, including metals, plasticizers and other chemicals integrated into the polymer, and contaminants sorbed from surrounding waters or sediments.

Sorption and concentration of metal contaminants onto synthetic polymers has not been well documented. A paper by Holmes et al. (2014) is one of several studies published in 2014 that address this knowledge gap. One primary consideration when assessing trace metals

associated with a plastic matrix is the methodological techniques used within laboratories that consistently rely on the use of plastic equipment and materials when analyzing metals in environmental samples. Method development would seem to be in early but promising stages. Holmes et al. (2014) determined greater sorption to beached polyethylene pellets than virgin pellets for all metals assessed (cadmium, cobalt, chromium, copper, nickel, and lead), and reported various shifts in adsorption based on salinity regime and pH though adsorption is less likely to occur onto synthetic polymers than to sediments. Rochman, Hentschel and The (2014) assessed dynamics of metal accumulation onto five synthetic polymers (polyethylene terephthalate (PET), high-density polyethylene (HDPE), polyvinyl chloride, low-density polyethylene (LDPE), and polypropylene (PP) during a year-long exposure experiment in San Diego Bay. Trace metal sorption did not significantly differ among the five polymers, though all metal concentrations increased during the twelve-month study which implies persistent debris fragments accumulate trace metals over time. In a new study by Graca et al. (2014), researchers analyzed virgin and beached foamed plastics for total inorganic mercury content and found an order of magnitude higher mercury content in foam debris (5.20 ng/g) over virgin foam pieces (0.23 n/g). The presence of biofilms and the interaction of the particles with solar radiation lead to seasonal peaks in Hg concentration during summer months (Graca et al. 2014). Authors noted occasional foam debris particles exceeded bottom sediment and soil standards by measuring approximately 3,000 ng/g mercury, and suggest that with

age these particles could be enhanced in their ability to transport mercury in the environment (Graca et al. 2014).

Researchers continue to use a combination of modeling, laboratory, and field conditions to develop a better understanding of the factors affecting sorption kinetics, and the connection between sorption and desorption upon particle ingestion. A modeling approach assessed the leaching of plastic additives to marine organisms by applying a biodynamic model to calculate potential for transfer of nonylphenol (NP) and bisphenol A (BPA) to intestinal tissues of the lugworm (*Arenicola marina*) and North Sea cod (*Gadus morhua*) (Koelmans et al. 2014a). Public concern over the leaching of such contaminants to seafood emphasizes the importance of the study's major finding that plastic ingestion is a negligible source of plastic additives to cod when compared with other pathways, though plastic particle ingestion could be a substantial pathway for introducing these chemicals to the lugworm.

Nilsen et al. (2014) propose to characterize ingested plastic particles by so-called "indicator chemicals" sorbed to the particle, in order to better document particles and fragments that are not able to be characterized as a particular polymer or resin type. According to this analysis, approximately 68.4% of unidentifiable plastic fragments ingested by Laysan albatross from Kure Atoll are polypropylene, while 20.5% are polyvinyl chloride (Nilsen et al. 2014). Authors suggest that infrequently recycled plastic resins are the more prevalent in this study and potentially more persistent in the environment (Nilsen et al. 2014). Resin type may influence and govern risks posed by

debris ingestion, especially as risk relates to the leaching of chemicals within the digestive tract, and thus this approach to identifying indicators of resin type may prove useful in future risk calculations.

Rochman, Kurobe, Flores and Teh (2014) built on previous work to investigate the potential for endocrine disruption in Japanese medaka (*Oryzias latipes*), a common organism used in toxicity testing, derived from exposure to plastic marine debris. Dietary exposures reveal altered gene expression in male and female fish exposed to polyethylene pellets that had spent three months in San Diego Bay, including down-regulation of choriogenin in males and vitellogenin in females, which the authors suggest warrants further investigation. Another study tested the hypothesis that chemical concentrations of plastic particles would be greater in areas predicted to be debris “hotspots” by a previously published predictive oceanographic model (Rochman et al. 2014). The study involved field collections to examine accumulation of POPs in myctophid fish inhabiting pelagic waters in the South Atlantic Ocean, as well as to document debris concentration in this region. Contaminant concentrations in the fish varied without respect to debris density except for several higher-brominated congeners of polybrominated diphenyl ethers (PBDE) measured in fish tissue (Rochman et al. 2014c). Results highlight the difficulty in determining the relative contribution of various sources of persistent organic pollutants and trace metals to marine organisms; it is challenging to determine relative contributions of chemicals sourced from marine debris versus other

exposure pathways, as tissue residues reflect integration of inputs over long time frames and large spatial scales.

Oil Spills: The Deepwater Horizon

Major oil spills, the Deepwater Horizon (DWH), Helbei Spirit, Prestige, and Exxon Valdez, continued to generate a research long after the events. This section explores papers stemming from the 2010 DWH incident.

Reviews. Pennings et al. (2014) conducted a literature review of effects of oil exposure to terrestrial arthropods and provided research topics for future studies. Field observations conducted at sites impacted by Deepwater Horizon showed an initial decline in terrestrial arthropods and a fast recovery of healthy populations. The authors suggest further investigations into the relationship of arthropods and their associative habitats with various floras in conjunction with their resilience to quantity of oiling.

Özhan et al. (2014) conducted a literature review on the effects of crude oil to phytoplankton species. The paper cites phytoplankton EC50's range from 1.03- >50mg/L when exposed to crude oil and 1.01-1031mg/L for crude oil compounds. The authors also provide citations regarding phytoplankton community diversity shift toward higher concentration of petroleum degrading microbes as a response to exposure to oil from the Deep Water Horizon event.

Exposure Concentrations. Two years after the Deepwater Horizon oil spill, Bianchi et al. (2014) identified dissolved organic carbon (DOC) concentrations ($284 \pm 64 \mu\text{M}$; $n = 3$) and oil-derived chromophoric dissolved

organic matter (CDOM) signatures at certain stations in the deep waters of the Gulf of Mexico that remain higher than typical deep water values ((ca.40–50 μM).

Gray et al. (2014) used liquid chromatography/tandem mass spectrometry with isotope-dilution quantification to measure concentrations of dioctyl sulfosuccinate (DOSS) at the surface and in the water column in the vicinity of the Macondo (a.k.a. Deepwater Horizon) well head from May 27-June 4, 2010 during the Deepwater Horizon oil spill. The highest concentrations of DOSS were recorded within 3 km of the wellhead, but only three sites had concentrations which exceeded the U.S. Environmental Protection Agency's aquatic life benchmark of 40 $\mu\text{g L}^{-1}$.

Smith, Flemings and Fulton (2014) studied the transport of hydrocarbons from two deep water vents in the Gulf of Mexico incorporating salinity and temperature gradients to include multi-phase hydrocarbon distribution. The author's one dimensional advection-diffusion model predicted hydrocarbon flux to exceed over 100 times previous estimates and attributes natural seeping hydrocarbons supporting bacteria communities that contributed to the high biodegradation rates observed during the Deepwater Horizon event.

Microbial Community Response. MacDonald et al. (2014) introduced the special issue of Environmental Research Letters which contains papers related to the Deepwater Horizon oil spill and covers topics ranging from the transience and persistence of discharged hydrocarbons to microbial responses to the large input of oil derived carbon.

Crespo-Medina et al. (2014) tracked methanotrophic activity in the deep sea during and after the Deepwater Horizon oil. They found that the methanotrophic population peaked in May/early June 2010, but then crashed in late June, even though deep water methane concentrations remained elevated (tens of μM , with a maximum of 180 μM) above background. Since methane remained abundant, the authors suggest that physiological or environmental factors such as kinetic selection of the methanotroph population, trophic interactions or mortality (for example, viral lysis, selective grazing pressure), nutrient or trace metal limitation are among the likely causes.

Kostka et al. (2014) discussed the impact that cutting edge molecular and biogeochemical techniques (including high throughput sequencing, isotope tracers, and _omic approaches) are having in advancing understanding of the biogeochemical processes and metabolic pathways that control hydrocarbon biodegradation in marine systems. They then outlined the major themes of the papers contained within the Research Topic, including diversity of and impacts to the microbial community in the Gulf of Mexico from the Deepwater Horizon oil spill, bridging laboratory studies of biodegradation to the field, and environmental controls of oil biodegradation in marine sediments.

Engel and Gupta (2014) found that the microbial community of sandy beaches in Grand Isle, Louisiana and Dauphin Island, Alabama that were impacted by oil from the Deepwater Horizon oil spill shifted from bacterial communities that are associated with fecal contamination

(pre-oiling) to open ocean associated communities known to consume hydrocarbons (post-oiling). The authors suggest that sand washing and tilling caused the regime shift.

Lamendella et al. (2014) reported a significant increase in microbial cell density and a shift in the microbial community toward a hydrocarbon degrading consortium on a beach in Louisiana after it was heavily contaminated with oil from the Deepwater Horizon oil spill. 16SrNA gene sequencing and Metatranscriptome profiling established the presence of both known and suspected hydrocarbon degrading microorganisms including *Marinobacter*, *Roseobacter*, and *Pseudomonas* species, among others.

Scott et al. (2014) compared sediments impacted by the Deepwater Horizon event, sediments exposed to natural seeps found in Santa Barbara, and reference sediments collected in Gulf of Mexico to assess the impact of polyaromatic hydrocarbon exposure to the nitrification processes in the upper layer of marine surface sediments. The authors specifically measured effects on genes involved with nitrogen metabolism and nitrogen cycling and reported significant differences between the three sediments in relative turnover rates for nitrogen metabolites.

Thomas et al. (2014) conducted a 3 month microcosm study using field collected Apalachicola Bay, Florida oysters, water and sediments, as well as, including oil-degrading bacteria colonies inoculated with oil from the Deepwater Horizon spill site. The authors report changes in the diversity of bacteria in the microcosms with respect to

the biodegradation process of oil, however, they report there was not a reference “no oil” microcosm to compare the results.

Joye et al. (2014) reviewed recent literature outlining the effects of the Deepwater Horizon oil spill on microbial community composition and activity in various Gulf of Mexico ecosystems. The microbial community shifted rapidly in response to the oil and gas input, a wide variety of microorganisms, including alkane, PAH, and methane degraders and nitrifying microorganisms responded, marine oil snow formed, and dispersants appear to have had an adverse effect on the microbial community and microbe-based food web, among other effects.

Effects on Flora. Judy et al. (2014) reported that common reed (*Phragmites australis*) was resilient to oiling of its shoots (0-100% cover) by both weathered and emulsified Macondo oil from the Deepwater Horizon spill, but that oil applied to the soil (0-16 l m⁻²) and repeated oiling of shoots resulted in reductions in above-and belowground plant growth.

Effects on Phytoplankton. Özhan, Parsons and Bargu (2014) reviewed the factors that influence the toxicity of oil to phytoplankton which includes among many other things, diversity of phytoplankton physiologies, geographic location, oceanographic and meteorological conditions, nutrient concentrations, seasonal variations, oil type, and oil dosage (impacts generally seen between 1 and 100 mg/l), and outlined direct and indirect (e.g. formation of oil films that limit gas exchange and reduce light penetration) toxic effects. The authors then summarized recent studies of effects in the phytoplankton community

from the Deepwater Horizon oil spill including phytoplankton blooms and the formation of marine snow, as well as laboratory studies that examined the toxicity of Macondo oil and dispersants.

Effects on Mollusks. Using stable carbon (C14) analysis of the tissues of barnacles (*Balanus sp.*) and marsh mussels (*Geukensia demissa*), Fry and Anderson (2014) concluded that little to no oil from the Deepwater Horizon oil spill was incorporated into estuarine food webs. The authors also measured respiration rates in estuarine waters that were within the range of previously reported values for uncontaminated waters, leading them to conclude that there was very little microbial community enhancement in estuarine waters as a result of the oil spill.

Synder et al. (2014) monitored PAH concentrations in field collected Coquina clams (*Donax spp*) and shoreline sediments for 21 months. The authors report PAH concentrations in clams ranging from 5-1000ppb and sand 5-500ppb. They suggest clams are a useful tool to monitor PAH pollution in high energy surf zones.

Effects on Crustaceans. Yednock and Neigel (2014) field collect blue crabs, *Callinectes sapidus* from Louisiana and Texas coastline during May to July 2010 and 2011 to conduct genetic sequencing analysis across 9 sample locations. Significant differences were reported for adult individuals and the authors speculated results were due to exposure to oil and Corexit 9500A mixtures during the DWH event coupled with the shrimp fishery closures that would predictably negatively impact adult blue distribution.

Effects on Pelagic Fish. Brette et al. (2014) demonstrated that crude oil from the Deepwater Horizon oil spill affects the regulation of cellular excitability in cardiomyocytes isolated from juvenile bluefin (*Thunnus orientalis*) and yellowfin (*T. albacares*) tuna. While effects were seen at ΣPAH concentrations as low as 4 µg/liter, the cardiotoxicity of the oil correlated to the concentrations of three-ringed PAHs in the oil samples.

Incardona et al. (2014) found adverse effects on cardiac function in Bluefin tuna, Yellowfin tuna, and Amberjack (*Seriola dumerili*) fish embryos at PAH exposure levels (1-15µg/L total PAH) consistent with those documented in the Gulf of Mexico during the Deepwater Horizon oil spill.

Mager et al. (2014) reported that juvenile mahi-mahi (*Coryphaena hippurus*) displayed impaired swimming performance when exposed to the water accommodated fraction of oil from the Deepwater Horizon oil spill. Embryos/larvae that were exposed for 48-hour at 1.2±0.6 µg L-1 ΣPAHs and then raised to juveniles displayed reduced swimming efficiency as a latent effect, while exposed juveniles displayed such effects only at the highest concentration tested (30±7 µg L-1ΣPAHs).

Effects on Nearshore Fish. Crowe et al. (2014) exposed Gulf killifish (*Fundulus grandis*) to water-accommodated fractions (WAFs) of crude oil (7.0 _0.10 mg/L C6-C28 at time 0) from the Deepwater Horizon oil spill for 12, 24, and 48-hour intervals. They observed a significant increase (66%) in the expression of the CYP1A in the fish exposed for 24-hours, and significant increases

in antioxidant capacity of nonenzymatic antioxidants in exposed fish at each time point.

Murawski et al. (2014) investigated reports of offshore fishes exhibiting skin lesions following the Deepwater Horizon well blowout and documented a significant decline in overall lesion frequency (1.9% to 0.9% (all species sampled) and bile PAH levels (Red Snapper (*Lutjanus campechanus*) between 2011 and 2012 sampling events at the Northern Gulf of Mexico sampling sites. There was a strong correlation ($r^2=0.82$; $p<0.001$) between oil collected at the Deepwater Horizon wellhead and the composition of PAH parent compounds and alkylated homologs in the Red Snapper liver samples.

Seafood Safety. Fitzgerald and Gohlke (2014) sampled seven species of Gulf of Mexico reef fishes for PAHs, Dioctyl Sodium sulfosuccinate (a component of the dispersants Corexit 9500A and 9527A), and several metals in the aftermath of the Deepwater Horizon spill as part of a fishing industry led study to ensure the safety of their catch. Of 92 samples, none exceeded federal safety standards for benzo(a)-pyrene-equivalents or contained DOSS, and metals were absent or within expected values.

Effects on Deep Sea Coral. Fisher et al. (2014a) documented adverse impacts attributable to the Deepwater Horizon oil spill to two additional deep sea coral communities located six kilometers and 22 kilometers away from the well-head. They also surveyed numerous coral communities around the northern Gulf of Mexico, and found no acute impacts to corals at depths between 400 and 850 m and greater than 30 km from the well-head. Fisher et al. (2014b) reviewed the effects of oil from the

Deepwater Horizon oil spill on corals and soft-sediment ecosystems in the Northern Gulf of Mexico below 400 m. Damage to coral colonies in three separate coral communities was easily observed by the presence of dead/dying branches that had been colonized by hydroids. Examination of sediment cores from areas with 30km of the Macondo (Deepwater Horizon) well site contained significantly higher levels of saturated hydrocarbons and PAHs (459 – 47,600 $\mu\text{g}/\text{kg}$) and less macrofaunal and meiofaunal diversity than previously recorded in the Gulf of Mexico (0-1030 $\mu\text{g}/\text{kg}$), although distribution was patchy. Coral associated sediment communities displayed similar decreases in diversity and shifts in community structure.

Fredericq et al. (2014) documented a dramatic die off of rich algal assemblages (rhodoliths) after the Deepwater Horizon oil spill at two previously documented sites in the northern Gulf of Mexico. They speculate that the corals may have suffered “chemical bleaching” from the possible induction of oil into porous bedrock, stimulating anaerobic sulfate reducers and an associated production of hydrogen sulfide, or from a potential increase in their release of dimethylsulfoniopropionate production. Bare rubble brought back from the sites was naturally recolonized in the laboratory setting, while the same rubble at the sampling sites had not recovered as of October 2013.

Effects on Birds. Bergeon Burns et al. (2014) reviewed the effects of oiling on terrestrial vertebrates which may include reduced water repelling properties, thermoregulation difficulties, immunosuppression, habitat alteration and prey impacts with subsequent effects on the

organisms, reduced reproductive success, and compromised ability to handle additional stressors, among others. The authors then outlined their approach in attempting to determine potential effects of oil from the Deepwater Horizon oil spill on seaside sparrows (*Ammodramus maritimus*) and marsh rice rats (*Oryzomys palustris*) living in heavily oiled salt marsh in Louisiana, while noting the difficulty of distinguishing the effects of the oil from many other complex and potentially confounding factors present in the Gulf of Mexico (e.g. hurricanes, subsidence, erosion, development pressure, salinization, other variability introduced by the presence of oil (e.g. pathogen increases) etc.). The authors' referenced preliminary data that shows seaside sparrow nests are significantly less likely to fledge in oiled areas than are those in non-oiled areas, but have not yet analyzed the data in the context of other confounding factors.

Franci et al. (2014) compared body mass, corticosterone, and prolactin levels in two populations of Northern gannets (*Morus bassanus*): one which overwinters in the Gulf of Mexico and potentially was exposed to oil from the Deepwater Horizon oil spill and one that overwinters on the Atlantic coast. The authors found no difference in the populations, and no variation in the hormone levels over the early to late incubation period.

Paruk et al. (2014) field collected 38 common loons (*Gavia immer*) off the coast of Louisiana near Barataria Bay during Jan-March of 2011 and 2012 and extracted blood samples along with biological measurements prior to release. In 2011, 3 of the 17 birds captured tested positive for Anthracene with concentrations

ranging from 1.7-2.3 ng/g. In 2012, 13 of the 21 birds captured tested positive for Anthracene concentrations ranging from 3.4-8.0ng/g and 1 tested positive for Fluoranthene at a concentration of 1.6ng/g.

Using an exposure probability model incorporating oil slick size, bird density, and proportionate mortality for their calculations, Haney et al. (2014) estimated that between 36,000 and 670,000 birds died in the offshore (+40km) of the Gulf of Mexico (GOM) as a result of exposure to oil from the Deepwater Horizon oil spill.

Using both a carcass sampling model and an exposure probability model, Haney et al. (2014b) calculated that approximately 700,000 birds died in the coastal GOM as a result of the Deepwater Horizon oil spill, but the numbers may be as low as 320,000 and as high as 1,900,000 within the 95% uncertainty bounds.

Walter et al. (2014) banded over 1,000 Brown Pelicans (*Pelecanus occidentalis*) between 2007 and 2009 and observed their migration and age distribution at study sites between 2008 and 2011. The authors observed 54 pelicans in 2011 after the DWH event and specifically 12 of the 182 banded pelicans that underwent oiling, rehabilitation, and release. The authors could not attribute that age structure variation was a cause of exposure to oil from the DWH event.

Effects on Marine Mammals. Schwacke et al. (2014) conducted an observational study on bottlenose dolphins (*Tursiops truncatus*) using Barataria Bay, LA, a heavily oiled location impacted by the Deepwater Horizon event, and Sarasota, FL as a reference site. Dolphins were

captured, evaluated, and released. The authors reported dolphins in Barataria Bay showed common symptoms due to oil exposure, such as lung disease, hypoadrenocorticism, and pulmonary consolidation as well as mortality rates higher than previously recorded. A critique and reply followed publication.

Rice et al. (2014) collected acoustic recordings of the Bryde's whale (*Balaenoptera edeni*) in the Gulf of Mexico between 2010-2012. The authors suggest the existence of the small group of whales in the Gulf of Mexico and should be considered before dispersant use.

Using a Bayesian modeling approach that incorporated a conditional occupancy estimator and prior information on detection probability, Martin et al. (2014) estimated that at the time of their aerial surveys, fewer than 2.4% of the manatee (*Trichechus manatus latirostris*) habitat potentially impacted by oil from the Deepwater Horizon oil spill may have contained manatees, and probably contained 107 or fewer manatees.

Use of –Omics Methods. Bik (2014) reviewed how the use of metagenomics is revolutionizing our understanding of ecosystem functioning. _Omics studies in the aftermath of the Deepwater Horizon oil spill were cited to illustrate how the use of –omics data is increasing our understanding of biological responses to the influx of oil in the Gulf of Mexico.

Other Oil and Fuel Spills and Research

Spain (Prestige Spill). Barros et al. (2014) documented long term reproductive impairment (+10 years) in the European Shag (*Phalacrocorax aristotelis*) as

a result of oil exposure from the 2002 Prestige oil spill. Reproductive success was reduced by 45% in oiled colonies vs. unoiled colonies after the spill as compared to the years prior to the spill. Castège et al. (2014) examined the effects of the Prestige oil spill on the benthic community of the rocky shore of Guéthary (south of the Bay of Biscay, France). Three years post spill, taxonomic richness had returned to pre-spill levels, but the community structure took almost five years to reestablish. Junoy et al. (2014) found that although macrofaunal assemblages of O Rostro Beach (Galicia, NW Spain), the beach most heavily oiled by the Prestige oil spill, displayed negative effects (reduced abundance and species richness) in the first six months after the spill, the community had recovered to pre-spill conditions by 2004.

South Korea (Hebei Spirit oil spill). Two years after the cleanup of the 2007 Hebei Spirit oil spill, Kim, Hong, Kim and Yang (2014) found that PAH levels in bottom sediments both inside and outside of Mohang Harbor, Korea ranged from 24-366 µg/kg, posing minimal ecological risk. Lee, Kim, Jeong et al. (2014) documented chamber breakage in 71.6% of the benthic foraminifera, *Ammonia beccarii*, collected from the substrata sediment of Sogunri tidal flat, Taean Peninsula, Korea. The authors attributed the breakage to decalcification caused by low pH (6.98) of the sediments contaminated with oil-mineral aggregates deposited from the Hebei Spirit oil spill. After three years post-spill monitoring Seo et al (2014) reported that subtidal sediment PAH concentrations returned to below background. There were moderate changes in the benthic infauna community, and no mass mortalities of amphipods

except at two stations. The opportunistic colonial polychaete, *Prionospio paradisea* dominated one site 10 months after the spill.

India (Ship collision). Sukumaran et al. (2014) conducted a 15 month environmental monitoring study to assess the impact of 800 tons of fuel oil spilled during a ship collision between MSC Chitra and MV Khalija 3 in Mumbai Harbour in 2010. The authors collected sediment cores for macrobenthic organism analysis, sediment characteristic and hydrocarbon concentration. Macrobenthic organisms showed less diversity after the oil spill than the next year and the authors reported a range of hydrocarbon concentration in sediments to be 5.6-311.3 µg/g.

Alaska (Exxon Valdez). Fukuyama et al. (2014) compared intertidal infaunal communities in unoiled, oiled and untreated or lightly treated (oiled), and oiled sites treated with high-pressure hot water washing (treated) over the period of 1990-2000 to evaluate impacts and recovery after the Exxon 1989 Valdez oil spill. They found that the infaunal communities in the oiled and treated sites had largely recovered by 2000, but treated sites were lacking large sized little neck clams (*Leukoma (Protothaca) staminea*) found in untreated and oiled sites, correlating to fewer fine grained sediments in the treated sites related to cleanup activities. Harwell and Gentile (2014) used new estimates of subsurface oil residue encounter rates for sea otters (*Enhydra lutris*) at northern Knight Island to update their individual-based model risk assessment. The authors concluded that the risk to sea otters from these encounters

is small, and that the population has fully recovered from the Exxon Valdez oil spill.

San Francisco (Dubai Star). Hwang, Stanton, McBride and Anderson (2014) found that body burdens of PAHs and levels of lysosomal membrane destabilization in mussels were higher from animals collected from moderately oiled shorelines than those collected from the same shoreline prior to oiling from the 2009 Dubai Star release (average 1077±519 ng/g (dry weight) vs up to 87,554 ng/g post spill). Body burdens of PAHs had returned to pre-oiling levels three months post-spill.

Arctic. Dunton et al. (2014) sampled the benthos of the northeastern Chukchi Sea to establish baseline conditions for chemical and biological characteristics. Metals, PAHs, and aliphatic hydrocarbons were at natural background levels at nearly all sites sampled, and there is a rich benthic community of organisms with a complex food web.

Jörundsdóttir et al. (2014) documented concentrations of non-alkylated PAHs and inorganic trace elements (arsenic, cadmium, mercury, lead) in blue mussels (*Mytilus edulis*) and PAH metabolites in cod (*Gadus morhua*) bile in the Nordic arctic and sub-Arctic coastlines to establish background conditions. In general, PAH concentrations and inorganic trace elements were low in mussels (Σ16PAH ranged between 28 and 480 ng/g d.w.) and 1-OH-pyrene was only quantifiable (between 44 and 140 ng/ml bile) in samples collected from the Norwegian coast. Harvey et al. (2014) documented very low concentrations of PAHs (<1600 ng g⁻¹ dry wt) in the top 0-1 cm of sediments of the Chukchi Sea, and found that

common Arctic cod (*Boreogadus saida*) CYP1A1, GST, and SOD enzyme levels were comparable to baseline levels in other pristine systems. They also opportunistically sampled Northern whelk (*Neptunea heros*), finding lower concentrations of PAHs (4.5–10.7 ng g⁻¹ wet wt), but larger concentrations of aliphatic n-alkanes (C19-C33) (0.655–5.20 µg g⁻¹ wet wt) in larger animals as compared to smaller animals.

Payne et al. (2014) conducted 14-day multi-species community toxicity tests and 7-day single species toxicity tests using field collected Antarctic zooplankton *Oncaea curvata*, *Oithona similis*, and *Stephos longipes* exposed to Special Antarctic Blend diesel water accommodated fractions. Three replicates of zooplankton community exposed to SAB diesel WAF treatments reported LC50 values ranging from 186-1091 µg TPH/L. The authors reported LC50 values of 158, 176, 188 µg TPH/L for *O. curvata*, *O. similis* and *S. longipes*.

General. Reviews. Chang et al. (2014) reviewed the consequences of oil spills from tankers and developed an overview framework to be used as a basis for planning and discussion by focusing on the following areas: the oil spill itself, disaster management, the physical marine environment, marine biology, human health, economy, and policy. The authors then used the expected increase in tanker traffic in Vancouver, Canada as a case study to illustrate potential impacts of a spill and their associated complexities, including the need to account unique conditions associated with each spill and locality.

Farrington (2014) outlined the fate and effects of oil spills in the marine environment and discussed some

cleanup and mitigation measures, including the use of dispersants, Environmental Sensitivity Indices, and oil spill models.

Kirby et al. (2014) outlined the following eight principles of effective post-oil spill monitoring: scientific guidance, skills and knowledge, equipment, funding, responsibility and management, integration and coordination, support and buy-in, and practice. The authors then provided a case study of the United Kingdom cross-government program known as Premium (Pollution Response in Emergencies: Marine Impact Assessment and Monitoring) which illustrates the implementation of the principles.

Klemas and Blažauskas provided a brief overview of information and actions needed to minimize the damage from oil spills and facilitate cleanup efforts, including the use of environmental sensitivity maps. The authors then introduced the Baltica Special Issue publication that stemmed from the common Lithuanian-Russian project, “Development of solutions for effective oil spill management in the South-Eastern Baltic.”

Ventikos and Sotiropoulos (2014) provide a literature review on the costs of responding and cleaning oil spill incidents and statistical models from a data set including 107 spills ultimately illustrating size and location directly influence costs. The authors also categorize accident types: sinking, grounding, collision, other and quantity spilled 1-10,000 tons to provide cost estimates of impact. The authors do not take in to account type of oil spilled and location.

Suzdalev et al. (2014) summarized the increase of shipping and potential threat of oil spills in the Baltic Sea. The authors provide examples of past spills in the area of concern and highlight improvements of spill response planning that have occurred in the area including area contingency plan updates, updating sensitive shoreline mapping tools, and use of new oil fate modelling tools.

Peters and Siuda (2014) reviewed the history of tar balls and tar mats observed in the Sargasso Sea from 1960-present and asserted detectable levels of tar balls have declined over time due to increased regulations and environmental monitoring.

Laboratory and field investigations. Han et al. (2014) found that exposure to the water accommodated fraction of Iranian crude oil at 20%, 40%, 60%, and 80% of stock solution 25 g crude oil/L caused significant delays in development, molting, and hatching rates in the copepod *Tigriopus japonicus*, as well as increased the activities of antioxidant enzymes. There was no mortality, even at 100% WAF exposure. The authors also identified three TJ-CYP genes as potential biomarkers of oil exposure.

Harms et al. (2015) found significant increases in key hematological values in loggerhead turtle (*Caretta caretta*) hatchlings exposed to Gulf Coast mixed sweet crude oil (0.833 mL/L) and/or dispersant (Corexit 9500A, 0.083 mL/L) as compared to non-exposed controls, with the greatest changes in the animals exposed to combined oil/dispersant mixtures.

Jurelevicius et al. (2014) dosed marine (Massambaba Beach) and hypersaline waters (Vermelha Lagoon) from the Massambaba Environmental Protection

Area, Rio de Janeiro, Brazil with crude oil (1 % v/v), naphthalene as a representative PAH (1 % w/v), or heptadecane as a representative aliphatic hydrocarbon (1 % v/v) to observe the effects on the Archaeal communities. No Archaeal communities were detected in the marine waters contaminated with hydrocarbons, but they were present in the hypersaline hydrocarbon contaminated water.

Lindgren et al. (2014) found that the addition of Swedish Mk-1 diesel at nominal PAH concentrations of 1300 µg ΣPAH/kg dw sediment in three different sediment types (muddy, sandy, and organic) resulted in significant negative impacts to the microbial community as measured by potential nitrification and denitrification, although the negative effect was less in muddy sediments. PAH availability was greater in the sandy and organic sediments than in the muddy sediments. No effect was observed on the meiofaunal community in any of the sediments.

Kang et al. (2014) reported significant declines in the density of meiofauna in all plots treated with 0.125 L, 0.25 L, 0.625 L, or 1.25L of crude oil, and increases in nematode species diversity in all treated plots except the 1.25L treatment as compared to the control plots with no treatment. However, the community recovered to pre-spill character and structure within one month of the spill.

Leite et al. (2014) found no significant differences in nematode total density, diversity, and community structure as compared between marine diesel oil treated (2500 ml application) and control plots located in unvegetated tidal flats of the Paranaguá Estuarine Complex (Southern Brazil). Total aliphatics in experimental plots ranged from 2.31 to 30.8 µg g⁻¹ dry

sediment in the treated plots as compared to 1.42 to 2.77 $\mu\text{g g}^{-1}$ in the control plots.

Morandin and O'Hara (2014) found that feathers exposed to sardine oil sheens (ranging from 0.04 to 3 μm in thickness) displayed significant feather microstructure disruption and significant weight gain from oil and water uptake at all sheen thicknesses tested. From this data and in conjunction with interviews with wildlife rehabilitation experienced with oiled bird rehabilitation, the authors concluded that edible oils are at least as harmful to seabirds as petroleum oil.

Based on data collected from both live and dead oiled birds, Henkel et al. (2014) estimate that >1,000 birds are oiled oil on an annual basis on the California coast by natural petroleum seeps and/or other chronic sources of oil (i.e. shipwrecks Luckenbach and Palo Alto). Oiling peaks in the late winter (Jan.-Apr.).

Stauffert et al. (2014) conducted a 270 day microcosm study evaluating the effect of oil exposure to field collected sediments containing Archaea. The authors quantified Archaea population diversity by using terminal restriction fragment length polymorphism (T-RFLP) fingerprints and conducted 16S rRNA gene and 16S cDNA sequencing. The authors reported oiled treatments had a significant effect on Archaea community structure.

Sugahara et al. (2014) exposed fertilized embryos and larval Takifugu niphobles to pyrene and phenanthrene to assess morphological and behavioral abnormalities due to exposure. Concentrations of pyrene ranged from 10-100ppb and phenanthrene test concentration was 200ppb. The authors reported effect on larval swimming behavior at

10ppb of pyrene and assert the effect was due to morphological stunting in the brain. No effects were observed for phenanthrene exposures.

Witt et al. (2014) conducted Microtox toxicity tests using *Vibrio fischeri* exposed to 128 field collected sediment samples to generate EC50 values and establish data that assists in developing a matrix to identify effects of pollutants in sediments. The authors used EC50 percentages ($\leq 1\%$ Very Toxic; $1\% < \text{EC}50 < 2\%$ -Toxic; $\text{EC}50 \geq 2\%$ - Non-Toxic) to predict and model toxicity of sediments of the Gdansk basin area and reported the toxicity decreased in a trend from the northeast to southwest with median EC50% ranging from 0.63-9.33% for the 5 areas.

Modeling Effect of Oil Exposure. Redman et al. (2014) investigated the bioavailability of hydrocarbons to rainbow trout (*Oncorhynchus mykiss*) using two methods, a PETROTOX model and solid-phase microextraction (SPME) approach. The goal of the research was to assess the appropriateness of using SPME methods to model toxicity and associated pathways as well as identify toxic compounds included in complex heavy fuel oil. The authors suggests the SPME method is a valid simple model to predict toxicity as the values compared appropriately with the values generated by the PETROTOX model.

Vaiene et al. (2014) compared the results of using two bioaccumulation models to predict PAH concentrations in 5 marine organisms: fish species *Gadus morhua*, *Paralichthys olivaceus*, and *Scophthalmus maximus*, the mollusk *Mytilus edulis*, and a crustacean *Pandalus borealis*. The authors investigated the difference between

bioaccumulation models that did and did not incorporate ingestion of oil droplets and reported that no difference between models was observed for all species except the mollusk, *Mytilus edulis*.

Risk Assessment. Weller et al. (2014) modeled the effects of multiple stressors on the endangered African penguin *Spheniscus demersus* including anthropogenic influences on food availability, oil spills, and predator/prey dynamics. Data was derived by multiple published studies and expert opinion. The authors reported the results of the model for managing the African penguin indicated preventing oil exposure from incidents coupled with increasing food resources would positively impact population sustainability.

Rogowska et al. (2014) conducted toxicity assays using field collected sediments coupled with a self organizing map algorithm to complete a risk assessment of potential pollution from a shipwreck in Gdańsk Bay. Toxicity tests included *Vibrio fischeri* microtox test, *Sinapis alba* phytotoxicity test, and *Heterocypris incongruens* chronic toxicity test. Sediments were analyzed for PAH and metal concentrations. All available data were utilized in the SOM algorithm which generated a series of similarities between endpoints and pollution concentrations.

Dispersants and Other Oil Spill Treating Agents

Use of chemical dispersants to combat the 2010 Deepwater Horizon (DWH, Gulf of Mexico, USA) oil spill stimulated increased research of the effects of dispersants and dispersed oil on marine and aquatic organisms. Studies

in 2014 included effects of dispersant products and dispersed oils on viruses, bacteria, phytoplankton, zooplankton, and adult or early life stages of mollusks, crustacean, and fishes., and also studies on other oil spill chemical treating agents. Readers should be aware that (1) inconsistent reporting of toxicity in terms of nominal concentrations, measured concentrations and/or mass vs % volume dilutions creates confusion and (2) there remains debate over whether dispersants and oil are synergistic or not.

Viruses. Pham et al. (2014) exposed the aquatic viruses: viral hemorrhagic septicemia virus (VHSV), infectious pancreatic necrosis virus (IPNV), chum salmon reovirus (CSV), and frog virus 3 (FV3) to Corexit 9500 using cell lines propagated from fathead minnow and CHSE-214. The authors reported exposures resulted in no effect for IPNV, reduced infectability at concentrations of .0001-10% v/v in VHSV and FV3, and increased infectability cells at concentrations of 1%v/v and higher in CSV.

Phytoplankton. Diatoms *Isochrysis galbana* and *Chaetoceros sp.* to weathered and non-weathered MC-252 crude oil, and Corexit 9500A to conduct acute and growth inhibition toxicity studies (Garr et al. 2014). No statistically significant effects observed for oil only toxicity tests; however, 96hr IC50 values reported for Chemically Enhance Water Accommodated Fractions (CEWAFs) of non-weathered MC252 crude oil were observed at 149 mg/L and 200 mg/L for *I. galbana* and *Chaetoceros sp* and CEWAFS of weathered oil 48hr test observed at 174 mg/L and 91 mg/L for the same respective species. Özhan and

Bargu (2014) conducted a 10 day microcosm study using field collected phytoplankton as the test organisms and Louisiana crude oil (SLC), Corexit 9500A, and Corexit 9500A treated SLC treatments. The authors reported Corexit 9500A only treatment showed significant effect of growth inhibition at a concentration of 63ppm. The authors had additional test treatments where additive nutrients were applied and observed less of a toxic effect from LSC exposure to specifically sensitive species. Phytotoxicity assays were also conducted to assess the effect of exposure to South Louisiana sweet crude oil (SLC) and Corexit 9500A-dispersed SLC on growth of marine diatoms and dinoflagellates (Özhan et al 2014). The authors reported TPH EC50 values for the oil only tests as 2498, 1835, 1751, 1138, 1025 ppb for *Ditylum brightwellii*, *Chaetoceros socialis*, *Pyrocystis lunula*, *Scrippsiella trochoidea*, *Heterocapsa triquetra*, respectively. The Corexit 9500A dispersed SLC treatments EC50 is reported as <100ppm for both diatoms and dinoflagellates species.

Planktonic Animals and Life Stages. The active ingredients in Corexit 9500, propylene glycol and 2-butoxyethanol, were evaluated for toxic effects using 48hr acute toxicity methods and the marine foraminifer *Amphistegina gibbosa* (Ross and Hallock 2014). The authors reported findings as Acute Concentration (AC50) due to organism recovery after 48hr exposure and LC50 after 48hour exposure with a 24hr observational period. The 48hr AC50 and 72hr LC50 for propylene glycol was 3% and 6%v/v, respectively and 0.2% and 1%v/v for 2-butoxyethanol. Investigations on the toxicity of MC-252

and Corexit 9500A were conducted by Cohen et al (2014) using copepods *Labidocera aestiva* in 24hr and 48hr acute toxicity tests. The authors reported 48 hr LC50's in the form of total petroleum hydrocarbons of crude oil WAF treatments as 40.5 µL/L 24hr, 37.5 µL/L 48hr and CEWAFs as 190.4 µL/L 24hr and 74.3 µL/L 48hr. Corexit only treatments were reported as 7.8mg/L and 4.5mg/L for 24hr and 48hr treatments, respectively.

Almeda and associates evaluated the effect of Louisiana sweet crude (LSC), Corexit 9500A, and Corexit 9500A treated LSC mixtures exposed to lower trophic level organism. Almeda et al (2014a) reported 48hr EC50s for crude oil, dispersant, and dispersant/oil mixtures for each of the following species: *Stombidium sp* (1.73, 0.08, 1.04 µL/L), *Spirostombidium sp* (0.99, 0.04, 0.85 µL/L), *Eutintinnus pectinis* (1.07, 0.03, 0.15 µL/L), *Favella ehrenbergii* (4.87, 0.2, 2.29 µL/L), *Gyrodinium spirale* (16.42, 0.76, 13.40 µL/L), and *Protoberidinium divergens* (13.73,0.28,5.69 µL/L). Additional 48hr tests with the same toxicants using calanoid copepods, *Arcartia tonsa*, *Temora turbinata*, and *Parvocalanus crassirostris* and investigated ingestion of crude oil droplets on sublethal endpoints (Almeda et al 2014b). At test concentrations of 1 µL/L SLC, 0.5 µL/L Corexit 9500A, 1 µL/L LSC/Corexit, the authors reported % reduction of observed endpoints were most toxic for dispersant/oil mixtures ranging from 45-54% egg production rates, 28-41% fecal pellet production rates, and 11-31% egg hatching. Using the same test concentrations as above, Almeda et al (2014c) conducted 72hr toxicity tests with test organism barnacle nauplii (*Amphibalanus improvises*) and tornaria larvae

(*Schizocardium sp.*). The authors reported no calculable effect concentrations for *Amphibalanus improvises* and *Schizocardium sp.* as having a higher sensitivity with no reported mortalities at test concentrations. Growth inhibition concentrations (GC50) were calculated for *Schizocardium sp.* as 2.5 µL/L LSC only, 0.03 µL/L dispersant only, 0.88 µL/L dispersant/oil mixture.

Multi-species Toxicity. Median effect concentrations of three dispersants: Corexit 9500A, Dasic NS, and Gamlen OD400, and five surface washing agents: Hela saneringsvæske, Bios, Bioversal, Corext 9580, and Absorrep K212 on four pelagic species: *Skeletonema costatum*, *Acartia tonsa*, *Calanus finmarchicus*, *Calanus glacialis*, and one benthic species, *Corophium volutator* was evaluated (Hansen et al 2014). The 72 hr EC50 values for each species were reported as 9.3 - 2270mg/L for *S. costatum*; 48hr LC50 6.5 - 2790mg/L for *A. tonsa*, 96hr LC50 11.5 - >20435mg/L for *C. finmarchicus*, 144h LC50 9- >20435mg/L for *C. glacialis*, and 10-d LC50 140- >15347mg/L for *C. volutator*. Due to the variability of sensitivities to each product and species interaction, the authors ranked the species 1 thru 5 and concluded *A. tonsa* as the most sensitive and *C. volutator* as the least sensitive species.

Koyama et al (2014) exposed diatoms, *Chaetoceras gracilis* and *Skeletonema costatum*, amphipod *Hyale barbicornis*, and red sea bream fish embryos *Pagrus major* to physically dispersed (PDWAFs) and chemically dispersed (CEWAFs) water accommodated fractions and using heavy C oil and D1128 dispersant. The EC50 reported for PDWAF/CEWAF exposures for each species

include 0.39/035 mg/L *C. gracilis*, 0.1/0.34 mg/L *S. costatum*, 0.09/0.06 mg/L *H. barbicornis*, and 0.11/0.018 mg/L *P. major*. Toxicity results were utilized in a simulation model for Tokyo Bay, Japan to determine areas of greatest risk to higher oil concentrations due to environmental factors of wind and currents, as well as, chemical dispersion. Rial et al (2014) tested 2 marine bacteria, *Phaeobacter sp.* (Ph) and *Pseudomonas sp.* (Ps), one terrestrial bacteria, *Leuconostoc mesenteroides*(Lm), and embryo/larval stage sea urchin *Paracentrotus lividus* to four spill response agents Cytosol, Finasol OSR 51, Agma OSD 569 and OD4000. The authors reported the 2 marine bacteria species showed no effect to all exposure levels ranging from 0-2000 µL/L and *Leuconostoc m.* growth inhibition EC50's of 754 µL/L Finasol OSR 51 and 129 µL/L OD4000. The sea urchin embryo tests resulted in EC50 values of 34 µL/L Agma OSD 569, 26.3 µL/L CytoSol, 2.2 µL/L OD4000, and 1.2 µL/L Finasol OSR 51.

Echinoderms. Development toxicity tests using the sea urchin *Paracentrotus lividus* (fertilization) exposed to toxicant mixtures of Maya crude oil and four spill response agents (Cytosol, Agma OSD 569, OD4000, Finasol OSR 51) were conducted to determine median effect concentrations (Rial, Vazquez and Murado 2014). The authors reported 48 hr EC50 dispersant only concentrations 15.1 mL/L Cytosol, 9.8 mL/L Agma OSD 569, 2.6 mL/L OD4000, and 1.8 mL/L Finasol OSR.

Crustaceans. Larval and juvenile stage blue crab, *Callinectes sapidus*, were used to conduct 96hr acute toxicity tests using a range of Corexit 9500 concentrations (Lively and Mckenzie 2014). The larval tests were exposed

to a range of 0.1-100 ppm and juvenile tests were conducted at a range of 1-1000 ppm. The authors reported no LC50 value for juvenile stage blue crab due to no significant mortality at the highest concentration and reported a 48hr LC50 value of 40.9 mg/L for larval stage tests due to no changes in mortality after 48hrs. Rodd et al. (2014) conducted 24hr acute toxicity tests using larval stage *Artemia franciscana* exposed to surface engineered carbon nanoparticles (CB) and powered activated carbon (PAC). The authors reported an LC50 value of 370 mg/L for CB exposures and 750 mg/L for PAC exposures.

Anderson, Kuhl and Anderson (2014) conducted 96hr static toxicity tests using field collected mud crabs, *Rhithropranopeus harrisi* and Louisiana sweet crude (LSC) only and Corexit 9500 treated oil test solutions. The authors did not report an LD50 for the LSC only treatment due to lack of mortality. The 96hr LD50 for the Corexit 9500A/LSC treatment was 25 g/L.

Using copepods *Labidocera aestiva*, Cohen et al. (2014) conducted acute toxicity tests with MC-252 crude oil, Corexit 9500A, and Corexit 9500A treated crude oil treatments. The authors reported 48 hr LC50's in the form of total petroleum hydrocarbons of crude oil WAF treatments as 40.5 µL/L 24hr, 37.5 µL/L 48hr and Corexit 9500A treated crude oil as 190.4 µL/L 24hr and 74.3 µL/L 48hr. Corexit only treatments were reported as 7.8 mg/L and 4.5 mg/L for 24hr and 48hr treatments, respectively. Oil-only and oil/dispersant mixture 48-hr toxicity assays were conducted with the shrimp *Litopenaeus vannamei* exposed to Persian Gulf-Khark crude oil (CO), Iranian Naftroob crude oil dispersant (IND), Radiagreen OSD

dispersant(RD), and mixtures (Delshad et al. 2014). The authors reported LC50 values of 1741 mg/L for CO, 17 mg/L for IND, 43 mg/L for RD, 631 mg/L for CO-IND, and 357 mg/L CO-RD. Acute toxicity test using standard test organism *Americamysis (=Mysidopsis)bahia* exposed to 6 crude oils and No. 2 oil physically dispersed water accommodated fractions (PDWAFs) and chemically dispersed water accommodated fractions (CEWAFs) using Corexit 9500 and 9527 at a ratio of 1:10 was evaluated by Letinski et al. (2014). The authors reported Lethal Loading values (LL50) for PDWAFs ranging from 3-533 mg/L with the lighter crude oils resulting in higher toxicity values. The results of Corexit 9500 and 9527 were combined to report LL50s ranging from 7-64 mg/L and the authors identified lower effect concentrations were due to increase exposure to hydrocarbons and not exposure to dispersant

Molluscs. Laramore et al. (2014) conducted acute and sublethal toxicity studies using the eastern oyster *Crassostrea virginica* to assess exposure effects of Macondo Canyon (MC) 252 oil alone and combined with Corexit 9500A dispersant and reported effect levels based on the following endpoints: fertilization, development, survival, and motility. The authors report negative effects of all endpoints when exposed to concentrations of total polycyclic aromatic hydrocarbon (TPAH) concentrations ranging 1.3-5 mg/L and 0.02-0.2 mg/L which were at or above measured field concentrations during the 2010 DWH event.

Fish. In order to determine presence of synergistic toxicity effects for dispersant/oil mixtures, Adams et al (2014) compared toxicity values by conducting

two separate investigations. The first test exposed Atlantic herring (*Clupea arengus*) embryos to weathered Medium South American Crude (MESA) and a 1:10 dilution of Corexit 9500A:MESA solution with embryo hatching at day 19 as the endpoint. The second test exposed Rainbow trout (*Oncorhynchus mykiss*) embryos to HFO-7102, Nujol mineral oil (as control), 1:20 dilution of Corexit 9500A:HFO-7102 and a range of dilutions from 1:2.5-1:20 for Corexit 9500a:Nujol oil solutions. The authors reported a 19 day EC50 for *C. arengus* as 0.15 mg/L (normal development) and 1.02mg/L (hatching success) of oil concentrations measured by flurometry. The use of a control dispersant/oil mixture resulted in no effects observed when exposed to *O. mykiss* and the authors suggest Corexit 9500A was not exposed to embryos due to binding with oil particles, 24-day LC50 values for Corexit:HFO is reported as .052%v/v and Corexit alone .00311%v/v. Mu, Jin, Ma, Lin and Wang (2014) conducted oil only WAF (Petrozuata heavy crude), oil/chemical dispersant mixtures CE-WAF (Petrozuata heavy crude and Shuangxiang No. 1), and oil/biological dispersant mixtures BE-WAF (Petrozuata heavy crude and Weipu MD-66) 25 day chronic toxicity tests using the standard test species marine medaka, *Orzias melastigma*, with deformity being primary endpoint. The authors reported EC20 values expressed as total petroleum hydrocarbons resulting in the following concentrations: 0.56 mg/L WAF, 0.78 mg/L CE-WAF, and 5.6 mg/L BE-WAF.

Soil and Sediment Organisms and Functions.

Oil-dispersant toxicity effects on reproductive gene expression using a standard test worm, *Caenorhabditis*

elegans and test solutions mixed with crude oil obtained from the Macondo well, MC-252, and Corexit 9500A was evaluated (Polli et al 2014) investigated. Effects of increased development of apoptotic cells in reproductive regions of the worm were observed at all concentrations reported as dilutions of a stock 1:20 dispersant-oil solution ranging from 500x, 2000x, and 5000x.

Field collected marsh sediment received doses of Macondo 252 crude oil (1 mL), Corexit 9500A (0.1 mL) and oil-dispersant mixture (1 mL oil combined with 0.1 mL Corexit 9500A) in a study conducted in 2 phases marked by nitrate additions (50ppm and 100ppm) and monitored N₂O and CO₂ productions to determine changes in organic matter mineralization and denitrification (Shi and Yu 2014). The authors reported no significant changes to oil only exposures, significant inhibition of denitrification and increased mineralization when exposed to dispersants. Sediments were dosed with IFO-15 crude oil and IFO-15/Corexit 9500 dispersant mixture to investigate effects of the dispersant on natural hydrocarbon degradation in marine sediment (Macías-Zamora et al. 2014). The authors reported no significant differences between oil only and dispersant/oil mixtures for hydrocarbon degradation, aromatics were readily degraded, and PAH degradation increased linearly with oxygen availability.

Tissue Culture Toxicity. Zheng et al. (2014) conducted MMT cytotoxicity assays with skin cancer cells (B16/BL6), neuronal cells (H19-7), human astrocytoma cells (1321N1), kidney cell I (HEK-293), kidney cell II (HK2) and Corexit 9500 as the toxicant. Test concentrations ranges were 20, 40, 80, 160, and 200 ppm

and exposures lasted 48hrs. The authors reported LC50 values as 16 ppm, 33 ppm, 70 ppm, 93 ppm, 95 ppm for BL16/B6, 1321N1, H19-7, HEK293, and HK-2, respectively. Sperm whale skin fibroblast cells were field collected by Wise et al. (2014) and exposed to Corexit 9500 and 9527 for 24 hours to determine cytotoxicity and clastogenicity effects. Using a clonogenic assay, the authors reported Corexit 9500 induced increased cytotoxic effects with 3% survival at the highest concentration tested of 0.1% and Corexit 9527 resulted in 26% survival at the highest concentration of 0.1%. A chromosomal aberration assay determined Corexit 9527 induced increased genotoxic effects when compared to Corexit 9500.

Comparison of Dispersants and House-hold Cleaners. Word et al. (2014) contracted two accredited EPA Laboratories to conduct 48hr and 96hr acute toxicity tests to compare LC50's among 8 household cleaning products and Corexit 9500 using standard test organisms *Americamysis (=Mysidopsis) bahia* and *Menidia beryllina*. The authors reported mean LC50 results from 48hr *A. bahia* tests for the two laboratories (lab A-Lab B) to be 28.9-35.1 ppm, 48.0-91.2 ppm, 13.3-20.7 ppm, 32.9-45.3 ppm, 35.4-67.7 ppm, 177-413 ppm, 10.7-12.4 ppm, 328-387 ppm, 40-45.3 ppm for Dawn dish soap, Restore the Earth dish soap, Palmolive dish soap, Green Works dish soap, Cascade dish detergent, Johnson's Baby shampoo, Tide Laundry detergent, Green Works all purpose cleaner, and Corexit 9500, respectively. The reported mean LC50 values from 96hr *M. beryllina* tests for the two laboratories (Lab A- Lab B) are 8.9-8.3 ppm, 26.9-21.2 ppm, 7.1-5.4 ppm, 7.8-9.9 ppm, 56.6-55.6 ppm, 38.8-42.0 ppm, 4-11.8

ppm, 386-591 ppm, 35.4-110 ppm for the same respective test solutions as above.

Disclaimer

Reference herein to any specific commercial products, process, or service by trade name, trademark, manufacturer, or otherwise, does not necessarily constitute or imply its endorsement, recommendation, or favoring by the United States Government. The views and opinions of authors expressed herein do not necessarily state or reflect those of the United States Government, and shall not be used for advertising or product endorsement purposes.

References

- Acampora, H.; Schuyler, Q. A.; Townsend, K. A.; Hardesty, B. D. (2014) Comparing Plastic Ingestion in Juvenile and Adult Stranded Short-Tailed Shearwaters (*Puffinus tenuirostris*) in Eastern Australia. *Marine Pollution Bulletin*, **78**, 63-68.
- Adams, J.; Sweezy, M.; Hodson, P. V. (2014) Oil and Oil Dispersant Do Not Cause Synergistic Toxicity to Fish Embryos. *Environmental Toxicology and Chemistry*, **33**, 107-114.
- Adimey, N. M.; Hudak, C. A.; Powell, J. R.; Bassos-Hull, K.; Foley, A.; Farmer, N. A.; White, L.; Minch, K. (2014) Fishery Gear Interactions from Stranded Bottlenose Dolphins, Florida Manatees and Sea Turtles in Florida, USA. *Marine Pollution Bulletin*, **81**, 103-115.
- Almeda, R.; Hyatt, C.; Buskey, E. J. (2014a) Toxicity of

- Dispersant Corexit 9500a and Crude Oil to Marine Microzooplankton. *Ecotoxicology and Environmental Safety*, **106**, 76-85.
- Almeda, R.; Baca, S.; Hyatt, C.; Buskey, E. J. (2014b) Ingestion and Sublethal Effects of Physically and Chemically Dispersed Crude Oil on Marine Planktonic Copepods. *Ecotoxicology*, **23**, 988-1003.
- Almeda, R.; Bona, S.; Foster, C. R.; Buskey, E. J. (2014c) Dispersant Corexit 9500a and Chemically Dispersed Crude Oil Decreases the Growth Rates of Meroplanktonic Barnacle Nauplii (*Amphibalanus improvisus*) and Tornaria Larvae (*Schizocardium* sp.). *Marine Environmental Research*, **99**, 212-217.
- Amoozadeh, E.; Malek, M.; Rashidinejad, R.; Nabavi, S.; Karbassi, M.; Ghayoumi, R.; Ghorbanzadeh-Zafarani, G.; Salehi, H.; Sures, B. (2014) Marine Organisms as Heavy Metal Bioindicators in the Persian Gulf and the Gulf of Oman. *Environmental Science and Pollution Research*, **21**, 2386-2395.
- Anderson, J. A.; Alford, A. B. (2014) Ghost Fishing Activity in Derelict Blue Crab Traps in Louisiana. *Marine Pollution Bulletin*, **79**, 261-267.
- Anderson, J. A.; Kuhl, A. J.; Anderson, A. N. (2014) Toxicity of Oil and Dispersed Oil on Juvenile Mud Crabs, *Rhithropanopeus harrisi*. [Bulletin of Environmental Contamination and Toxicology](#), **92**, 375-380.
- Apostolopoulou, M.-V.; Monteyne, E.; Krikonis, K.; Pavlopoulos, K.; Roose, P.; Dehairs, F. (2014) Monitoring Polycyclic Aromatic Hydrocarbons in the Northeast Aegean Sea Using *Posidonia oceanica* Seagrass and Synthetic Passive Samplers. *Marine Pollution Bulletin*, **87**, 338-344.
- Arai, T. (2014) Variation in Organochlorine Accumulation in Relation to the Life History of the Japanese Eel *Anguilla japonica*. *Marine Pollution Bulletin*, **80**, 186-193.
- Arantes de Carvalho, G. G.; Manoel Degaspari, I. A.; Branco, V.; Canario, J.; de Amorim, A. F.; Kennedy, V. H.; Ferreira, J. R. (2014) Assessment of Total and Organic Mercury Levels in Blue Sharks (*Prionace glauca*) from the South and Southeastern Brazilian Coast. *Biological Trace Element Research*, **159**, 128-134.
- Arrhenius, A.; Backhaus, T.; Hilvarsson, A.; Wendt, I.; Zgrundo, A.; Blanck, H. (2014) A Novel Bioassay for Evaluating the Efficacy of Biocides to Inhibit Settling and Early Establishment of Marine Biofilms. *Marine Pollution Bulletin*, **87**, 292-299.
- Arthur, C.; Sutton-Grier, A. E.; Murphy, P.; Bamford, H. (2014) Out of Sight but Not out of Mind: Harmful Effects of Derelict Traps in Selected US Coastal Waters. *Marine Pollution Bulletin*, **86**, 19-28.
- Bakir, A.; Rowland, S. J.; Thompson, R. C. (2014a) Enhanced Desorption of Persistent Organic

- Pollutants from Microplastics under Simulated Physiological Conditions. *Environmental Pollution*, **185**, 16-23.
- Bakir, A.; Rowland, S. J.; Thompson, R. C. (2014b) Transport of Persistent Organic Pollutants by Microplastics in Estuarine Conditions. *Estuarine Coastal and Shelf Science*, **140**, 14-21.
- Balbi, T.; Smerilli, A.; Fabbri, R.; Ciacci, C.; Montagna, M.; Grasselli, E.; Brunelli, A.; Pojana, G.; Marcomini, A.; Gallo, G.; Canesi, L. (2014) Co-Exposure to Ni²⁺ and Cd²⁺ Results in Interactive Effects on Biomarker Responses but Not in Increased Toxicity in the Marine Bivalve *M. galloprovincialis*. *Science of the Total Environment*, **493**, 355-364.
- Balcioglu, E. B.; Aksu, A.; Balkis, N.; Ozturk, B. (2014) T-Pah Contamination in Mediterranean Mussels (*Mytilus galloprovincialis*, Lamarck, 1819) at Various Stations of the Turkish Straits System. *Marine Pollution Bulletin*, **88**, 344-346.
- Banaru, D.; Carlotti, F.; Barani, A.; Gregori, G.; Neffati, N.; Harmelin-Vivien, M. (2014) Seasonal Variation of Stable Isotope Ratios of Size-Fractionated Zooplankton in the Bay of Marseille (Nw Mediterranean Sea). *Journal of Plankton Research*, **36**, 145-156.
- Bao, V. W. W.; Lui, G. C. S.; Leung, K. M. Y. (2014) Acute and Chronic Toxicities of Zinc Pyriithione Alone and in Combination with Copper to the Marine Copepod *Tigriopus japonicus*. *Aquatic Toxicology*, **157**, 81-93.
- Barhoumi, B.; Clerandau, C.; Gourves, P.-Y.; Le Menach, K.; El Megdiche, Y.; Peluhet, L.; Budzinski, H.; Baudrimont, M.; Driss, M. R.; Cachot, J. (2014) Pollution Biomonitoring in the Bizerte Lagoon (Tunisia), Using Combined Chemical and Biomarker Analyses in Grass Goby, *Zosterisessor ophiocephalus* (Teleostei, Gobiidae). *Marine Environmental Research*, **101**, 184-195.
- Barreiros, J. P.; Raykov, V. S. (2014) Lethal Lesions and Amputation Caused by Plastic Debris and Fishing Gear on the Loggerhead Turtle *Caretta caretta* (Linnaeus, 1758). Three Case Reports from Terceira Island, Azores (Ne Atlantic). *Marine Pollution Bulletin*, **86**, 518-522.
- Barros, A.; Alvarez, D.; Velando, A. (2014) Long-Term Reproductive Impairment in a Seabird after the Prestige Oil Spill. *Biology Letters*, **10**.
- Baulch, S.; Perry, C. (2014) Evaluating the Impacts of Marine Debris on Cetaceans. *Marine Pollution Bulletin*, **80**, 210-221.
- Becherucci, M. E.; Benavides, H.; Vallarino, E. A. (2014) Effect of Taxonomic Aggregation in Macroalgae Assemblages in a Rocky Shore of Mar Del Plata, Argentina, Southwest Atlantic Ocean. *Thalassas*, **30**, 9-20.
- Beeden, R. J.; Turner, M. A.; Dryden, J.; Merida, F.; Goudkamp, K.; Malone, C.; Marshall, P. A.; Birtles, A.; Maynard, J. A. (2014) Rapid Survey Protocol That Provides Dynamic Information on Reef Condition to Managers of the Great Barrier

- Reef. *Environmental Monitoring and Assessment*, **186**, 8527-8540.
- Bellas, J.; Albentosa, M.; Vidal-Linan, L.; Besada, V.; Angeles Franco, M.; Fumega, J.; Gonzalez-Quijano, A.; Vinas, L.; Beiras, R. (2014) Combined Use of Chemical, Biochemical and Physiological Variables in Mussels for the Assessment of Marine Pollution Along the N-Nw Spanish Coast. *Marine Environmental Research*, **96**, 105-117.
- Bendell, L. I. (2014) Community Composition of the Intertidal in Relation to the Shellfish Aquaculture Industry in Coastal British Columbia, Canada. *Aquaculture*, **433**, 384-394.
- Beretta, M.; Britto, V.; Tavares, T. M.; Teixeira da Silva, S. M.; Pletsch, A. L. (2014) Occurrence of Pharmaceutical and Personal Care Products (PPCPs) in Marine Sediments in the Todos Os Santos Bay and the North Coast of Salvador, Bahia, Brazil. *Journal of Soils and Sediments*, **14**, 1278-1286.
- Bhattacharya, B. D.; Hwang, J.-S.; Tseng, L.-C.; Sarkar, S. K.; Rakshit, D.; Mitra, S. (2014) Bioaccumulation of Trace Elements in Dominant Mesozooplankton Group Inhabiting in the Coastal Regions of Indian Sundarban Mangrove Wetland. *Marine Pollution Bulletin*, **87**, 345-351.
- Bianchi, T. S.; Osburn, C.; Shields, M. R.; Yvon-Lewis, S.; Young, J.; Guo, L.; Zhou, Z. (2014) Deepwater Horizon Oil in Gulf of Mexico Waters after 2 Years: Transformation into the Dissolved Organic Matter Pool. *Environmental Science & Technology*, **48**, 9288-9297.
- Bik, H. M. (2014) Deciphering Diversity and Ecological Function from Marine Metagenomes. *Biological Bulletin*, **227**, 107-116.
- Bilkovic, D. M.; Havens, K.; Stanhope, D.; Angstadt, K. (2014) Derelict Fishing Gear in Chesapeake Bay, Virginia: Spatial Patterns and Implications for Marine Fauna. *Marine Pollution Bulletin*, **80**, 114-123.
- Binnington, M. J.; Wania, F. (2014) Clarifying Relationships between Persistent Organic Pollutant Concentrations and Age in Wildlife Biomonitoring: Individuals, Cross-Sections, and the Roles of Lifespan and Sex. *Environmental Toxicology and Chemistry*, **33**, 1415-1426.
- Bo, M.; Bava, S.; Canese, S.; Angiolillo, M.; Cattaneo-Vietti, R.; Bavestrello, G. (2014) Fishing Impact on Deep Mediterranean Rocky Habitats as Revealed by ROV Investigation. *Biological Conservation*, **171**, 167-176.
- Boalt, E.; Miller, A.; Dahlgren, H. (2014) Distribution of Cadmium, Mercury, and Lead in Different Body Parts of Baltic Herring (*Clupea harengus*) and Perch (*Perca fluviatilis*): Implications for Environmental Status Assessments. *Marine Pollution Bulletin*, **78**, 130-136.
- Bode, A.; Fernandez, C.; Mompean, C.; Parra, S.; Rozada, F.; Valencia-Vila, J.; Viana, I. G. (2014) Differential Processing of Anthropogenic Carbon and Nitrogen in Benthic Food Webs of a Coruna

- (NW Spain) Traced by Stable Isotopes. *Deep-Sea Research Part II-Topical Studies in Oceanography*, **106**, 198-206.
- Boldina, I.; Beninger, P. G.; Le Coz, M. (2014) Effect of Long-Term Mechanical Perturbation on Intertidal Soft-Bottom Meiofaunal Community Spatial Structure. *Journal of Sea Research*, **85**, 85-91.
- Bolognesi, C.; Cirillo, S. (2014) Genotoxicity Biomarkers in Aquatic Bioindicators. *Current Zoology*, **60**, 273-284.
- Bond, A. L.; Provencher, J. F.; Daoust, P.-Y.; Lucas, Z. N. (2014) Plastic Ingestion by Fulmars and Shearwaters at Sable Island, Nova Scotia, Canada. *Marine Pollution Bulletin*, **87**, 68-75.
- Bossus, M. C.; Guler, Y. Z.; Short, S. J.; Morrison, E. R.; Ford, A. T. (2014) Behavioural and Transcriptional Changes in the Amphipod *Echinogammarus marinus* Exposed to Two Antidepressants, Fluoxetine and Sertraline. *Aquatic Toxicology*, **151**, 46-56.
- Boulcott, P.; Millar, C. P.; Fryer, R. J. (2014) Impact of Scallop Dredging on Benthic Epifauna in a Mixed-Substrate Habitat. *Ices Journal of Marine Science*, **71**, 834-844.
- Brenner, M.; Broeg, K.; Frickenhaus, S.; Buck, B. H.; Koehler, A. (2014) Multi-Biomarker Approach Using the Blue Mussel (*Mytilus edulis* L.) to Assess the Quality of Marine Environments: Season and Habitat-Related Impacts. *Marine Environmental Research*, **95**, 13-27.
- Brette, F.; Machado, B.; Cros, C.; Incardona, J. P.; Scholz, N. L.; Block, B. A. (2014) Crude Oil Impairs Cardiac Excitation-Contraction Coupling in Fish. *Science*, **343**, 772-776.
- Brind'Amour, A.; Laffargue, P.; Morin, J.; Vaz, S.; Foveau, A.; Le Bris, H. (2014) Morphospecies and Taxonomic Sufficiency of Benthic Megafauna in Scientific Bottom Trawl Surveys. *Continental Shelf Research*, **72**, 1-9.
- Browne, M. A.; Niven, S. J.; Galloway, T. S.; Rowland, S. J.; Thompson, R. C. (2013) Microplastic Moves Pollutants and Additives to Worms, Reducing Functions Linked to Health and Biodiversity. *Current Biology*, **23**, 2388-2392.
- Bu-Olayan, A. H.; Thomas, B. V. (2014) Dispersion Model and Bioaccumulation Factor Validating Trace Metals in Sea Bream Inhabiting Wastewater Drain Outfalls. *International Journal of Environmental Science and Technology*, **11**, 795-804.
- Buffet, P.-E.; Poirier, L.; Zalouk-Vergnoux, A.; Lopes, C.; Amiard, J.-C.; Gaudin, P.; Risso-de Faverney, C.; Guibbolini, M.; Gilliland, D.; Perrein-Ettajani, H.; Valsami-Jones, E.; Mouneyrac, C. (2014a) Biochemical and Behavioural Responses of the Marine Polychaete *Hediste diversicolor* to Cadmium Sulfide Quantum Dots (CDS QDs): Waterborne and Dietary Exposure. *Chemosphere*, **100**, 63-70.
- Buffet, P.-E.; Zalouk-Vergnoux, A.; Chatel, A.; Berthet, B.; Metais, I.; Perrein-Ettajani, H.; Poirier, L.; Luna-Acosta, A.; Thomas-Guyon, H.; Risso-de

- Faverney, C.; Guibbolini, M.; Gilliland, D.; Valsami-Jones, E.; Mouneyrac, C. (2014b) A Marine Mesocosm Study on the Environmental Fate of Silver Nanoparticles and Toxicity Effects on Two Endobenthic Species: The Ragworm *Hediste diversicolor* and the Bivalve Mollusc *Scrobicularia plana*. *Science of the Total Environment*, **470**, 1151-1159.
- Burd, B. J.; Macdonald, T. A.; Macdonald, R. W.; Ross, P. S. (2014) Distribution and Uptake of Key Polychlorinated Biphenyl and Polybrominated Diphenyl Ether Congeners in Benthic Infauna Relative to Sediment Organic Enrichment. *Archives of Environmental Contamination and Toxicology*, **67**, 310-334.
- Burns, C. M. B.; Olin, J. A.; Woltmann, S.; Stouffer, P. C.; Taylor, S. S. (2014) Effects of Oil on Terrestrial Vertebrates: Predicting Impacts of the Macondo Blowout. *Bioscience*, **64**, 820-828.
- Cabral-Oliveira, J.; Bevilacqua, S.; Terlizzi, A.; Pardal, M. A. (2014) Are Eulittoral Assemblages Suitable for Detecting the Effects of Sewage Discharges in Atlantic and Mediterranean Coastal Areas? *Italian Journal of Zoology*, **81**, 584-592.
- Cabral-Oliveira, J.; Dolbeth, M.; Pardal, M. A. (2014) Impact of Sewage Pollution on the Structure and Functioning of a Rocky Shore Benthic Community. *Marine and Freshwater Research*, **65**, 750-758.
- Calder, D. R.; Choong, H. H. C.; Carlton, J. T.; Chapman, J. W.; Miller, J. A.; Geller, J. (2014) Hydroids (Cnidaria: Hydrozoa) from Japanese Tsunami Marine Debris Washing Ashore in the Northwestern United States. *Aquatic Invasions*, **9**, 425-440.
- Camedda, A.; Marra, S.; Matiddi, M.; Massaro, G.; Coppa, S.; Perilli, A.; Ruiu, A.; Briguglio, P.; de Lucia, G. A. (2014) Interaction between Loggerhead Sea Turtles (*Caretta caretta*) and Marine Litter in Sardinia (Western Mediterranean Sea). *Marine Environmental Research*, **100**, 25-32.
- Canario, J.; Poissant, L.; Pilote, M.; Blaise, C.; Constant, P.; Ferard, J.-F.; Gagne, F. (2014) Toxicity Survey of Canadian Arctic Marine Sediments. *Journal of Soils and Sediments*, **14**, 196-203.
- Canesi, L.; Frenzilli, G.; Balbi, T.; Bernardeschi, M.; Ciacci, C.; Corsolini, S.; Della Torre, C.; Fabbri, R.; Faleri, C.; Focardi, S.; Guidi, P.; Kocan, A.; Marcomini, A.; Mariottini, M.; Nigro, M.; Pozo-Gallardo, K.; Rocco, L.; Scarcelli, V.; Smerilli, A.; Corsi, I. (2014) Interactive Effects of N-Tio2 and 2,3,7,8-TCDD on the Marine Bivalve *Mytilus galloprovincialis*. *Aquatic Toxicology*, **153**, 53-65.
- Carregosa, V.; Velez, C.; Soares, A. M. V. M.; Figueira, E.; Freitas, R. (2014) Physiological and Biochemical Responses of Three Veneridae Clams Exposed to Salinity Changes. *Comparative Biochemistry and Physiology B-Biochemistry & Molecular Biology*, **177**, 1-9.
- Carroll, A. G.; Przeslawski, R.; Radke, L. C.; Black, J.

- R.; Picard, K.; Moreau, J. W.; Haese, R. R.; Nichol, S. (2014) Environmental Considerations for Subseabed Geological Storage of CO₂: A Review. *Continental Shelf Research*, **83**, 116-128.
- Castege, I.; Milon, E.; Pautrizel, F. (2014) Response of Benthic Macrofauna to an Oil Pollution: Lessons from the "Prestige" Oil Spill on the Rocky Shore of Guethary (South of the Bay of Biscay, France). *Deep-Sea Research Part II-Topical Studies in Oceanography*, **106**, 192-197.
- Castro-Bugallo, A.; Gonzalez-Fernandez, A.; Guisande, C.; Barreiro, A. (2014) Comparative Responses to Metal Oxide Nanoparticles in Marine Phytoplankton. *Archives of Environmental Contamination and Toxicology*, **67**, 483-493.
- Chalkiadaki, O.; Dassenakis, M.; Paraskevopoulou, V.; Lydakis-Simantiris, N. (2014) Experimental Study of Cadmium Bioaccumulation in Three Mediterranean Marine Bivalve Species: Correlation with Selected Biomarkers. *Pure and Applied Chemistry*, **86**, 1189-1204.
- Chang, S. E.; Stone, J.; Demes, K.; Piscitelli, M. (2014) Consequences of Oil Spills: A Review and Framework for Informing Planning. *Ecology and Society*, **19**.
- Chen, L.; Zhang, H.; Sun, J.; Wong, Y.-H.; Han, Z.; Au, D. W. T.; Bajic, V. B.; Qian, P.-Y. (2014) Proteomic Changes in Brain Tissues of Marine Medaka (*Oryzias melastigma*) after Chronic Exposure to Two Antifouling Compounds: Butenolide and 4,5-Dichloro-2-N-Octyl-4-Isothiazolin-3-One (Dcoit). *Aquatic Toxicology*, **157**, 47-56.
- Choi, J. Y.; Yang, D. B.; Hong, G. H.; Shin, K. H. (2014) Distribution and Bioaccumulation of Polychlorinated Biphenyls and Organochlorine Pesticides Residues in Sediments and Manila Clams (*Ruditapes philippinarum*) from Along the Mid-Western Coast of Korea. *Marine Pollution Bulletin*, **85**, 672-678.
- Choi, M.-H.; Hwang, Y.; Lee, H. U.; Kim, B.; Lee, G.-W.; Oh, Y.-K.; Andersen, H. R.; Lee, Y.-C.; Huh, Y. S. (2014) Aquatic Ecotoxicity Effect of Engineered Aminoclay Nanoparticles. *Ecotoxicology and Environmental Safety*, **102**, 34-41.
- Chua, E. M.; Shimeta, J.; Nugegoda, D.; Morrison, P. D.; Clarke, B. O. (2014) Assimilation of Polybrominated Diphenyl Ethers from Microplastics by the Marine Amphipod, *Allorchestes compressa*. *Environmental Science & Technology*, **48**, 8127-8134.
- Clarke, S.; Tully, O. (2014) Baci Monitoring of Effects of Hydraulic Dredging for Cockles on Intertidal Benthic Habitats of Dundalk Bay, Ireland. *Journal of the Marine Biological Association of the United Kingdom*, **94**, 1451-1464.
- Cohen, J. H.; McCormick, L. R.; Burkhardt, S. M. (2014) Effects of Dispersant and Oil on Survival and Swimming Activity in a Marine Copepod. *Bulletin of Environmental Contamination and*

- Toxicology*, **92**, 381-387.
- Cozzi, S.; Mistaro, A.; Sparnocchia, S.; Colugnati, L.; Bajt, O.; Toniatti, L. (2014) Anthropogenic Loads and Biogeochemical Role of Urea in the Gulf of Trieste. *Science of the Total Environment*, **493**, 271-281.
- Crespo-Medina, M.; Meile, C. D.; Hunter, K. S.; Diercks, A. R.; Asper, V. L.; Orphan, V. J.; Tavormina, P. L.; Nigro, L. M.; Battles, J. J.; Chanton, J. P.; Shiller, A. M.; Joung, D. J.; Amon, R. M. W.; Bracco, A.; Montoya, J. P.; Villareal, T. A.; Wood, A. M.; Joye, S. B. (2014) The Rise and Fall of Methanotrophy Following a Deepwater Oil-Well Blowout. *Nature Geoscience*, **7**, 423-427.
- Cresson, P.; Fabri, M. C.; Bouchoucha, M.; Papa, C. B.; Chavanon, F.; Jadaud, A.; Knoery, J.; Miralles, F.; Cossa, D. (2014) Mercury in Organisms from the Northwestern Mediterranean Slope: Importance of Food Sources. *Science of the Total Environment*, **497**, 229-238.
- Crowe, K. M.; Newton, J. C.; Kaltenboeck, B.; Johnson, C. (2014) Oxidative Stress Responses of Gulf Killifish Exposed to Hydrocarbons from the Deepwater Horizon Oil Spill: Potential Implications for Aquatic Food Resources. *Environmental Toxicology and Chemistry*, **33**, 370-374.
- Dadar, M.; Peyghan, R.; Memari, H. R. (2014) Evaluation of the Bioaccumulation of Heavy Metals in White Shrimp (*Litopenaeus vannamei*) Along the Persian Gulf Coast. *Bulletin of Environmental Contamination and Toxicology*, **93**, 339-343.
- Dahms, S.; van der Bank, F. H.; Greenfield, R. (2014) A Baseline Study of Metal Contamination Along the Namibian Coastline for *Perna perna* and *Choromytilus meridionalis*. *Marine Pollution Bulletin*, **85**, 297-305.
- Daief, Z.; Borja, A.; Joulami, L.; Azzi, M.; Fahde, A.; Bazairi, H. (2014) Assessing Benthic Ecological Status of Urban Sandy Beaches (Northeast Atlantic, Morocco) Using M-AMBI. *Ecological Indicators*, **46**, 586-595.
- Dannheim, J.; Brey, T.; Schroeder, A.; Mintenbeck, K.; Knust, R.; Arntz, W. E. (2014) Trophic Look at Soft-Bottom Communities - Short-Term Effects of Trawling Cessation on Benthos. *Journal of Sea Research*, **85**, 18-28.
- De Backer, A.; Van Hoey, G.; Coates, D.; Vanaverbeke, J.; Hostens, K. (2014) Similar Diversity-Disturbance Responses to Different Physical Impacts: Three Cases of Small-Scale Biodiversity Increase in the Belgian Part of the North Sea. *Marine Pollution Bulletin*, **84**, 251-262.
- de Souza, K. B.; Jutfelt, F.; Kling, P.; Forlin, L.; Sturve, J. (2014) Effects of Increased CO₂ on Fish Gill and Plasma Proteome. *Plos One*, **9**.
- Dedeh, A.; Ciutat, A.; Tran, D.; Bourdineaud, J.-P. (2014) DNA Alterations Triggered by Environmentally Relevant Polymetallic Concentrations in Marine Clams *Ruditapes philippinarum* and Polychaete Worms *Hediste diversicolor*. *Archives of*

- Environmental Contamination and Toxicology*, **67**, 651-658.
- Del-Pilar-Ruso, Y.; Antonio de-la-Ossa-Carretero, J.; Gimenez-Casaldueiro, F.; Luis Sanchez-Lizaso, J.; San Martin, G. (2014) Checking the Concurrence among Macrobenthic Organism Distribution Patterns at Different Taxonomic Scales in Relation to Environmental Factors. *Journal of Sea Research*, **86**, 49-57.
- Della Torre, C.; Bergami, E.; Salvati, A.; Faleri, C.; Cirino, P.; Dawson, K. A.; Corsi, I. (2014) Accumulation and Embryotoxicity of Polystyrene Nanoparticles at Early Stage of Development of Sea Urchin Embryos *Paracentrotus lividus*. *Environmental Science & Technology*, **48**, 12302-12311.
- DeLorenzo, M. E.; Key, P. B.; Chung, K. W.; Sapozhnikova, Y.; Fulton, M. H. (2014) Comparative Toxicity of Pyrethroid Insecticides to Two Estuarine Crustacean Species, *Americamysis bahia* and *Palaemonetes pugio*. *Environmental Toxicology*, **29**, 1099-1106.
- Delshad, N.; Emtiazjoo, M.; Khezri, M. (2014) Toxicity Effect of Oil Spill Dispersants on *Litopenaeus vannamei*. *International Journal of Environmental Research*, **8**, 1027-1030.
- Deng, X.; Pan, L.; Miao, J.; Cai, Y.; Hu, F. (2014) Digital Gene Expression Analysis of Reproductive Toxicity of Benzo a Pyrene in Male Scallop *Chlamys farreri*. *Ecotoxicology and Environmental Safety*, **110**, 190-196.
- Diepens, N. J.; Arts, G. H. P.; Brock, T. C. M.; Smidt, H.; Van den Brink, P. J.; Van den Heuvel-Greve, M. J.; Koelmans, A. A. (2014) Sediment Toxicity Testing of Organic Chemicals in the Context of Prospective Risk Assessment: A Review. *Critical Reviews in Environmental Science and Technology*, **44**, 255-302.
- Diez, I.; Santolaria, A.; Muguerza, N.; Gorostiaga, J. M. (2014) Capacity for Recovery of Rocky Subtidal Assemblages Following Pollution Abatement in a Scenario of Global Change. *Marine Pollution Bulletin*, **86**, 197-209.
- Dodder, N. G.; Maruya, K. A.; Ferguson, P. L.; Grace, R.; Kiosterhaus, S.; La Guardia, M. J.; Lauenstein, G. G.; Ramirez, J. (2014) Occurrence of Contaminants of Emerging Concern in Mussels (*Mytilus spp.*) Along the California Coast and the Influence of Land Use, Storm Water Discharge, and Treated Wastewater Effluent. *Marine Pollution Bulletin*, **81**, 340-346.
- Dolores Basallote, M.; De Orte, M. R.; Angel DelValls, T.; Riba, I. (2014) Studying the Effect of CO₂-Induced Acidification on Sediment Toxicity Using Acute Amphipod Toxicity Test. *Environmental Science & Technology*, **48**, 8864-8872.
- Donnelly-Greenan, E. L.; Harvey, J. T.; Nevins, H. M.; Hester, M. M.; Walker, W. A. (2014) Prey and Plastic Ingestion of Pacific Northern Fulmars (*Fulmarus glacialis rogersii*) from Monterey Bay, California. *Marine Pollution Bulletin*, **85**,

- 214-224.
- dos Santos, D. M.; Santos, G. S.; Cestari, M. M.; de Oliveira Ribeiro, C. A.; Silva de Assis, H. C.; Yamamoto, F.; Guiloski, I. C.; Rodrigues de Marchi, M. R.; Montone, R. C. (2014) Bioaccumulation of Butyltins and Liver Damage in the Demersal Fish *Cathorops spixii* (Siluriformes, Ariidae). *Environmental Science and Pollution Research*, **21**, 3166-3174.
- Dutton, J.; Fisher, N. S. (2014) Modeling Metal Bioaccumulation and Tissue Distribution in Killifish (*Fundulus heteroclitus*) in Three Contaminated Estuaries. *Environmental Toxicology and Chemistry*, **33**, 89-101.
- Edmunds, P. J.; Gray, S. C. (2014) The Effects of Storms, Heavy Rain, and Sedimentation on the Shallow Coral Reefs of St. John, Us Virgin Islands. *Hydrobiologia*, **734**, 143-158.
- El-Din, N. G. S.; Mohamedein, L. I.; El-Moselhy, K. M. (2014) Seaweeds as Bioindicators of Heavy Metals Off a Hot Spot Area on the Egyptian Mediterranean Coast During 2008-2010. *Environmental Monitoring and Assessment*, **186**, 5865-5881.
- Engel, A. S.; Gupta, A. A. (2014) Regime Shift in Sandy Beach Microbial Communities Following Deepwater Horizon Oil Spill Remediation Efforts. *Plos One*, **9**.
- Fabbri, R.; Montagna, M.; Balbi, T.; Raffo, E.; Palumbo, F.; Canesi, L. (2014) Adaptation of the Bivalve Embryotoxicity Assay for the High Throughput Screening of Emerging Contaminants in *Mytilus galloprovincialis*. *Marine Environmental Research*, **99**, 1-8.
- Faimali, M.; Garaventa, E.; Piazza, V.; Costa, E.; Greco, G.; Mazzola, V.; Beltrandi, M.; Bongiovanni, E.; Lavorano, S.; Gnone, G. (2014) Ephyra Jellyfish as a New Model for Ecotoxicological Bioassays. *Marine Environmental Research*, **93**, 93-101.
- Farrington, J. W. (2014) Oil Pollution in the Marine Environment II: Fates and Effects of Oil Spills. *Environment*, **56**, 16-31.
- Farrington, J. W.; Takada, H. (2014) Persistent Organic Pollutants (POPs), Polycyclic Aromatic Hydrocarbons (PAHs), and Plastics Examples of the Status, Trend, and Cycling of Organic Chemicals of Environmental Concern in the Ocean. *Oceanography*, **27**, 196-213.
- Field, L. J.; Norton, S. B. (2014) Regional Models for Sediment Toxicity Assessment. *Environmental Toxicology and Chemistry*, **33**, 708-717.
- Fisher, C. R.; Demopoulos, A. W. J.; Cordes, E. E.; Baums, I. B.; White, H. K.; Bourque, J. R. (2014a) Coral Communities as Indicators of Ecosystem-Level Impacts of the Deepwater Horizon Spill. *Bioscience*, **64**, 796-807.
- Fisher, C. R.; Hsing, P.-Y.; Kaiser, C. L.; Yoerger, D. R.; Roberts, H. H.; Shedd, W. W.; Cordes, E. E.; Shank, T. M.; Berlet, S. P.; Saunders, M. G.; Larcom, E. A.; Brooks, J. M. (2014b) Footprint of Deepwater Horizon Blowout Impact to Deep-

- Water Coral Communities. *Proceedings of the National Academy of Sciences of the United States of America*, **111**, 11744-11749.
- Fitzgerald, T. P.; Gohlke, J. M. (2014) Contaminant Levels in Gulf of Mexico Reef Fish after the Deepwater Horizon Oil Spill as Measured by a Fishermen-Led Testing Program. *Environmental Science & Technology*, **48**, 1993-2000.
- Fong, C. C.; Shi, Y. F.; Yu, W. K.; Wei, F.; van de Merwe, J. P.; Chan, A. K. Y.; Ye, R.; Au, D. W. T.; Wu, R. S. S.; Yang, M. S. (2014) Itraq-Based Proteomic Profiling of the Marine Medaka (*Oryzias melastigma*) Gonad Exposed to Bde-47. *Marine Pollution Bulletin*, **85**, 471-478.
- Fossi, M. C.; Coppola, D.; Baini, M.; Giannetti, M.; Guerranti, C.; Marsili, L.; Panti, C.; de Sabata, E.; Clo, S. (2014) Large Filter Feeding Marine Organisms as Indicators of Microplastic in the Pelagic Environment: The Case Studies of the Mediterranean Basking Shark (*Cetorhinus maximus*) and Fin Whale (*Balaenoptera physalus*). *Marine Environmental Research*, **100**, 17-24.
- Foster, S. D.; Hosack, G. R.; Hill, N. A.; Barrett, N. S.; Lucieer, V. L. (2014) Choosing between Strategies for Designing Surveys: Autonomous Underwater Vehicles. *Methods in Ecology and Evolution*, **5**, 287-297.
- Fox, M.; Ohlason, C.; Sharpe, A. D.; Brown, R. J. (2014) The Use of a *Corophium volutator* Chronic Sediment Study to Support the Risk Assessment of Medetomidine for Marine Environments. *Environmental Toxicology and Chemistry*, **33**, 937-942.
- Franci, C. D.; Guillemette, M.; Pelletier, E.; Chastel, O.; Bonnefoi, S.; Verreault, J. (2014) Endocrine Status of a Migratory Bird Potentially Exposed to the Deepwater Horizon Oil Spill: A Case Study of Northern Gannets Breeding on Bonaventure Island, Eastern Canada. *Science of the Total Environment*, **473**, 110-116.
- Franciskovic-Bilinski, S.; Cukrov, N. (2014) A Critical Evaluation of Using Bulk Sediment Instead of Fine Fraction in Environmental Marine Studies, Investigated on Example of Rijeka Harbor, Croatia. *Environmental Earth Sciences*, **71**, 341-356.
- Fredericq, S.; Arakaki, N.; Camacho, O.; Gabriel, D.; Kravesky, D.; Self-Kravesky, S.; Rees, G.; Richards, J.; Sauvage, T.; Venera-Ponton, D.; Schmidt, W. E. (2014) A Dynamic Approach to the Study of Rhodoliths: A Case Study for the Northwestern Gulf of Mexico. *Cryptogamie Algologie*, **35**, 77-98.
- Freitas, R.; Martins, R.; Antunes, S.; Velez, C.; Moreira, A.; Cardoso, P.; Pires, A.; Soares, A. M. V. M.; Figueira, E. (2014) *Venerupis decussata* under Environmentally Relevant Lead Concentrations: Bioconcentration, Tolerance, and Biochemical Alterations. *Environmental Toxicology and Chemistry*, **33**, 2786-2794.
- Frias, J. P. G. L.; Otero, V.; Sobral, P. (2014) Evidence of

- Microplastics in Samples of Zooplankton from Portuguese Coastal Waters. *Marine Environmental Research*, **95**, 89-95.
- Friedlander, A. M.; Ballesteros, E.; Fay, M.; Sala, E. (2014) Marine Communities on Oil Platforms in Gabon, West Africa: High Biodiversity Oases in a Low Biodiversity Environment. *Plos One*, **9**.
- Fry, B.; Anderson, L. C. (2014) Minimal Incorporation of Deepwater Horizon Oil by Estuarine Filter Feeders. *Marine Pollution Bulletin*, **80**, 282-287.
- Fukuyama, A. K.; Shigenaka, G.; Coats, D. A. (2014) Status of Intertidal Infaunal Communities Following the Exxon Valdez Oil Spill in Prince William Sound, Alaska. *Marine Pollution Bulletin*, **84**, 56-69.
- Galgani, F.; Claro, F.; Depledge, M.; Fossi, C. (2014) Monitoring the Impact of Litter in Large Vertebrates in the Mediterranean Sea within the European Marine Strategy Framework Directive (MSFD): Constraints, Specificities and Recommendations. *Marine Environmental Research*, **100**, 3-9.
- Ganesh, T.; Rakhesh, M.; Raman, A. V.; Nanduri, S.; Moore, S.; Rajanna, B. (2014) Macrobenthos Response to Sewage Pollution in a Tropical Inshore Area. *Environmental Monitoring and Assessment*, **186**, 3553-3566.
- Garr, A. L.; Laramore, S.; Krebs, W. (2014) Toxic Effects of Oil and Dispersant on Marine Microalgae. [Bulletin of Environmental Contamination and Toxicology](#), **93**, 654-659.
- Giannakopoulou, L.; Neofitou, C. (2014) Heavy Metal Concentrations in *Mullus barbatus* and *Pagellus erythrinus* in Relation to Body Size, Gender, and Seasonality. *Environmental Science and Pollution Research*, **21**, 7140-7153.
- Goldstein, M. C.; Carson, H. S.; Eriksen, M. (2014) Relationship of Diversity and Habitat Area in North Pacific Plastic-Associated Rafting Communities. *Marine Biology*, **161**, 1441-1453.
- Gomes, I. D. L.; Lemos, M. F. L.; Soares, A. M. V. M.; Barata, C.; Faria, M. (2014) The Use of Cholinesterase as Potential Biomarker: In Vitro Characterization in the Polychaete *Capitella teleta*. *Marine Pollution Bulletin*, **85**, 179-185.
- Gomiero, A.; Viarengo, A. (2014) Effects of Elevated Temperature on the Toxicity of Copper and Oxytetracycline in the Marine Model, *Euplotes crassus*: A Climate Change Perspective. *Environmental Pollution*, **194**, 262-271.
- Gonzalez Carman, V.; Marcelo Acha, E.; Maxwell, S. M.; Albareda, D.; Campagna, C.; Mianzan, H. (2014) Young Green Turtles, *Chelonia mydas*, Exposed to Plastic in a Frontal Area of the Sw Atlantic. *Marine Pollution Bulletin*, **78**, 56-62.
- Gonzalez, S. A.; Stotz, W.; Lancellotti, D. (2014) Effects of the Discharge of Iron Ore Tailings on Subtidal Rocky-Bottom Communities in Northern Chile. *Journal of Coastal Research*, **30**, 500-514.
- Gonzalez-Rivero, M.; Bongaerts, P.; Beijbom, O.; Pizarro, O.; Friedman, A.; Rodriguez-Ramirez, A.; Upcroft, B.; Laffoley, D.; Kline, D.;

- Bailhache, C.; Vevers, R.; Hoegh-Guldberg, O. (2014) The Catlin Seaview Survey - Kilometre-Scale Seascape Assessment, and Monitoring of Coral Reef Ecosystems. *Aquatic Conservation-Marine and Freshwater Ecosystems*, **24**, 184-198.
- Grabowski, J. H.; Bachman, M.; Demarest, C.; Eayrs, S.; Harris, B. P.; Malkoski, V.; Packer, D.; Stevenson, D. (2014) Assessing the Vulnerability of Marine Benthos to Fishing Gear Impacts. *Reviews in Fisheries Science & Aquaculture*, **22**, 142-155.
- Graca, B.; Beldowska, M.; Wrzesien, P.; Zgrundo, A. (2014) Styrofoam Debris as a Potential Carrier of Mercury within Ecosystems. *Environmental Science and Pollution Research*, **21**, 2263-2271.
- Gray, J. L.; Kanagy, L. K.; Furlong, E. T.; Kanagy, C. J.; McCoy, J. W.; Mason, A.; Lauenstein, G. (2014) Presence of the Corexit Component Dioctyl Sodium Sulfosuccinate in Gulf of Mexico Waters after the 2010 Deepwater Horizon Oil Spill. *Chemosphere*, **95**, 124-130.
- Greenfield, R.; Brink, K.; Degger, N.; Wepener, V. (2014) The Usefulness of Transplantation Studies in Monitoring of Metals in the Marine Environment: South African Experience. *Marine Pollution Bulletin*, **85**, 566-573.
- Guerranti, C.; Baini, M.; Casini, S.; Focardi, S. E.; Giannetti, M.; Mancusi, C.; Marsili, L.; Perra, G.; Fossi, M. C. (2014) Pilot Study on Levels of Chemical Contaminants and Porphyrins in *Caretta caretta* from the Mediterranean Sea. *Marine Environmental Research*, **100**, 33-37.
- Haemer, J.; Gutow, L.; Koehler, A.; Saborowski, R. (2014) Fate of Microplastics in the Marine Lsopod *Idotea emarginata*. *Environmental Science & Technology*, **48**, 13451-13458.
- Han, J.; Won, E.-J.; Hwang, D.-S.; Shin, K.-H.; Lee, Y. S.; Leung, K. M.-Y.; Lee, S.-J.; Lee, J.-S. (2014) Crude Oil Exposure Results in Oxidative Stress-Mediated Dysfunctional Development and Reproduction in the Copepod *Tigriopus japonicus* and Modulates Expression of Cytochrome P450 (Cyp) Genes. *Aquatic Toxicology*, **152**, 308-317.
- Handley, S. J.; Willis, T. J.; Cole, R. G.; Bradley, A.; Cairney, D. J.; Brown, S. N.; Carter, M. E. (2014) The Importance of Benchmarking Habitat Structure and Composition for Understanding the Extent of Fishing Impacts in Soft Sediment Ecosystems. *Journal of Sea Research*, **86**, 58-68.
- Handoh, I. C.; Kawai, T. (2014) Modelling Exposure of Oceanic Higher Trophic-Level Consumers to Polychlorinated Biphenyls: Pollution 'Hotspots' in Relation to Mass Mortality Events of Marine Mammals. *Marine Pollution Bulletin*, **85**, 824-830.
- Haney, J. C.; Geiger, H. J.; Short, J. W. (2014a) Bird Mortality from the Deepwater Horizon Oil Spill. I. Exposure Probability in the Offshore Gulf of Mexico. *Marine Ecology Progress Series*, **513**, 225-237.
- Haney, J. C.; Geiger, H. J.; Short, J. W. (2014b) Bird

- Mortality from the Deepwater Horizon Oil Spill. ii. Carcass Sampling and Exposure Probability in the Coastal Gulf of Mexico. *Marine Ecology Progress Series*, **513**, 239-252.
- Hansen, B. H.; Altin, D.; Bonaunet, K.; Overjordet, I. B. (2014) Acute Toxicity of Eight Oil Spill Response Chemicals to Temperate, Boreal, and Arctic Species. *Journal of Toxicology and Environmental Health-Part a-Current Issues*, **77**, 495-505.
- Hao, Q.; Sun, Y.-X.; Xu, X.-R.; Yao, Z.-W.; Wang, Y.-S.; Zhang, Z.-W.; Luo, X.-J.; Mai, B.-X. (2014) Occurrence of Persistent Organic Pollutants in Marine Fish from the Natuna Island, South China Sea. *Marine Pollution Bulletin*, **85**, 274-279.
- Hariharan, G.; Purvaja, R.; Ramesh, R. (2014) Toxic Effects of Lead on Biochemical and Histological Alterations in Green Mussel (*Perna viridis*) Induced by Environmentally Relevant Concentrations. *Journal of Toxicology and Environmental Health-Part a-Current Issues*, **77**, 246-260.
- Harms, C.A.; P. McClellan-Green; M.H. Godfrey; E.F. Christiansen; H.J. Broadhurst and C. Goddard-Codding. 2014. Clinical Pathology Effects of Crude Oil and Dispersant on Hatchling Loggerhead Sea Turtles (*Caretta caretta*). Abstract. 2014 International Association of Aquatic Animal Medicine Conference, May 17-22, 2014, Gold Coast, Australia.
- Harrison, J. P.; Schratzberger, M.; Sapp, M.; Osborn, A. M. (2014) Rapid Bacterial Colonization of Low-Density Polyethylene Microplastics in Coastal Sediment Microcosms. *Bmc Microbiology*, **14**.
- Harvey, H. R.; Taylor, K. A.; Pie, H. V.; Mitchelmore, C. L. (2014) Polycyclic Aromatic and Aliphatic Hydrocarbons in Chukchi Sea Biota and Sediments and Their Toxicological Response in the Arctic Cod, *Boreogadus saida*. *Deep-Sea Research Part II-Topical Studies in Oceanography*, **102**, 32-55.
- Harwell, M. A.; Gentile, J. H. (2014) Assessing Risks to Sea Otters and the Exxon Valdez Oil Spill: New Scenarios, Attributable Risk, and Recovery. [*Human and Ecological Risk Assessment*, **20**, 889-916.](#)
- He, S. Y.; Zhang, X.; Wang, W. C.; Wu, Q. L. (2014) Distribution and Toxicity Risks of PCBs in the Tidal Flat Ecosystem near the 37th PCBs Sealed Site on the Coast of Zhejiang Province, China. [*Human and Ecological Risk Assessment*, **21**, 415-433.](#)
- Henkel, L. A.; Nevins, H.; Martin, M.; Sugarman, S.; Harvey, J. T.; Ziccardi, M. H. (2014) Chronic Oiling of Marine Birds in California by Natural Petroleum Seeps, Shipwrecks, and Other Sources. *Marine Pollution Bulletin*, **79**, 155-163.
- Hernandez-Almaraz, P.; Mendez-Rodriguez, L.; Zenteno-Savin, T.; Garcia-Dominguez, F.; Vazquez-Botello, A.; Serviere-Zaragoza, E. (2014) Metal Mobility and Bioaccumulation Differences at Lower Trophic Levels in Marine Ecosystems

- Dominated by Sargassum Species. *Journal of the Marine Biological Association of the United Kingdom*, **94**, 435-442.
- Ho, K. K. Y.; Leung, K. M. Y. (2014) Organotin Contamination in Seafood and Its Implication for Human Health Risk in Hong Kong. *Marine Pollution Bulletin*, **85**, 634-640.
- Hoang, T. C.; Rand, G. M. (2014) Effects of Contaminated St. Lucie River Saltwater Sediments on an Amphipod (*Ampelisca abdita*) and a Hard-Shell Clam (*Mercenaria mercenaria*). [*Archives of Environmental Contamination and Toxicology*](#), **67**, 224-233.
- Hoarau, L.; Ainley, L.; Jean, C.; Ciccione, S. (2014) Ingestion and Defecation of Marine Debris by Loggerhead Sea Turtles, *Caretta caretta*, from by-Catches in the South-West Indian Ocean. *Marine Pollution Bulletin*, **84**, 90-96.
- Holmes, L. A.; Turner, A.; Thompson, R. C. (2014) Interactions between Trace Metals and Plastic Production Pellets under Estuarine Conditions. *Marine Chemistry*, **167**, 25-32.
- Hong, H.; Li, D.; Shen, R.; Wang, X.; Shi, D. (2014) Mechanisms of Hexabromocyclododecanes Induced Developmental Toxicity in Marine Medaka (*Oryzias melastigma*) Embryos. *Aquatic Toxicology*, **152**, 173-185.
- Hong, S.; Khim, J. S.; Park, J.; Son, H.-S.; Choi, S.-D.; Choi, K.; Ryu, J.; Kim, C.-Y.; Chang, G. S.; Giesy, J. P. (2014a) Species- and Tissue-Specific Bioaccumulation of Arsenicals in Various Aquatic Organisms from a Highly Industrialized Area in the Pohang City, Korea. *Environmental Pollution*, **192**, 27-35.
- Hong, W.-J.; Jia, H.; Liu, C.; Zhang, Z.; Sun, Y.; Li, Y.-F. (2014b) Distribution, Source, Fate and Bioaccumulation of Methyl Siloxanes in Marine Environment. *Environmental Pollution*, **191**, 175-181.
- Hook, S. E.; Osborn, H. L.; Gissi, F.; Moncuquet, P.; Twine, N. A.; Wilkins, M. R.; Adams, M. S. (2014) RNA-SEQ Analysis of the Toxicant-Induced Transcriptome of the Marine Diatom, *Ceratoneis closterium*. *Marine Genomics*, **16**, 45-53.
- Horvat, M.; Degenek, N.; Lipej, L.; Tratnik, J. S.; Faganeli, J. (2014) Trophic Transfer and Accumulation of Mercury in Ray Species in Coastal Waters Affected by Historic Mercury Mining (Gulf of Trieste, Northern Adriatic Sea). *Environmental Science and Pollution Research*, **21**, 4163-4176.
- Howe, P. L.; Reichelt-Brushett, A. J.; Clark, M. W. (2014a) Development of a Chronic, Early Life-Stage Sub-Lethal Toxicity Test and Recovery Assessment for the Tropical Zooxanthellate Sea Anemone *Aiptasia pulchella*. *Ecotoxicology and Environmental Safety*, **100**, 138-147.
- Howe, P. L.; Reichelt-Brushett, A. J.; Clark, M. W. (2014b) Effects of Cd, Co, Cu, Ni and Zn on Asexual Reproduction and Early Development of the Tropical Sea Anemone *Aiptasia pulchella*.

- Ecotoxicology*, **23**, 1593-1606.
- Hu, W.; Culloty, S.; Darmody, G.; Lynch, S.; Davenport, J.; Ramirez-Garcia, S.; Dawson, K. A.; Lynch, I.; Blasco, J.; Sheehan, D. (2014) Toxicity of Copper Oxide Nanoparticles in the Blue Mussel, *Mytilus edulis*: A Redox Proteomic Investigation. *Chemosphere*, **108**, 289-299.
- Huge, D. H.; Schofield, P. J.; Jacoby, C. A.; Frazer, T. K. (2014) Total Mercury Concentrations in Lionfish (*Pterois volitans* Miles) from the Florida Keys National Marine Sanctuary, USA. *Marine Pollution Bulletin*, **78**, 51-55.
- Hughes, D. J. (2014) Benthic Habitat and Megafaunal Zonation across the Hebridean Slope, Western Scotland, Analysed from Archived Seabed Photographs. *Journal of the Marine Biological Association of the United Kingdom*, **94**, 643-658.
- Hwang, H.-M.; Stanton, B.; McBride, T.; Anderson, M. J. (2014) Polycyclic Aromatic Hydrocarbon Body Residues and Lysosomal Membrane Destabilization in Mussels Exposed to the Dubai Star Bunker Fuel Oil (Intermediate Fuel Oil 380) Spill in San Francisco Bay. *Environmental Toxicology and Chemistry*, **33**, 1117-1121.
- Hwang, J.; Suh, S.-S.; Chang, M.; Park, S. Y.; Ryu, T. K.; Lee, S.; Lee, T.-K. (2014) Effects of Triclosan on Reproductive Parameters and Embryonic Development of Sea Urchin, *Strongylocentrotus nudus*. *Ecotoxicology and Environmental Safety*, **100**, 148-152.
- Hwang, S. W.; Lee, H. G.; Choi, K. H.; Kim, C. K.; Lee, T. W. (2014) Impact of Sand Extraction on Fish Assemblages in Gyeonggi Bay, Korea. *Journal of Coastal Research*, **30**, 1251-1259.
- Incardona, J. P.; Gardner, L. D.; Linbo, T. L.; Brown, T. L.; Esbaugh, A. J.; Mager, E. M.; Stieglitz, J. D.; French, B. L.; Labenia, J. S.; Laetz, C. A.; Tagal, M.; Sloan, C. A.; Elizur, A.; Benetti, D. D.; Grosell, M.; Block, B. A.; Scholz, N. L. (2014) Deepwater Horizon Crude Oil Impacts the Developing Hearts of Large Predatory Pelagic Fish. *Proceedings of the National Academy of Sciences of the United States of America*, **111**, E1510-E1518.
- Ingels, J.; Dashfield, S. L.; Somerfield, P. J.; Widdicombe, S.; Austen, M. C. (2014) Interactions between Multiple Large Macrofauna Species and Nematode Communities - Mechanisms for Indirect Impacts of Trawling Disturbance. *Journal of Experimental Marine Biology and Ecology*, **456**, 41-49.
- Ivar do Sul, J. A.; Costa, M. F. (2014) The Present and Future of Microplastic Pollution in the Marine Environment. *Environmental Pollution*, **185**, 352-364.
- Jang, Y. C.; Hong, S.; Lee, J.; Lee, M. J.; Shim, W. J. (2014) Estimation of Lost Tourism Revenue in Geoje Island from the 2011 Marine Debris Pollution Event in South Korea. *Marine Pollution Bulletin*, **81**, 49-54.
- Jebali, J.; Chicano-Galvez, E.; Fernandez-Cisnal, R.; Banni, M.; Chouba, L.; Boussetta, H.; Lopez-

- Barea, J.; Alhama, J. (2014) Proteomic Analysis in Caged Mediterranean Crab (*Carcinus maenas*) and Chemical Contaminant Exposure in Teboulba Harbour, Tunisia. *Ecotoxicology and Environmental Safety*, **100**, 15-26.
- Johansson, C. H.; Janmar, L.; Backhaus, T. (2014) Toxicity of Ciprofloxacin and Sulfamethoxazole to Marine Periphytic Algae and Bacteria. *Aquatic Toxicology*, **156**, 248-258.
- Jones, H. J.; Swadling, K. M.; Butler, E. C. V.; Barry, L. A.; Macleod, C. K. (2014) Application of Stable Isotope Mixing Models for Defining Trophic Biomagnification Pathways of Mercury and Selenium. *Limnology and Oceanography*, **59**, 1181-1192.
- Jorundsdottir, H. O.; Jensen, S.; Hylland, K.; Holth, T. F.; Gunnlaugsdottir, H.; Svavarsson, J.; Olafsdottir, A.; El-Taliawy, H.; Riget, F.; Strand, J.; Nyberg, E.; Bignert, A.; Hoydal, K. S.; Halldorsson, H. P. (2014) Pristine Arctic: Background Mapping of Paks, Pah Metabolites and Inorganic Trace Elements in the North-Atlantic Arctic and Sub-Arctic Coastal Environment. *Science of the Total Environment*, **493**, 719-728.
- Joye, S. B.; Teske, A. P.; Kostka, J. E. (2014) Microbial Dynamics Following the Macondo Oil Well Blowout across Gulf of Mexico Environments. *Bioscience*, **64**, 766-777.
- Judy, C. R.; Graham, S. A.; Lin, Q.; Hou, A.; Mendelsohn, I. A. (2014) Impacts of Macondo Oil from Deepwater Horizon Spill on the Growth Response of the Common Reed *Phragmites australis*: A Mesocosm Study. *Marine Pollution Bulletin*, **79**, 69-76.
- Junoy, J.; Castellanos, C.; Bernardo-Madrid, R.; Riera, R.; Vieitez, J. M. (2014) Macroinfaunal Recovery on the Beach Most Severely Affected by the 'Prestige' Oil Spill (O Rostro, Galicia, North-West Spain). *Journal of the Marine Biological Association of the United Kingdom*, **94**, 17-24.
- Jurelevicius, D.; de Almeida Couto, C. R.; Alvarez, V. M.; Vollu, R. E.; Dias, F. d. A.; Seldin, L. (2014) Response of the Archaeal Community to Simulated Petroleum Hydrocarbon Contamination in Marine and Hypersaline Ecosystems. *Water Air and Soil Pollution*, **225**.
- Kalantzi, I.; Papageorgiou, N.; Sevastou, K.; Black, K. D.; Pergantis, S. A.; Karakassis, I. (2014) Metals in Benthic Macrofauna and Biogeochemical Factors Affecting Their Trophic Transfer to Wild Fish around Fish Farm Cages. *Science of the Total Environment*, **470**, 742-753.
- Kang, T.; Min, W.-G.; Rho, H. S.; Park, H.-S.; Kim, D. (2014) Differential Responses of a Benthic Meiofaunal Community to an Artificial Oil Spill in the Intertidal Zone. *Journal of the Marine Biological Association of the United Kingdom*, **94**, 219-231.
- Kapsimalis, V.; Panagiotopoulos, I. P.; Talagani, P.; Hatzianestis, I.; Kaberi, H.; Rousakis, G.; Kanellopoulos, T. D.; Hatiris, G. A. (2014)

- Organic Contamination of Surface Sediments in the Metropolitan Coastal Zone of Athens, Greece: Sources, Degree, and Ecological Risk. *Marine Pollution Bulletin*, **80**, 312-324.
- Kenchington, E.; Javier Murillo, F.; Lirette, C.; Sacau, M.; Koen-Alonso, M.; Kenny, A.; Ollerhead, N.; Wareham, V.; Beazley, L. (2014) Kernel Density Surface Modelling as a Means to Identify Significant Concentrations of Vulnerable Marine Ecosystem Indicators. *Plos One*, **9**.
- Kim, B.-M.; Rhee, J.-S.; Jeong, C.-B.; Seo, J. S.; Park, G. S.; Lee, Y.-M.; Lee, J.-S. (2014) Heavy Metals Induce Oxidative Stress and Trigger Oxidative Stress-Mediated Heat Shock Protein (Hsp) Modulation in the Intertidal Copepod *Tigriopus japonicus*. *Comparative Biochemistry and Physiology C-Toxicology & Pharmacology*, **166**, 65-74.
- Kim, C. J.; Hong, G. H.; Kim, H. E.; Yang, D. B. (2014) Polycyclic Aromatic Hydrocarbons (PAHs) in Starfish Body and Bottom Sediments in Mohang Harbor (Taeon), South Korea. *Environmental Monitoring and Assessment*, **186**, 4343-4356.
- Kirby, M. F.; Gioia, R.; Law, R. J. (2014) The Principles of Effective Post-Spill Environmental Monitoring in Marine Environments and Their Application to Preparedness Assessment. *Marine Pollution Bulletin*, **82**, 11-18.
- Klemas, V.; Blazauskas, N. (2014) Reducing the Oil Spill Threat to the Marine Environment (Foreword). *Baltica*, **27**, 1-2.
- Ko, F.-C.; Chang, C.-W.; Cheng, J.-O. (2014) Comparative Study of Polycyclic Aromatic Hydrocarbons in Coral Tissues and the Ambient Sediments from Kenting National Park, Taiwan. *Environmental Pollution*, **185**, 35-43.
- Ko, F. C.; We, N.-Y.; Chou, L.-S. (2014) Bioaccumulation of Persistent Organic Pollutants in Stranded Cetaceans from Taiwan Coastal Waters. *Journal of Hazardous Materials*, **277**, 127-133.
- Koelmans, A. A.; Besseling, E.; Foekema, E. M. (2014) Leaching of Plastic Additives to Marine Organisms. *Environmental Pollution*, **187**, 49-54.
- Konovalenko, L.; Bradshaw, C.; Kumblad, L.; Kautsky, U. (2014) Radionuclide Transfer in Marine Coastal Ecosystems, a Modelling Study Using Metabolic Processes and Site Data. *Journal of Environmental Radioactivity*, **133**, 48-59.
- Kostka, J. E.; Teske, A. P.; Joye, S. B.; Head, I. M. (2014) The Metabolic Pathways and Environmental Controls of Hydrocarbon Biodegradation in Marine Ecosystems. *Frontiers in Microbiology*, **5**.
- Koyama, J.; Imakado, C.; Uno, S.; Kuroda, T.; Hara, S.; Majima, T.; Shiota, H.; Anasco, N. C. (2014) Simulated Distribution and Ecotoxicity-Based Assessment of Chemically-Dispersed Oil in Tokyo Bay. *Marine Pollution Bulletin*, **85**, 487-493.
- Kumar, M. K. P.; Shyama, S. K.; Sonaye, B. S.; Naik, U. R.; Kadam, S. B.; Bipin, P. D.; D'Costa, A.; Chaubey, R. C. (2014) Evaluation of Gamma-

- Radiation-Induced DNA Damage in Two Species of Bivalves and Their Relative Sensitivity Using Comet Assay. *Aquatic Toxicology*, **150**, 1-8.
- Kwok, K. W. H.; Batley, G. E.; Wenning, R. J.; Zhu, L.; Vangheluwe, M.; Lee, S. (2014) Sediment Quality Guidelines: Challenges and Opportunities for Improving Sediment Management. *Environmental Science and Pollution Research*, **21**, 17-27.
- Kwon, B. G.; Saïdo, K.; Koizumi, K.; Sato, H.; Ogawa, N.; Chung, S. Y.; Kusui, T.; Kodera, Y.; Kogure, K. (2014) Regional Distribution of Styrene Analogues Generated from Polystyrene Degradation Along the Coastlines of the North-East Pacific Ocean and Hawaii. *Environmental Pollution*, **188**, 45-49.
- Lacroix, C.; Coquille, V.; Guyomarch, J.; Auffret, M.; Moraga, D. (2014) A Selection of Reference Genes and Early-Warning Mrna Biomarkers for Environmental Monitoring Using *Mytilus* Spp. As Sentinel Species. *Marine Pollution Bulletin*, **86**, 304-313.
- Lamendella, R.; Strutt, S.; Borglin, S.; Chakraborty, R.; Tas, N.; Mason, O. U.; Hultman, J.; Prestat, E.; Hazen, T. C.; Jansson, J. K. (2014) Assessments of the Deepwater Horizon Oil Spill Impact on Gulf Coast Microbial Communities. *Frontiers in Microbiology*, **5**.
- Lange, L.; Griffiths, C. L. (2014) Large-Scale Spatial Patterns within Soft-Bottom Epibenthic Invertebrate Assemblages Along the West Coast of South Africa, Based on the Nansen Trawl Survey. *African Journal of Marine Science*, **36**, 111-124.
- Laramore, S.; Krebs, W.; Garr, A. (2014) Effects of Macondo Canyon 252 Oil (Naturally and Chemically Dispersed) on Larval *Crassostrea virginica* (Gmelin, 1791). *Journal of Shellfish Research*, **33**, 709-718.
- Lavender Law, K. L.; Thompson, R. C. (2014) Microplastics in the Seas. *Science*, **345**, 144-145.
- Lavers, J. L.; Bond, A. L.; Hutton, I. (2014) Plastic Ingestion by Flesh-Footed Shearwaters (*Puffinus carneipes*): Implications for Fledgling Body Condition and the Accumulation of Plastic-Derived Chemicals. *Environmental Pollution*, **187**, 124-129.
- Law, R. J. (2014) An Overview of Time Trends in Organic Contaminant Concentrations in Marine Mammals: Going up or Down? *Marine Pollution Bulletin*, **82**, 7-10.
- Le Bihanic, F.; Perrichon, P.; Landi, L.; Clerandau, C.; Le Menach, K.; Budzinski, H.; Cousin, X.; Cachot, J. (2014) Development of a Reference Artificial Sediment for Chemical Testing Adapted to the Mela Sediment Contact Assay. *Environmental Science and Pollution Research*, **21**, 13689-13702.
- Lee, H.; Shim, W. J.; Kwon, J.-H. (2014) Sorption Capacity of Plastic Debris for Hydrophobic Organic Chemicals. *Science of the Total Environment*, **470**, 1545-1552.

- Lee, J., Hong, S., Jang, Y.C., Lee, M.J., Kang, D., Shim, W.J. (2014a) Finding solutions for the Styrofoam buoy debris problem through participatory workshops. *Marine Policy* **51**: 182-189.
- Lee, M.-A.; Guo, R.; Ki, J.-S. (2014) Different Transcriptional Responses of Heat Shock Protein 20 in the Marine Diatom *Ditylum brightwellii* Exposed to Metals and Endocrine-Disrupting Chemicals. *Environmental Toxicology*, **29**, 1379-1389.
- Lee, Y. G.; Kim, S.; Jeong, D. U.; Lee, J. S.; Woo, H. J.; Park, M. W.; Kim, B. H.; Son, M. H.; Choi, Y. H. (2014) Decalcification of Benthic Foraminifera Due to "Hebei Spirit" Oil Spill, Korea. *Marine Pollution Bulletin*, **87**, 276-285.
- Lehnert, K.; Mueller, S.; Weirup, L.; Ronnenberg, K.; Pawliczka, I.; Rosenberger, T.; Siebert, U. (2014) Molecular Biomarkers in Grey Seals (*Halichoerus grypus*) to Evaluate Pollutant Exposure, Health and Immune Status. *Marine Pollution Bulletin*, **88**, 311-318.
- Lehtonen, K. K.; Sundelin, B.; Lang, T.; Strand, J. (2014) Development of Tools for Integrated Monitoring and Assessment of Hazardous Substances and Their Biological Effects in the Baltic Sea. *Ambio*, **43**, 69-81.
- Leite, D. S.; Sandrini-Neto, L.; Camargo, M. Z.; Thomas, M. C.; Lana, P. C. (2014) Are Changes in the Structure of Nematode Assemblages Reliable Indicators of Moderate Petroleum Contamination? *Marine Pollution Bulletin*, **83**, 38-47.
- Leon, V. M.; Garcia, I.; Martinez-Gomez, C.; Campillo, J. A.; Benedicto, J. (2014) Heterogeneous Distribution of Polycyclic Aromatic Hydrocarbons in Surface Sediments and Red Mullet Along the Spanish Mediterranean Coast. *Marine Pollution Bulletin*, **87**, 352-363.
- Letinski, D.; Parkerton, T.; Redman, A.; Manning, R.; Bragin, G.; Febbo, E.; Palandro, D.; Nedwed, T. (2014) Use of Passive Samplers for Improving Oil Toxicity and Spill Effects Assessment. *Marine Pollution Bulletin*, **86**, 274-282.
- Leung, P. T. Y.; Park, T. J.; Wang, Y.; Che, C. M.; Leung, K. M. Y. (2014) Isoform-Specific Responses of Metallothioneins in a Marine Pollution Biomonitor, the Green-Lipped Mussel *Perna viridis*, Towards Different Stress Stimulations. *Proteomics*, **14**, 1796-1807.
- Li, A. J.; Leung, P. T. Y.; Bao, V. W. W.; Yi, A. X. L.; Leung, K. M. Y. (2014) Temperature-Dependent Toxicities of Four Common Chemical Pollutants to the Marine Medaka Fish, Copepod and Rotifer. *Ecotoxicology*, **23**, 1564-1573.
- Li, H.; Lu, L.; Huang, W.; Yang, J.; Ran, Y. (2014) In-Situ Partitioning and Bioconcentration of Polycyclic Aromatic Hydrocarbons among Water, Suspended Particulate Matter, and Fish in the Dongjiang and Pearl Rivers and the Pearl River Estuary, China. *Marine Pollution Bulletin*, **83**, 306-316.
- Lindgren, J. F.; Hasselov, I.-M.; Dahllöf, I. (2014) Pah

- Effects on Meio- and Microbial Benthic Communities Strongly Depend on Bioavailability. *Aquatic Toxicology*, **146**, 230-238.
- Liu, C.; Chang, V. W. C.; Gin, K. Y. H. (2014) Oxidative Toxicity of Perfluorinated Chemicals in Green Mussel and Bioaccumulation Factor Dependent Quantitative Structure-Activity Relationship. *Environmental Toxicology and Chemistry*, **33**, 2323-2332.
- Liu, C.; Chang, V. W. C.; Gin, K. Y. H.; Viet Tung, N. (2014) Genotoxicity of Perfluorinated Chemicals (PFCs) to the Green Mussel (*Perna viridis*). *Science of the Total Environment*, **487**, 117-122.
- Liu, C.; Gin, K. Y. H.; Chang, V. W. C. (2014) Multi-Biomarker Responses in Green Mussels Exposed to PFCs: Effects at Molecular, Cellular, and Physiological Levels. *Environmental Science and Pollution Research*, **21**, 2785-2794.
- Liu, H.-H.; Xiang, L.-X.; Shao, J.-Z. (2014) A Novel ClqDC Domain-Containing (ClqDC) Protein from *Mytilus coruscus* with the Transcriptional Analysis against Marine Pathogens and Heavy Metals. *Developmental and Comparative Immunology*, **44**, 70-75.
- Liu, J.; Zhang, X. (2014) Comparative Toxicity of New Halophenolic DBPs in Chlorinated Saline Wastewater Effluents against a Marine Alga: Halophenolic DBPs Are Generally More Toxic Than Haloaliphatic Ones. *Water Research*, **65**, 64-72.
- Liu, L.; Li, B.; Lin, K.; Cai, W.; Wang, Q. (2014) Assessing Benthic Ecological Status in Coastal Area near Changjiang River Estuary Using AMBI and M-AMBI. *Chinese Journal of Oceanology and Limnology*, **32**, 290-305.
- Liu, N.; Pan, L.; Gong, X.; Tao, Y.; Hu, Y.; Miao, J. (2014) Effects of Benzo(a)Pyrene on Differentially Expressed Genes and Haemocyte Parameters of the Clam *Venerupis philippinarum*. *Ecotoxicology*, **23**, 122-132.
- Lively, J. A. A.; McKenzie, J. (2014) Toxicity of the Dispersant Corexit 9500 to Early Life Stages of Blue Crab, *Callinectes Sapidus*. *Bulletin of Environmental Contamination and Toxicology*, **93**, 649-653.
- Llorca, M.; Farre, M.; Karapanagioti, H. K.; Barcelo, D. (2014) Levels and Fate of Perfluoroalkyl Substances in Beached Plastic Pellets and Sediments Collected from Greece. *Marine Pollution Bulletin*, **87**, 286-291.
- Lopes-dos-Santos, R. M. A.; Galante-Oliveira, S.; Lopes, E.; Almeida, C.; Barroso, C. (2014) Assessment of Imposéx and Butyltin Concentrations in *Gemphos viverratus* (Kiener, 1834), from Sao Vicente, Republic of Cabo Verde (Africa). *Environmental Science and Pollution Research*, **21**, 10671-10677.
- Lozano, P.; Trombini, C.; Crespo, E.; Blasco, J.; Moreno-Garrido, I. (2014) Roi-Scavenging Enzyme Activities as Toxicity Biomarkers in Three Species of Marine Microalgae Exposed to Model

- Contaminants (Copper, Irgarol and Atrazine). *Ecotoxicology and Environmental Safety*, **104**, 294-301.
- Lucia, M.; Bocher, P.; Chambosse, M.; Delaporte, P.; Bustamante, P. (2014) Trace Element Accumulation in Relation to Trophic Niches of Shorebirds Using Intertidal Mudflats. *Journal of Sea Research*, **92**, 134-143.
- Lyons, K.; Lavado, R.; Schlenk, D.; Lowe, C. G. (2014) Bioaccumulation of Organochlorine Contaminants and Ethoxyresorufin-O-Deethylase Activity in Southern California Round Stingrays (*Urobatis halleri*) Exposed to Planar Aromatic Compounds. *Environmental Toxicology and Chemistry*, **33**, 1380-1390.
- Ma, X.; Chen, C.; Zhang, H.; Gao, Y.; Wang, Z.; Yao, Z.; Chen, J.; Chen, J. (2014a) Congener-Specific Distribution and Bioaccumulation of Short-Chain Chlorinated Paraffins in Sediments and Bivalves of the Bohai Sea, China. *Marine Pollution Bulletin*, **79**, 299-304.
- Ma, X.; Zhang, H.; Wang, Z.; Yao, Z.; Chen, J.; Chen, J. (2014b) Bioaccumulation and Trophic Transfer of Short Chain Chlorinated Paraffins in a Marine Food Web from Liaodong Bay, North China. *Environmental Science & Technology*, **48**, 5964-5971.
- MacDonald, I. R.; Kammen, D. M.; Fan, M. (2014) Science in the Aftermath: Investigations of the DWH Hydrocarbon Discharge. *Environmental Research Letters*, **9**.
- Macias-Zamora, J. V.; Melendez-Sanchez, A. L.; Ramirez-Alvarez, N.; Gutierrez-Galindo, E. A.; Orozco-Borbon, M. V. (2014) On the Effects of the Dispersant Corexit 9500((C)) During the Degradation Process of N-Alkanes and PAHs in Marine Sediments. *Environmental Monitoring and Assessment*, **186**, 1051-1061.
- Madeira, D.; Mendonca, V.; Dias, M.; Roma, J.; Costa, P. M.; Diniz, M. S.; Vinagre, C. (2014) Physiological and Biochemical Thermal Stress Response of the Intertidal Rock Goby *Gobius Paganellus*. *Ecological Indicators*, **46**, 232-239.
- Madeira, D.; Vinagre, C.; Costa, P. M.; Diniz, M. S. (2014) Histopathological Alterations, Physiological Limits, and Molecular Changes of Juvenile *Sparus aurata* in Response to Thermal Stress. *Marine Ecology Progress Series*, **505**, 253-266.
- Madeira Di Benedetto, A. P.; Arruda Ramos, R. M. (2014) Marine Debris Ingestion by Coastal Dolphins: What Drives Differences between Sympatric Species? *Marine Pollution Bulletin*, **83**, 298-301.
- Mager, E. M.; Esbaugh, A. J.; Stieglitz, J. D.; Hoenig, R.; Bodinier, C.; Incardona, J. P.; Scholz, N. L.; Benetti, D. D.; Grosell, M. (2014) Acute Embryonic or Juvenile Exposure to Deepwater Horizon Crude Oil Impairs the Swimming Performance of Mahi-Mahi (*Coryphaena hippurus*). *Environmental Science & Technology*, **48**, 7053-7061.
- Majer, A. P.; Varella Petti, M. A.; Corbisier, T. N.;

- Ribeiro, A. P.; Sawamura Theophilo, C. Y.; de Lima Ferreira, P. A.; Lopes Figueira, R. C. (2014) Bioaccumulation of Potentially Toxic Trace Elements in Benthic Organisms of Admiralty Bay (King George Island, Antarctica). *Marine Pollution Bulletin*, **79**, 321-325.
- Malea, P.; Adamakis, I.-D. S.; Kevrekidis, T. (2014) Effects of Lead Uptake on Microtubule Cytoskeleton Organization and Cell Viability in the Seagrass *Cymodocea nodosa*. *Ecotoxicology and Environmental Safety*, **104**, 175-181.
- Malea, P.; Kevrekidis, T. (2014) Trace Element Patterns in Marine Macroalgae. *Science of the Total Environment*, **494**, 144-157.
- Mangano, M. C.; Kaiser, M. J.; Porporato, E. M. D.; Lambert, G. I.; Rinelli, P.; Spano, N. (2014) Infaunal Community Responses to a Gradient of Trawling Disturbance and a Long-Term Fishery Exclusion Zone in the Southern Tyrrhenian Sea. *Continental Shelf Research*, **76**, 25-35.
- Maranho, L. A.; Baena-Nogueras, R. M.; Lara-Martin, P. A.; DelValls, T. A.; Martin-Diaz, M. L. (2014) Bioavailability, Oxidative Stress, Neurotoxicity and Genotoxicity of Pharmaceuticals Bound to Marine Sediments. The Use of the Polychaete *Hediste diversicolor* as Bioindicator Species. *Environmental Research*, **134**, 353-365.
- Marsico, E. T.; Ferreira, M. S.; Sao Clemente, S. C.; Gouvea, R. C. S.; Jesus, E. F. O.; Conti, C. C.; Conte Junior, C. A.; Kelecom, A. G. A. C. (2014) Distribution of Po-210 in Two Species of Predatory Marine Fish from the Brazilian Coast. *Journal of Environmental Radioactivity*, **128**, 91-96.
- Marsili, L.; Maltese, S.; Coppola, D.; Carletti, L.; Mazzariol, S.; Fossi, M. C. (2014) Ecotoxicological Status of Seven Sperm Whales (*Physeter macrocephalus*) Stranded Along the Adriatic Coast of Southern Italy. *Aquatic Conservation-Marine and Freshwater Ecosystems*, **24**, 103-118.
- Martin, J.; Edwards, H. H.; Bled, F.; Fønnesbeck, C. J.; Dupuis, J. A.; Gardner, B.; Koslovsky, S. M.; Aven, A. M.; Ward-Geiger, L. I.; Carmichael, R. H.; Fagan, D. E.; Ross, M. A.; Reinert, T. R. (2014) Estimating Upper Bounds for Occupancy and Number of Manatees in Areas Potentially Affected by Oil from the Deepwater Horizon Oil Spill. *Plos One*, **9**.
- Martins, C. C.; Cabral, A. C.; Barbosa-Cintra, S. C. T.; Dauner, A. L. L.; Souza, F. M. (2014) An Integrated Evaluation of Molecular Marker Indices and Linear Alkylbenzenes (Labs) to Measure Sewage Input in a Subtropical Estuary (Babitonga Bay, Brazil). *Environmental Pollution*, **188**, 71-80.
- Maszkowska, J.; Stolte, S.; Kumirska, J.; Lukaszewicz, P.; Mioduszewska, K.; Puckowski, A.; Caban, M.; Wagil, M.; Stepnowski, P.; Bialk-Bielinska, A. (2014) Beta-Blockers in the Environment: Part II. Ecotoxicity Study. *Science of the Total Environment*, **493**, 1122-1126.

- McEneff, G.; Barron, L.; Kelleher, B.; Paull, B.; Quinn, B. (2014) A Year-Long Study of the Spatial Occurrence and Relative Distribution of Pharmaceutical Residues in Sewage Effluent, Receiving Marine Waters and Marine Bivalves. *Science of the Total Environment*, **476**, 317-326.
- Meador, J. P. (2014) Do Chemically Contaminated River Estuaries in Puget Sound (Washington, USA) Affect the Survival Rate of Hatchery-Reared Chinook Salmon? *Canadian Journal of Fisheries and Aquatic Sciences*, **71**, 162-180.
- Méndez-Fernandez, P.; Webster, L.; Chouvelon, T.; Bustamante, P.; Ferreira, M.; Gonzalez, A. F.; Lopez, A.; Moffat, C. F.; Pierce, G. J.; Read, F. L.; Russell, M.; Santos, M. B.; Spitz, J.; Vingada, J. V.; Caurant, F. (2014a) An Assessment of Contaminant Concentrations in Toothed Whale Species of the NW Iberian Peninsula: Part I. Persistent Organic Pollutants. *Science of the Total Environment*, **484**, 196-205.
- Méndez-Fernandez, P.; Webster, L.; Chouvelon, T.; Bustamante, P.; Ferreira, M.; Gonzalez, A. F.; Lopez, A.; Moffat, C. F.; Pierce, G. J.; Read, F. L.; Russell, M.; Santos, M. B.; Spitz, J.; Vingada, J. V.; Caurant, F. (2014b) An Assessment of Contaminant Concentrations in Toothed Whale Species of the NW Iberian Peninsula: Part II. Trace Element Concentrations. *Science of the Total Environment*, **484**, 206-217.
- Meseck, S. L.; Mercaldo-Allen, R.; Rose, J. M.; Clark, P.; Kuropat, C.; Pereira, J. J.; Goldberg, R. (2014) Effects of Hydraulic Dredging for *Mercenaria mercenaria*, Northern Quahog, on Sediment Biogeochemistry. *Journal of the World Aquaculture Society*, **45**, 301-311.
- Mestre, N. C.; Calado, R.; Soares, A. M. V. M. (2014) Exploitation of Deep-Sea Resources: The Urgent Need to Understand the Role of High Pressure in the Toxicity of Chemical Pollutants to Deep-Sea Organisms. *Environmental Pollution*, **185**, 369-371.
- Michel, J.; Rutherford, N. (2014) Impacts, Recovery Rates, and Treatment Options for Spilled Oil in Marshes. *Marine Pollution Bulletin*, **82**, 19-25.
- Minetto, D.; Libralato, G.; Ghirardini, A. V. (2014) Ecotoxicity of Engineered TiO₂ Nanoparticles to Saltwater Organisms: An Overview. *Environment International*, **66**, 18-27.
- Minguez, L.; Di Poi, C.; Farcy, E.; Ballandonne, C.; Benchouala, A.; Bojic, C.; Cossu-Leguille, C.; Costil, K.; Serpentine, A.; Lebel, J.-M.; Halm-Lemeille, M.-P. (2014) Comparison of the Sensitivity of Seven Marine and Freshwater Bioassays as Regards Antidepressant Toxicity Assessment. *Ecotoxicology*, **23**, 1744-1754.
- Mirto, S.; Arigo, C.; Genovese, L.; Pusceddu, A.; Gambi, C.; Danovaro, R. (2014) Nematode Assemblage Response to Fish-Farm Impact in Vegetated (*Posidonia oceanica*) and Non-Vegetated Habitats. *Aquaculture Environment Interactions*, **5**, 17-28.
- Morandin, L. A.; O'Hara, P. D. (2014) Fish Oil Disrupts

- Seabird Feather Microstructure and Waterproofing. *Science of the Total Environment*, **496**, 257-263.
- Morris, C. A.; Owen, J. R.; Thomas, M. C.; El-Hiti, G. A.; Harwood, J. L.; Kille, P. (2014) Intracellular Localization and Induction of a Dynamic RNA-Editing Event of Macro-Algal V-ATPase Subunit a (VHA-a) in Response to Copper. *Plant Cell and Environment*, **37**, 189-203.
- Mouneyrac, C.; Buffet, P.-E.; Poirier, L.; Zalouk-Vergnoux, A.; Guibolini, M.; Risso-de Faverney, C.; Gilliland, D.; Berhanu, D.; Dybowska, A.; Chatel, A.; Perrein-Ettajni, H.; Pan, J.-F.; Thomas-Guyon, H.; Reip, P.; Valsami-Jones, E. (2014) Fate and Effects of Metal-Based Nanoparticles in Two Marine Invertebrates, the Bivalve Mollusc *scrobicularia Plana* and the Annelid Polychaete *Hediste diversicolor*. *Environmental Science and Pollution Research*, **21**, 7899-7912.
- Mu, J.; Jin, F.; Ma, X.; Lin, Z.; Wang, J. (2014) Comparative Effects of Biological and Chemical Dispersants on the Bioavailability and Toxicity of Crude Oil to Early Life Stages of Marine Medaka (*Oryzias melastigma*). *Environmental Toxicology and Chemistry*, **33**, 2576-2583.
- Mu, J.; Wang, J.; Jin, F.; Wang, X.; Hong, H. (2014) Comparative Embryotoxicity of Phenanthrene and Alkyl-Phenanthrene to Marine Medaka (*Oryzias melastigma*). *Marine Pollution Bulletin*, **85**, 505-515.
- Munari, M.; Mann, M. G.; Matozzo, V. (2014) Effects of the Antidepressant Fluoxetine on the Immune Parameters and Acetylcholinesterase Activity of the Clam *Venerupis philippinarum*. *Marine Environmental Research*, **94**, 32-37.
- Muntadas, A.; Demestre, M.; de Juan, S.; Frid, C. L. J. (2014) Trawling Disturbance on Benthic Ecosystems and Consequences on Commercial Species: A Northwestern Mediterranean Case Study. *Scientia Marina*, **78**, 53-65.
- Murawski, S. A.; Hogarth, W. T.; Peebles, E. B.; Barbeiri, L. (2014) Prevalence of External Skin Lesions and Polycyclic Aromatic Hydrocarbon Concentrations in Gulf of Mexico Fishes, Post-Deepwater Horizon. *Transactions of the American Fisheries Society*, **143**, 1084-1097.
- Nabe-Nielsen, J.; Sibly, R. M.; Tougaard, J.; Teilmann, J.; Sveegaard, S. (2014) Effects of Noise and By-Catch on a Danish Harbour Porpoise Population. *Ecological Modelling*, **272**, 242-251.
- National Oceanic and Atmospheric Administration, Marine Debris Program. (2014a) Report on the entanglement of marine species in marine debris with an emphasis on species in the United States. Silver Spring, MD. 33p.
- National Oceanic and Atmospheric Administration, Marine Debris Program. (2014b) Report on the occurrence and health effects of anthropogenic debris ingested by marine organisms. Silver Spring, MD. 24p.
- Nel, P.; Branch, G. M. (2014) The Effect of Bait Collectors

- on Stocks of *Callichirus kraussi* and *Upogebia* Species in Langebaan Lagoon. *African Zoology*, **49**, 30-43.
- Neves, B. M.; Du Preez, C.; Edinger, E. (2014) Mapping Coral and Sponge Habitats on a Shelf-Depth Environment Using Multibeam Sonar and Rov Video Observations: Learmonth Bank, Northern British Columbia, Canada. *Deep-Sea Research Part II-Topical Studies in Oceanography*, **99**, 169-183.
- Nilsen, F.; Hyrenbach, K. D.; Fang, J.; Jensen, B. (2014) Use of Indicator Chemicals to Characterize the Plastic Fragments Ingested by Laysan Albatross. *Marine Pollution Bulletin*, **87**, 230-236.
- Niquil, N.; Baeta, A.; Marques, J. C.; Chaalali, A.; Lobry, J.; Patricio, J. (2014) Reaction of an Estuarine Food Web to Disturbance: Lindeman's Perspective. *Marine Ecology Progress Series*, **512**, 141-154.
- Noedler, K.; Voutsas, D.; Licha, T. (2014) Polar Organic Micropollutants in the Coastal Environment of Different Marine Systems. *Marine Pollution Bulletin*, **85**, 50-59.
- Nurulnadia, M. Y.; Koyama, J.; Uno, S.; Kito, A.; Kokushi, E.; Bacolod, E. T.; Ito, K.; Chuman, Y. (2014) Accumulation of Endocrine Disrupting Chemicals (EDCs) in the Polychaete *Paraprionospio* Sp from the Yodo River Mouth, Osaka Bay, Japan. *Environmental Monitoring and Assessment*, **186**, 1453-1463.
- Oberbeckmann, S.; Loeder, M. G. J.; Gerds, G.; Osborn, A. M. (2014) Spatial and Seasonal Variation in Diversity and Structure of Microbial Biofilms on Marine Plastics in Northern European Waters. *Fems Microbiology Ecology*, **90**, 478-492.
- Oliveira, D. D.; Souza-Santos, L. P.; Silva, H. K. P.; Macedo, S. J. (2014) Toxicity of Sediments from a Mangrove Forest Patch in an Urban Area in Pernambuco (Brazil). *Ecotoxicology and Environmental Safety*, **104**, 373-378.
- Özhan, K.; Bargu, S. (2014) Distinct Responses of Gulf of Mexico Phytoplankton Communities to Crude Oil and the Dispersant Corexit EC9500a under Different Nutrient Regimes. *Ecotoxicology*, **23**, 370-384.
- Özhan, K.; Miles, S. M.; Gao, H.; Bargu, S. (2014) Relative Phytoplankton Growth Responses to Physically and Chemically Dispersed South Louisiana Sweet Crude Oil. *Environmental Monitoring and Assessment*, **186**, 3941-3956.
- Özhan, K.; Parsons, M. L.; Bargu, S. (2014) How Were Phytoplankton Affected by the Deepwater Horizon Oil Spill? *Bioscience*, **64**, 829-836.
- Paredes, E.; Perez, S.; Rodil, R.; Quintana, J. B.; Beiras, R. (2014) Ecotoxicological Evaluation of Four UV Filters Using Marine Organisms from Different Trophic Levels *Isochrysis galbana*, *Mytilus galloprovincialis*, *Paracentrotus lividus*, and *Siriella armata*. *Chemosphere*, **104**, 44-50.
- Park, J.; Kim, S.; Yoo, J.; Lee, J.-S.; Park, J.-W.; Jung, J. (2014) Effect of Salinity on Acute Copper and Zinc Toxicity to *Tigriopus japonicus*: The

- Difference between Metal Ions and Nanoparticles. *Marine Pollution Bulletin*, **85**, 526-531.
- Paruk, J. D.; Long, D.; Perkins, C.; East, A.; Sigel, B. J.; Evers, D. C. (2014) Polycyclic Aromatic Hydrocarbons Detected in Common Loons (*Gavia immer*) Wintering Off Coastal Louisiana. *Waterbirds*, **37**, 85-93.
- Pastore, A. S.; Santacroce, M. P.; Narracci, M.; Cavallo, R. A.; Acquaviva, M. I.; Casalino, E.; Colamonaco, M.; Crescenzo, G. (2014) Genotoxic Damage of Benzo a Pyrene in Cultured Sea Bream (*Sparus Aurata* L.) Hepatocytes: Harmful Effects of Chronic Exposure. *Marine Environmental Research*, **100**, 74-85.
- Pavlidou, A.; Papadopoulos, V. P.; Hatzianestis, I.; Simboura, N.; Patiris, D.; Tsabaris, C. (2014) Chemical Inputs from a Karstic Submarine Groundwater Discharge (SGD) into an Oligotrophic Mediterranean Coastal Area. *Science of the Total Environment*, **488**, 1-13.
- Pawlowski, J.; Esling, P.; Lejzerowicz, F.; Cedhagen, T.; Wilding, T. A. (2014) Environmental Monitoring through Protist Next-Generation Sequencing Metabarcoding: Assessing the Impact of Fish Farming on Benthic Foraminifera Communities. *Molecular Ecology Resources*, **14**, 1129-1140.
- Payne, S. J.; King, C. K.; Zamora, L. M.; Virtue, P. (2014) Temporal Changes in the Sensitivity of Coastal Antarctic Zooplankton Communities to Diesel Fuel: A Comparison between Single- and Multi-Species Toxicity Tests. *Environmental Toxicology and Chemistry*, **33**, 882-890.
- Pearce, B.; Farinas-Franco, J. M.; Wilson, C.; Pitts, J.; deBurgh, A.; Somerfield, P. J. (2014) Repeated Mapping of Reefs Constructed by *Sabellaria spinulosa* Leuckart 1849 at an Offshore Wind Farm Site. *Continental Shelf Research*, **83**, 3-13.
- Peng, H.; Wan, Y.; Zhang, K.; Sun, J.; Hu, J. (2014) Trophic Transfer of Dechloranes in the Marine Food Web of Liaodong Bay, North China. *Environmental Science & Technology*, **48**, 5458-5466.
- Pennings, S. C.; McCall, B. D.; Hooper-Bui, L. (2014) Effects of Oil Spills on Terrestrial Arthropods in Coastal Wetlands. *Bioscience*, **64**, 789-795.
- Peters, A. J.; Siuda, A. N. S. (2014) A Review of Observations of Floating Tar in the Sargasso Sea. *Oceanography*, **27**, 217-221.
- Pham, P. H.; Huang, Y. J.; Chen, C.; Bols, N. C. (2014) Corexit 9500 Inactivates Two Enveloped Viruses of Aquatic Animals but Enhances the Infectivity of a Nonenveloped Fish Virus. *Applied and Environmental Microbiology*, **80**, 1035-1041.
- Podolska, M.; Nadolna, K.; Szostakowska, B. (2014) Acetylcholinesterase Activity in the Host-Parasite System of the Cod *Gadus morhua* and Acanthocephalan *Echinorhynchus gadi* from the Southern Baltic Sea. *Marine Pollution Bulletin*, **79**, 100-106.
- Polizzi, P. S.; Romero, M. B.; Chiodi Boudet, L. N.; Das,

- K.; Denuncio, P. E.; Rodriguez, D. H.; Gerpe, M. S. (2014) Metallothioneins Pattern During Ontogeny of Coastal Dolphin, *Pontoporia blainvillei*, from Argentina. *Marine Pollution Bulletin*, **80**, 275-281.
- Polli, J. R.; Zhang, Y.; Pan, X. (2014) Dispersed Crude Oil Amplifies Germ Cell Apoptosis in *Caenorhabditis elegans*, Followed a Cep-1-Dependent Pathway. *Archives of Toxicology*, **88**, 543-551.
- Poynton, H. C.; Robinson, W. E.; Blalock, B. J.; Hannigan, R. E. (2014) Correlation of Transcriptomic Responses and Metal Bioaccumulation in *Mytilus edulis* L. Reveals Early Indicators of Stress. *Aquatic Toxicology*, **155**, 129-141.
- Provencher, J. F.; Bond, A. L.; Hedd, A.; Montevecchi, W. A.; Bin Muzaffar, S.; Courchesne, S. J.; Gilchrist, H. G.; Jamieson, S. E.; Merkel, F. R.; Falk, K.; Durinck, J.; Mallory, M. L. (2014) Prevalence of Marine Debris in Marine Birds from the North Atlantic. *Marine Pollution Bulletin*, **84**, 411-417.
- Purroy, A.; Requena, S.; Gili, J.-M.; Canepa, A.; Sarda, R. (2014) Spatial Assessment of Artisanal Fisheries and Their Potential Impact on the Seabed: The Cap De Creus Regional Case Study (Northwestern Mediterranean Sea). *Scientia Marina*, **78**, 449-459.
- Ramos, A. S.; Antunes, S. C.; Goncalves, F.; Nunes, B. (2014) The Gooseneck Barnacle (*Pollicipes pollicipes*) as a Candidate Sentinel Species for Coastal Contamination. *Archives of Environmental Contamination and Toxicology*, **66**, 317-326.
- Redman, A. D.; Parkerton, T. F.; Letinski, D. J.; Manning, R. G.; Adams, J. E.; Hodson, P. V. (2014) Evaluating Toxicity of Heavy Fuel Oil Fractions Using Complementary Modeling and Biomimetic Extraction Methods. *Environmental Toxicology and Chemistry*, **33**, 2094-2104.
- Regoli, F.; Giuliani, M. E. (2014) Oxidative Pathways of Chemical Toxicity and Oxidative Stress Biomarkers in Marine Organisms. *Marine Environmental Research*, **93**, 106-117.
- Reisser, J.; Shaw, J.; Hallegraeff, G.; Proietti, M.; Barnes, D. K. A.; Thums, M.; Wilcox, C.; Hardesty, B. D.; Pattiaratchi, C. (2014) Millimeter-Sized Marine Plastics: A New Pelagic Habitat for Microorganisms and Invertebrates. *Plos One*, **9**.
- Renchen, G. F.; Pittman, S. J.; Clark, R.; Caldow, C.; Gall, S.; Olsen, D.; Hill, R. L. (2014) Impact of Derelict Fish Traps in Caribbean Waters: An Experimental Approach. *Bulletin of Marine Science*, **90**, 551-563.
- Rengstorf, A. M.; Mohn, C.; Brown, C.; Wisz, M. S.; Grehan, A. J. (2014) Predicting the Distribution of Deep-Sea Vulnerable Marine Ecosystems Using High-Resolution Data: Considerations and Novel Approaches. *Deep-Sea Research Part I-Oceanographic Research Papers*, **93**, 72-82.

- Renzi, M.; Roselli, L.; Giovani, A.; Focardi, S. E.; Basset, A. (2014) Early Warning Tools for Ecotoxicity Assessment Based on *Phaeodactylum tricornerutum*. *Ecotoxicology*, **23**, 1055-1072.
- Reshitnyk, L.; Costa, M.; Robinson, C.; Dearden, P. (2014) Evaluation of Worldview-2 and Acoustic Remote Sensing for Mapping Benthic Habitats in Temperate Coastal Pacific Waters. *Remote Sensing of Environment*, **153**, 7-23.
- Rhee, J.-S.; Jeong, C.-B.; Kim, B.-M.; Lee, J.-S. (2012) P-Glycoprotein (P-GP) in the Monogonont Rotifer, *Brachionus koreanus*: Molecular Characterization and Expression in Response to Pharmaceuticals. *Aquatic Toxicology*, **114**, 104-118.
- Rhee, J.-S.; Kim, B.-M.; Choi, B.-S.; Choi, I.-Y.; Park, H.; Ahn, I.-Y.; Lee, J.-S. (2014) Transcriptome Information of the Arctic Green Sea Urchin and Its Use in Environmental Monitoring. *Polar Biology*, **37**, 1133-1144.
- Rial, D.; Murado, M. A.; Beiras, R.; Vazquez, J. A. (2014) Toxicity of Four Spill-Treating Agents on Bacterial Growth and Sea Urchin Embryogenesis. *Chemosphere*, **104**, 57-62.
- Rial, D.; Vazquez, J. A.; Murado, M. A. (2014) Toxicity of Spill-Treating Agents and Oil to Sea Urchin Embryos. *Science of the Total Environment*, **472**, 302-308.
- Ribalta, C.; Sole, M. (2014) In Vitro Interaction of Emerging Contaminants with the Cytochrome P450 System of Mediterranean Deep-Sea Fish. *Environmental Science & Technology*, **48**, 12327-12335.
- Rice, A. N.; Palmer, K. J.; Tielens, J. T.; Muirhead, C. A.; Clark, C. W. (2014) Potential Bryde's Whale (*Balaenoptera edeni*) Calls Recorded in the Northern Gulf of Mexico. *Journal of the Acoustical Society of America*, **135**, 3066-3076.
- Richmond, S.; Stevens, T. (2014) Classifying Benthic Biotopes on Sub-Tropical Continental Shelf Reefs: How Useful Are Abiotic Surrogates? *Estuarine Coastal and Shelf Science*, **138**, 79-89.
- Riera, R.; Antonio de-la-Ossa-Carretero, J. (2014) Response of Benthic Opportunistic Polychaetes and Amphipods Index to Different Perturbations in Coastal Oligotrophic Areas (Canary Archipelago, North East Atlantic Ocean). *Marine Ecology-an Evolutionary Perspective*, **35**, 354-366.
- Rochman, C. M.; Hentschel, B. T.; Teh, S. J. (2014) Long-Term Sorption of Metals Is Similar among Plastic Types: Implications for Plastic Debris in Aquatic Environments. *Plos One*, **9**.
- Rochman, C. M.; Kurobe, T.; Flores, I.; Teh, S. J. (2014) Early Warning Signs of Endocrine Disruption in Adult Fish from the Ingestion of Polyethylene with and without Sorbed Chemical Pollutants from the Marine Environment. *Science of the Total Environment*, **493**, 656-661.
- Rochman, C. M.; Lewison, R. L.; Eriksen, M.; Allen, H.; Cook, A.-M.; Teh, S. J. (2014) Polybrominated Diphenyl Ethers (PPDEs) in Fish Tissue May Be

- an Indicator of Plastic Contamination in Marine Habitats. *Science of the Total Environment*, **476**, 622-633.
- Rodd, A. L.; Creighton, M. A.; Vaslet, C. A.; Rene Rangel-Mendez, J.; Hurt, R. H.; Kane, A. B. (2014) Effects of Surface-Engineered Nanoparticle-Based Dispersants for Marine Oil Spills on the Model Organism *Artemia franciscana*. *Environmental Science & Technology*, **48**, 6419-6427.
- Rodrigues, E. T.; Pardal, M. A. (2014) The Crab *Carcinus maenas* as a Suitable Experimental Model in Ecotoxicology. *Environment International*, **70**, 158-182.
- Rogowska, J.; Kudlak, B.; Tsakovski, S.; Wolska, L.; Simeonov, V.; Namiesnik, J. (2014) Novel Approach to Ecotoxicological Risk Assessment of Sediments Cores around the Shipwreck by the Use of Self-Organizing Maps. *Ecotoxicology and Environmental Safety*, **104**, 239-246.
- Ross, B. J.; Hallock, P. (2014) Chemical Toxicity on Coral Reefs: Bioassay Protocols Utilizing Benthic Foraminifers. *Journal of Experimental Marine Biology and Ecology*, **457**, 226-235.
- Rossi, G.; Monticelli, L. (2014) Modeling the Effect of Nano-Sized Polymer Particles on the Properties of Lipid Membranes. *Journal of Physics-Condensed Matter*, **26**.
- Rožič, P. Z.; Dolenc, T.; Bazdaric, B.; Karamarko, V.; Kniewald, G.; Dolenc, M. (2014) Element Levels in Cultured and Wild Sea Bass (*Dicentrarchus labrax*) and Gilthead Sea Bream (*Sparus aurata*) from the Adriatic Sea and Potential Risk Assessment. *Environmental Geochemistry and Health*, **36**, 19-39.
- Rožič, P. Z.; Dolenc, T.; Lojen, S.; Kniewald, G.; Dolenc, M. (2014) Using Stable Nitrogen Isotopes in *Patella Sp* to Trace Sewage-Derived Material in Coastal Ecosystems. *Ecological Indicators*, **36**, 224-230.
- Rubal, M.; Veiga, P.; Reis, P. A.; Bertocci, I.; Sousa-Pinto, I. (2014) Effects of Subtle Pollution at Different Levels of Biological Organisation on Species-Rich Assemblages. *Environmental Pollution*, **191**, 101-110.
- Ruelas-Inzunza, J.; Soto-Jimenez, M. F.; Ruiz-Fernandez, A. C.; Ramos-Osuna, M.; Mones-Saucedo, J.; Paez-Osuna, F. (2014) Po-210, Cd and Pb Distribution and Biomagnification in the Yellowfin Tuna *Thunnus Albacares* and Skipjack Tuna *Katsuwonus Pelamis* from the Eastern Pacific. *Marine Pollution Bulletin*, **87**, 98-103.
- Rumbold, D.; Wasno, R.; Hammerschlag, N.; Volety, A. (2014) Mercury Accumulation in Sharks from the Coastal Waters of Southwest Florida. [Archives of Environmental Contamination and Toxicology](#), **67**, 402-412.
- Santos-Neto, E. B.; Azevedo-Silva, C. E.; Bisi, T. L.; Santos, J.; Meirelles, A. C. O.; Carvalho, V. L.; Azevedo, A. F.; Guimaraes, J. E.; Lailson-Brito, J. (2014) Organochlorine Concentrations (PCBs, DDTs, HCHs, HCB and Mirex) in Delphinids

- Stranded at the Northeastern Brazil. *Science of the Total Environment*, **472**, 194-203.
- Sarkar, A.; Bhagat, J.; Sarker, S. (2014) Evaluation of Impairment of DNA in Marine Gastropod, *Morula granulata* as a Biomarker of Marine Pollution. *Ecotoxicology and Environmental Safety*, **106**, 253-261.
- Schuyler, Q.; Hardesty, B. D.; Wilcox, C.; Townsend, K. (2014a) Global Analysis of Anthropogenic Debris Ingestion by Sea Turtles. *Conservation Biology*, **28**, 129-139.
- Schuyler, Q. A.; Wilcox, C.; Townsend, K.; Hardesty, B. D.; Marshall, N. J. (2014b) Mistaken Identity? Visual Similarities of Marine Debris to Natural Prey Items of Sea Turtles. *Bmc Ecology*, **14**.
- Schwacke, L. H.; Smith, C. R.; Townsend, F. I.; Wells, R. S.; Hart, L. B.; Balmer, B. C.; Collier, T. K.; De Guise, S.; Fry, M. M.; Guillette, L. J., Jr.; Lamb, S. V.; Lane, S. M.; McFee, W. E.; Place, N. J.; Tumlin, M. C.; Ylitalo, G. M.; Zolman, E. S.; Rowles, T. K. (2014) Health of Common Bottlenose Dolphins (*Tursiops Truncatus*) in Barataria Bay, Louisiana Following the Deepwater Horizon Oil Spill (Vol 48, Pg 93, 2014). *Environmental Science & Technology*, **48**, 10528-10528.
- Scott, N. M.; Hess, M.; Bouskill, N. J.; Mason, O. U.; Jansson, J. K.; Gilbert, J. A. (2014) The Microbial Nitrogen Cycling Potential Is Impacted by Polyaromatic Hydrocarbon Pollution of Marine Sediments. *Frontiers in Microbiology*, **5**.
- Seabra Pereira, C. D.; Abessa, D. M. S.; Choueri, R. B.; Almagro-Pastor, V.; Cesar, A.; Maranhão, L. A.; Laura Martin-Diaz, M.; Torres, R. J.; Gusso-Choueri, P. K.; Almeida, J. E.; Cortez, F. S.; Mozeto, A. A.; Silbiger, H. L. N.; Sousa, E. C. P. M.; Angel Del Valls, T.; Bainy, A. C. D. (2014) Ecological Relevance of Sentinels' Biomarker Responses: A Multi-Level Approach. *Marine Environmental Research*, **96**, 118-126.
- Seixas, T. G.; Moreira, I.; Sicilian, S.; Maim, O.; Kehrig, H. A. (2014) Mercury and Selenium in Tropical Marine Plankton and Their Trophic Successors. *Chemosphere*, **111**, 32-39.
- Seixas, T. G.; Moreira, I.; Siciliano, S.; Malm, O.; Kehrig, H. A. (2014) Differences in Methylmercury and Inorganic Mercury Biomagnification in a Tropical Marine Food Web. *Bulletin of Environmental Contamination and Toxicology*, **92**, 274-278.
- Seo, J.-Y.; Kim, M.; Lim, H.-S.; Choi, J.-W. (2014) The Macrofaunal Communities in the Shallow Subtidal Areas for the First 3 Years after the Hebei Spirit Oil Spill. *Marine Pollution Bulletin*, **82**, 208-220.
- Seoane, M.; Rioboo, C.; Herrero, C.; Cid, A. (2014) Toxicity Induced by Three Antibiotics Commonly Used in Aquaculture on the Marine Microalga *Tetraselmis suecica* (Kylin) Butch. *Marine Environmental Research*, **101**, 1-7.
- Setälä, O.; Fleming-Lehtinen, V.; Lehtiniemi, M. (2014) Ingestion and Transfer of Microplastics in the

- Planktonic Food Web. *Environmental Pollution*, **185**, 77-83.
- Shi, R.; Yu, K. (2014) Impact of Exposure of Crude Oil and Dispersant (Corexit EC9500a) on Denitrification and Organic Matter Mineralization in a Louisiana Salt Marsh Sediment. *Chemosphere*, **108**, 300-305.
- Silva-Filho, E. V.; Kuetter, V. T.; Figueiredo, T. S.; Tessier, E.; Rezende, C. E.; Teixeira, D. C.; Silva, C. A.; Donard, O. F. X. (2014) Mercury Speciation in Plankton from the Cabo Frio Bay, SE - Brazil. *Environmental Monitoring and Assessment*, **186**, 8141-8150.
- Skejjic, S.; Bojanic, N.; Matijevic, S.; Vidjak, O.; Grbec, B.; Gladan, Z. N.; Sestanovic, S.; Santic, D. (2014) Analysis of the Phytoplankton Community in the Vicinity of Domestic Sewage Outflow During Stratified Conditions. *Mediterranean Marine Science*, **15**, 574-586.
- Slijkerman, D. M. E.; de Leon, R.; de Vries, P. (2014) A Baseline Water Quality Assessment of the Coastal Reefs of Bonaire, Southern Caribbean. *Marine Pollution Bulletin*, **86**, 523-529.
- Smith, A. J.; Flemings, P. B.; Fulton, P. M. (2014) Hydrocarbon Flux from Natural Deepwater Gulf of Mexico Vents. *Earth and Planetary Science Letters*, **395**, 241-253.
- Smith, M. D.; Asche, F.; Benneer, L. S.; Oglend, A. (2014) Spatial-Dynamics of Hypoxia and Fisheries: The Case of Gulf of Mexico Brown Shrimp. *Marine Resource Economics*, **29**, 111-131.
- Snyder, R. A.; Vestal, A.; Welch, C.; Barnes, G.; Pelot, R.; Ederington-Hagy, M.; Hileman, F. (2014) Pah Concentrations in Coquina (*Donax spp.*) on a Sandy Beach Shoreline Impacted by a Marine Oil Spill. *Marine Pollution Bulletin*, **83**, 87-91.
- Sondergaard, J.; Bach, L.; Gustavson, K. (2014) Measuring Bioavailable Metals Using Diffusive Gradients in Thin Films (DGT) and Transplanted Seaweed (*Fucus vesiculosus*), Blue Mussels (*Mytilus edulis*) and Sea Snails (*Littorina saxatilis*) Suspended from Monitoring Buoys near a Former Lead-Zinc Mine in West Greenland. *Marine Pollution Bulletin*, **78**, 102-109.
- Staniszewska, M.; Falkowska, L.; Grabowski, P.; Kwasniak, J.; Mudrak-Cegiolk, S.; Reindl, A. R.; Sokolowski, A.; Szumilo, E.; Zgrundo, A. (2014) Bisphenol a, 4-Tert-Octylphenol, and 4-Nonylphenol in the Gulf of Gdansk (Southern Baltic). *Archives of Environmental Contamination and Toxicology*, **67**, 335-347.
- Stankovic, S.; Tanaskovski, B.; Zlatic, B.; Arsenovic, M.; Pezo, L. (2014) Analysis of Trace Elements in Surface Sediments, Mussels, Seagrass and Seawater Along the Southeastern Adriatic Coast - a Chemometric Approach. *Pure and Applied Chemistry*, **86**, 1111-1127.
- Stark, J. S.; Kim, S. L.; Oliver, J. S. (2014) Anthropogenic Disturbance and Biodiversity of Marine Benthic Communities in Antarctica: A Regional

- Comparison. *Plos One*, **9**.
- Stauffert, M.; Duran, R.; Gassie, C.; Cravo-Laureau, C. (2014) Response of Archaeal Communities to Oil Spill in Bioturbated Mudflat Sediments. *Microbial Ecology*, **67**, 108-119.
- Stentiford, G. D.; Massoud, M. S.; Al-Mudhhi, S.; Al-Sarawi, M. A.; Al-Enezi, M.; Lyons, B. P. (2014) Histopathological Survey of Potential Biomarkers for the Assessment of Contaminant Related Biological Effects in Species of Fish and Shellfish Collected from Kuwait Bay, Arabian Gulf. *Marine Environmental Research*, **98**, 60-67.
- Stewart, M.; Olsen, G.; Hickey, C. W.; Ferreira, B.; Jelic, A.; Petrovic, M.; Barcelo, D. (2014) A Survey of Emerging Contaminants in the Estuarine Receiving Environment around Auckland, New Zealand. *Science of the Total Environment*, **468**, 202-210.
- Storelli, M. M.; Zizzo, N. (2014) Occurrence of Organochlorine Contaminants (PCBs, PCDDs and PCDFs) and Pathologic Findings in Loggerhead Sea Turtles, *Caretta caretta*, from the Adriatic Sea (Mediterranean Sea). *Science of the Total Environment*, **472**, 855-861.
- Strogyloudi, E.; Pancucci-Papadopoulou, M.-A.; Papadopoulos, G. L. (2014) Metal and Metallothionein Concentrations in *Paracentrotus lividus* from Amvrakikos Gulf (Ionian Sea-Greece). *Environmental Monitoring and Assessment*, **186**, 5489-5499.
- Subedi, B.; Yun, S.; Jayaraman, S.; Bergen, B. J.; Kannan, K. (2014) Retrospective Monitoring of Persistent Organic Pollutants, Including PCBs, PBDEs, and Polycyclic Musks in Blue Mussels (*Mytilus edulis*) and Sediments from New Bedford Harbor, Massachusetts, USA: 1991-2005. *Environmental Monitoring and Assessment*, **186**, 5273-5284.
- Suehring, R.; Byer, J.; Freese, M.; Pohlmann, J.-D.; Wolschke, H.; Moeller, A.; Hodson, P. V.; Alaei, M.; Hanel, R.; Ebinghaus, R. (2014) Brominated Flame Retardants and Dechloranes in European and American Eels from Glass to Silver Life Stages. *Chemosphere*, **116**, 104-111.
- Sugahara, Y.; Kawaguchi, M.; Itoyama, T.; Kurokawa, D.; Tosa, Y.; Kitamura, S.-I.; Handoh, I. C.; Nakayama, K.; Murakami, Y. (2014) Pyrene Induces a Reduction in Midbrain Size and Abnormal Swimming Behavior in Early-Hatched Pufferfish Larvae. *Marine Pollution Bulletin*, **85**, 479-486.
- Sukumaran, S.; Mulik, J.; Rokade, M. A.; Kamble, A. (2014) Impact of 'Chitra' Oil Spill on Tidal Pool Macrobenthic Communities of a Tropical Rocky Shore (Mumbai, India). *Estuaries and Coasts*, **37**, 1415-1431.
- Sun, Y.-X.; Hao, Q.; Xu, X.-R.; Luo, X.-J.; Wang, S.-L.; Zhang, Z.-W.; Mai, B.-X. (2014) Persistent Organic Pollutants in Marine Fish from Yongxing Island, South China Sea: Levels, Composition Profiles and Human Dietary

- Exposure Assessment. *Chemosphere*, **98**, 84-90.
- Suzdalev, S.; Gulbinskas, S.; Sivkov, V.; Bukanova, T. (2014) Solutions for Effective Oil Spill Management in the South-Eastern Part of the Baltic Sea. *Baltica*, **27**, 3-8.
- Symons, J.; Pirotta, E.; Lusseau, D. (2014) Sex Differences in Risk Perception in Deep-Diving Bottlenose Dolphins Leads to Decreased Foraging Efficiency When Exposed to Human Disturbance. [*Journal of Applied Ecology*](#), **51**, 1584-1592.
- Taylor, A. M.; Maher, W. A. (2014) Exposure-Dose-Response of *Tellina deltoidalis* to Contaminated Estuarine Sediments 3. Selenium Spiked Sediments. *Comparative Biochemistry and Physiology C-Toxicology & Pharmacology*, **166**, 34-43.
- Taylor, J. R.; DeVogelaere, A. P.; Burton, E. J.; Frey, O.; Lundsten, L.; Kuhnz, L. A.; Whaling, P. J.; Lovera, C.; Buck, K. R.; Barry, J. P. (2014) Deep-Sea Faunal Communities Associated with a Lost Intermodal Shipping Container in the Monterey Bay National Marine Sanctuary, Ca. *Marine Pollution Bulletin*, **83**, 92-106.
- Thera, J. C.; Rumbold, D. G. (2014) Biomagnification of Mercury through a Subtropical Coastal Food Web Off Southwest Florida. [*Environmental Toxicology and Chemistry*](#), **33**, 65-73.
- Thomas, J. C.; Wafula, D.; Chauhan, A.; Green, S. J.; Gragg, R.; Jagoe, C. (2014) A Survey of Deepwater Horizon (Dwh) Oil-Degrading Bacteria from the Eastern Oyster Biome and Its Surrounding Environment. *Frontiers in Microbiology*, **5**.
- Tian, S.; Zhang, Y.; Song, C.; Zhu, X.; Xing, B. (2014) Titanium Dioxide Nanoparticles as Carrier Facilitate Bioaccumulation of Phenanthrene in Marine Bivalve, Ark Shell (*Scapharca subcrenata*). *Environmental Pollution*, **192**, 59-64.
- Tomanek, L. (2014) Proteomics to Study Adaptations in Marine Organisms to Environmental Stress. *Journal of Proteomics*, **105**, 92-106.
- Tsangaris, C.; Stroglyoudi, E.; Hatzianestis, I.; Catsiki, V.-A.; Panagiotopoulos, I.; Kapsimalis, V. (2014) Impact of Dredged Urban River Sediment on a Saronikos Gulf Dumping Site (Eastern Mediterranean): Sediment Toxicity, Contaminant Levels, and Biomarkers in Caged Mussels. *Environmental Science and Pollution Research*, **21**, 6146-6161.
- Tsangaris, C.; Stroglyoudi, E.; Hatzianestis, I.; Catsiki, V.-A.; Panagiotopoulos, I.; Kapsimalis, V. (2014) Impact of Dredged Urban River Sediment on a Saronikos Gulf Dumping Site (Eastern Mediterranean): Sediment Toxicity, Contaminant Levels, and Biomarkers in Caged Mussels. *Environmental Science and Pollution Research*, **21**, 6146-6161.
- Tu, T.; Li, S.; Chen, L.; Zheng, F.; Huang, X.-G. (2014) Effect of Coastal Eutrophication on Heavy Metal Bioaccumulation and Oral Bioavailability in the

- Razor Clam, *Sinonovacula constricta*. *Aquatic Toxicology*, **155**, 269-274.
- Tucca, F.; Diaz-Jaramillo, M.; Cruz, G.; Silva, J.; Bay-Schmith, E.; Chiang, G.; Barra, R. (2014) Toxic Effects of Antiparasitic Pesticides Used by the Salmon Industry in the Marine Amphipod *Monocorophium insidiosum*. *Archives of Environmental Contamination and Toxicology*, **67**, 139-148.
- Van Cauwenberghe, L.; Janssen, C. R. (2014) Microplastics in Bivalves Cultured for Human Consumption. *Environmental Pollution*, **193**, 65-70.
- van Denderen, P. D.; Hintzen, N. T.; Rijnsdorp, A. D.; Ruardij, P.; van Kooten, T. (2014) Habitat-Specific Effects of Fishing Disturbance on Benthic Species Richness in Marine Soft Sediments. *Ecosystems*, **17**, 1216-1226.
- Van Geest, J. L.; Burridge, L. E.; Fife, F. J.; Kidd, K. A. (2014a) Feeding Response in Marine Copepods as a Measure of Acute Toxicity of Four Anti-Sea Lice Pesticides. *Marine Environmental Research*, **101**, 145-152.
- Van Geest, J. L.; Burridge, L. E.; Kidd, K. A. (2014b) Toxicity of Two Pyrethroid-Based Anti-Sea Lice Pesticides, Alphamax (R) and Excis (R), to a Marine Amphipod in Aqueous and Sediment Exposures. *Aquaculture*, **434**, 233-240.
- Van Geest, J. L.; Burridge, L. E.; Kidd, K. A. (2014c) The Toxicity of the Anti-Sea Lice Pesticide Alphamax (R) to the Polychaete Worm *Nereis virens*. *Aquaculture*, **430**, 98-106.
- Vasanthi, L. A.; Muruganandam, A.; Revathi, P.; Baskar, B.; Jayapriyan, K.; Baburajendran, R.; Munuswamy, N. (2014) The Application of Histo-Cytopathological Biomarkers in the Mud Crab *Scylla serrata* (Forsk.) to Assess Heavy Metal Toxicity in Pulicat Lake, Chennai. *Marine Pollution Bulletin*, **81**, 85-93.
- Velusamy, A.; Kumar, P. S.; Ram, A.; Chinnadurai, S. (2014) Bioaccumulation of Heavy Metals in Commercially Important Marine Fishes from Mumbai Harbor, India. *Marine Pollution Bulletin*, **81**, 218-224.
- Ventikos, N. P.; Sotiropoulos, F. S. (2014) Disutility Analysis of Oil Spills: Graphs and Trends. *Marine Pollution Bulletin*, **81**, 116-123.
- Ventura, S. P. M.; Silva, F. A. E.; Goncalves, A. M. M.; Pereira, J. L.; Goncalves, F.; Coutinho, J. A. P. (2014) Ecotoxicity Analysis of Cholinium-Based Ionic Liquids to *Vibrio fischeri* Marine Bacteria. *Ecotoxicology and Environmental Safety*, **102**, 48-54.
- Verlis, K. M.; Campbell, M. L.; Wilson, S. P. (2014) Marine Debris Is Selected as Nesting Material by the Brown Booby (*Sula leucogaster*) within the Swain Reefs, Great Barrier Reef, Australia. *Marine Pollution Bulletin*, **87**, 180-190.
- Viaene, K. P. J.; Janssen, C. R.; de Hoop, L.; Hendriks, A. J.; De Laender, F. (2014) Evaluating the Contribution of Ingested Oil Droplets to the Bioaccumulation of Oil Components - a

- Modeling Approach. *Science of the Total Environment*, **499**, 99-106.
- Vidal-Linan, L.; Bellas, J.; Etxebarria, N.; Nieto, O.; Beiras, R. (2014) Glutathione S-Transferase, Glutathione Peroxidase and Acetylcholinesterase Activities in Mussels Transplanted to Harbour Areas. *Science of the Total Environment*, **470**, 107-116.
- Walter, S. T.; Carloss, M. R.; Hess, T. J.; Leberg, P. L. (2014) Demographic Trends of Brown Pelicans in Louisiana before and after the Deepwater Horizon Oil Spill. *Journal of Field Ornithology*, **85**, 421-429.
- Wang, H.; Ho, K. T.; Scheckel, K. G.; Wu, F.; Cantwell, M. G.; Katz, D. R.; Horowitz, D. B.; Boothman, W. S.; Burgess, R. M. (2014) Toxicity, Bioaccumulation, and Biotransformation of Silver Nanoparticles in Marine Organisms. *Environmental Science & Technology*, **48**, 13711-13717.
- Wang, J.; Shi, T.; Yang, X.; Han, W.; Zhou, Y. (2014) Environmental Risk Assessment on Capsaicin Used as Active Substance for Antifouling System on Ships. *Chemosphere*, **104**, 85-90.
- Wang, J.; Zhao, F.; Yang, X.; Han, W.; Long, K.; Zhou, Y. (2014) Marine Environmental Risk Assessment Method for Active Substances Used in Antifouling Systems on Ships in China. *Environmental Engineering, Pts 1-4*, **864-867**, 962-972.
- Wang, Y.; Wang, J.; Mu, J.; Wang, Z.; Yao, Z.; Lin, Z. (2014) Aquatic Predicted No-Effect Concentration for Three Polycyclic Aromatic Hydrocarbons and Probabilistic Ecological Risk Assessment in Liaodong Bay of the Bohai Sea, China. *Environmental Science and Pollution Research*, **21**, 148-158.
- Wang, Z.; Wang, X.; Ke, C. (2014) Bioaccumulation of Trace Metals by the Live Macroalga *Gracilaria lemaneiformis*. *Journal of Applied Phycology*, **26**, 1889-1897.
- Waszak, I.; Dabrowska, H.; Komar-Szymczak, K. (2014) Comparison of Common Persistent Organic Pollutants (POPs) in Flounder (*Platichthys flesus*) from the Vistula (Poland) and Douro (Portugal) River Estuaries. *Marine Pollution Bulletin*, **81**, 225-233.
- Watts, A. J. R.; Lewis, C.; Goodhead, R. M.; Beckett, S. J.; Moger, J.; Tyler, C. R.; Galloway, T. S. (2014) Uptake and Retention of Microplastics by the Shore Crab *Carcinus maenas*. *Environmental Science & Technology*, **48**, 8823-8830.
- Weller, F.; Cecchini, L.-A.; Shannon, L.; Sherley, R. B.; Crawford, R. J. M.; Altwegg, R.; Scott, L.; Stewart, T.; Jarre, A. (2014) A System Dynamics Approach to Modelling Multiple Drivers of the African Penguin Population on Robben Island, South Africa. *Ecological Modelling*, **277**, 38-56.
- Williams, T. D.; Davies, I. M.; Wu, H.; Diab, A. M.; Webster, L.; Viant, M. R.; Chipman, J. K.; Leaver, M. J.; George, S. G.; Moffat, C. F.; Robinson, C. D. (2014) Molecular Responses of

- European Flounder (*Platichthys flesus*) Chronically Exposed to Contaminated Estuarine Sediments. *Chemosphere*, **108**, 152-158.
- Wise, C. F.; Wise, J. T. F.; Wise, S. S.; Thompson, W. D.; Wise, J. P., Jr.; Wise, J. P., Sr. (2014) Chemical Dispersants Used in the Gulf of Mexico Oil Crisis Are Cytotoxic and Genotoxic to Sperm Whale Skin Cells. *Aquatic Toxicology*, **152**, 335-340.
- Witt, M.; Kobusinska, M.; Maciak, J.; Niemirycz, E. (2014) Geostatistical Methods for Estimation of Toxicity of Marine Bottom Sediments Based on the Gdansk Basin Area. *Oceanological and Hydrobiological Studies*, **43**, 247-256.
- Won, E.-J.; Ra, K.; Kim, K.-T.; Lee, J.-S.; Lee, Y.-M. (2014) Three Novel Superoxide Dismutase Genes Identified in the Marine Polychaete *Perinereis nuntia* and Their Differential Responses to Single and Combined Metal Exposures. *Ecotoxicology and Environmental Safety*, **107**, 36-45.
- Word, J. Q.; Clark, J. R.; Word, L. S. (2015) Comparison of the Acute Toxicity of Corexit 9500 and Household Cleaning Products. *Human and Ecological Risk Assessment*, **21**, 707-725.
- Xia, B.; Chen, B.; Sun, X.; Cui, Y.; Zhao, J.; Qu, K. (2014) Toxicological Effects of Crude Oil: Integrated Biomarker Responses in the Hepatopancreas of Clam *Ruditapes philippinarum*. *Asian Journal of Chemistry*, **26**, 3631-3638.
- Xu, E. G. B.; Liu, S.; Ying, G.-G.; Zheng, G. J. S.; Lee, J. H. W.; Leung, K. M. Y. (2014) The Occurrence and Ecological Risks of Endocrine Disrupting Chemicals in Sewage Effluents from Three Different Sewage Treatment Plants, and in Natural Seawater from a Marine Reserve of Hong Kong. *Marine Pollution Bulletin*, **85**, 352-362.
- Xu, W. Z.; Cheung, S. G.; Shin, P. K. S. (2014) Structure and Taxonomic Composition of Free-Living Nematode and Macrofaunal Assemblages in a Eutrophic Subtropical Harbour, Hong Kong. *Marine Pollution Bulletin*, **85**, 764-773.
- Yednock, B. K.; Neigel, J. E. (2014) An Investigation of Genetic Population Structure in Blue Crabs, *Callinectes sapidus*, Using Nuclear Gene Sequences. *Marine Biology*, **161**, 871-886.
- Yi, A. X.; Han, J.; Lee, J.-S.; Leung, K. M. Y. (2014) Ecotoxicity of Triphenyltin on the Marine Copepod *Tigriopus japonicus* at Various Biological Organisations: From Molecular to Population-Level Effects. *Ecotoxicology*, **23**, 1314-1325.
- Yorio, P.; Marinao, C.; Suarez, N. (2014) Kelp Gulls (*Larus dominicanus*) Killed and Injured by Discarded Monofilament Lines at a Marine Recreational Fishery in Northern Patagonia. *Marine Pollution Bulletin*, **85**, 186-189.
- Yoshida, H.; Clavreul, J.; Scheutz, C.; Christensen, T. H. (2014) Influence of Data Collection Schemes on the Life Cycle Assessment of a Municipal Wastewater Treatment Plant. *Water Research*, **56**, 292-303.

- Yusof, S.; Ismail, A.; Alias, M. S. (2014) Effect of Glyphosate-Based Herbicide on Early Life Stages of Java Medaka (*Oryzias javanicus*): A Potential Tropical Test Fish. *Marine Pollution Bulletin*, **85**, 494-498.
- Zalewska, T. (2014) Bioaccumulation of Gamma Emitting Radionuclides in *Polysiphonia fucoides*. *Journal of Radioanalytical and Nuclear Chemistry*, **299**, 1489-1497.
- Zhang, H.; Pan, L.; Tao, Y. (2014) Toxicity Assessment of Environmental Pollutant Phenanthrene in Clam *Venerupis philippinarum* Using Oxidative Stress Biomarkers. *Environmental Toxicology and Pharmacology*, **37**, 697-704.
- Zhao, L.; Zhang, Y.; Liang, J.; Xu, X.; Wang, H.; Yang, F.; Yan, X. (2014) Environmental Cadmium Exposure Impacts Physiological Responses in Manila Clams. *Biological Trace Element Research*, **159**, 241-253.
- Zheng, M.; Ahuja, M.; Bhattacharya, D.; Clement, T. P.; Hayworth, J. S.; Dhanasekaran, M. (2014) Evaluation of Differential Cytotoxic Effects of the Oil Spill Dispersant Corexit 9500. *Life Sciences*, **95**, 108-117.
- Zhu, B.; Lai, N. L. S.; Wai, T.-C.; Chan, L. L.; Lam, J. C. W.; Lam, P. K. S. (2014) Changes of Accumulation Profiles from PBDEs to Brominated and Chlorinated Alternatives in Marine Mammals from the South China Sea. *Environment International*, **66**, 65-70.

Table 1—Examples of chemical residues reported from marine organisms. See text for more data. Always consult cited paper to confirm actual values, units and confidence limits. See footnotes for concentration units.

Chemical	Organism Location	Geographical µg/g dry weight	Concentration,	Reference	
Aluminum	Pelecypods	India	647-73019	Bhattacharya et al.	
		Namibia	1.02-3.2	Dahms et al.	
		So. Africa	5-1,400 ^c	Greenfield et al.	
Arsenic	Seaweed	Greenland	0.2-55 ^c	Søndergaard et al.	
	Macroalgae	Antarctica	14.8	Majer et al.	
	Nemerteans	Antarctica	18.6	Majer et al.	
	Amphipods	Antarctica	7.8-9.8	Majer et al.	
	Isopods	Antarctica	8.4	Majer et al.	
	Decapods	Antarctica	1.7-12	Majer et al.	
	Pelecypods	China	0.098-0.3 ^c	Tu et al.	
		Korea	3-25	Hong et al.a	
		So. Africa	1-13 ^a	Greenfield et al.	
		Adriatic Sea	4-30	Stanković et al.	
		Greenland	14-16 ^c	Søndergaard et al.	
		Gastropods	Greenland	11-14 ^c	Søndergaard et al.
			Antarctica	6-26	Majer et al.
		Echinoderms	Antarctica	5.02-6	Majer et al.
	Fish	Korea	0.64-5.4	,a et al.	
		Maryland	16-66 ^{a,c}	Dutton and Fisher	
		Adriatic Sea	4.01	Rozic et al.	
	Mammals	Portugal	<0.67-1.2 ^a	Méndez-Fernandez et al. b	
	Cadmium	Green algae	Gulf of California	0.7-2.4	Hernández-Almaraz et al.
			Egypt	1.1-1.5	El-Din et al.
Brown algae		Gulf of California	5.2-9.6	Hernández-Almaraz et al.	
		Egypt	2.2-8	El-Din et al.	
Red algae		Gulf of California	5.1-6	Hernández-Almaraz et al.	
		Egypt	1-4	El-Din et al.	
Sea grass		Adriatic Sea	0.22-1.86	Stanković et al.	
Plankton		India	4.2-21.6	Bhattacharya et al.	
Nemerteans		Antarctica	5	Majer et al.	
Polychaetes		Persian Gulf	1.2-82.2	Amoozadeh et al.	
		France	0.1-0.5 ^c	Dedeh et al.	
Barnacles		Persian Gulf	2.73-484	Amoozadeh et al.	
Amphipods		Antarctica	0.5-2.3	Majer et al.	
Isopods		Antarctica	1.07	Majer et al.	
Decapods		Persian Gulf	0	Dadar et al.	
		Persian Gulf	0.76-5.6	Amoozadeh et al.	
Pelecypods		Persian Gulf	7.4-119,4	Amoozadeh et al.	
		Namibia	0.12-1.65	Dahms et al.	
		So.Africa	0.15-3.8 ^c	Greenfield et al.	
		France	0.1-2.7 ^c	Dedeh. et al.	
		Adriatic Sea	1.97-4.1	Stanković. et al.	
Gastropods		Greenland	2.44-3.03 ^c	Søndergaard et al.	
		Antarctica	1.76	Majer et al.	
		Gulf of California	4.6-17.7	Hernández-Almaraz et al.	
Echinoderms		Greece	0.04-1.2	Strogylaudi et al.	
		Antarctica	0.03-0.98	Majer et al.	
Fish		Sweden	0.11-0.16	Boalt et al.	
		Greece	0.1-1.2	Giannakopoulou & Neofitou	
		Maryland	6-46 ^{a,c}	Dutton & Fisher	
Cadmium (cont.)		Fish	Gulf of California	0.18-20.6	Ruelas-Inzuinza. et al.

	Mammals	Adriatic Sea	0.01-0.016	Rozic et al.
	Birds	Portugal	<0.07-30 ^a	Méndez-Fernandez et al.b
		France	0.03-18.7	Leucie et al.
Chromium	Seaweed	Greenland	0.05-0.6 ^c	Søndergaard et al.
	Seagrass	Adriatic Sea	3.2-37	Stantović et al.
	Plankton	India	21-194	Bhattacharya et al.
	Pelecypods	Namibia	0.36-1.2	Dahms et al.
		So. Africa	25-110 ^c	Greenfield et al.
	Adriatic Sea		0.8-2.9	Stantović et al.
		Greenland	1.26-2.65 ^c	Søndergaard et al.
	Gastropods	Greenland	0.4-1.08 ^c	Søndergaard et al.
	Echinoderms	Greece	0.3-13.7	Strogylaudi et al.
	Fish	India	0-1.55	Velusany et al.
		Greece	0-5	Giannakopoulou & Neufitou
		Maryland	4.-54a.c	Dutton & Fisher
		Adriatic Sea	49.1	Rozic et al.
Mammals	Kuwait	4-6	Bu-Olayan & Thomas	
	Portugal	<0.7-1.8 ^a	Méndez-Fernandez et al. b	
Cobalt	Plankton	India	32-111	Bhattacharya et al.
	Pelecypods	Namibia	0.36-1.2	Dahms et al.
	Decapods	Persian Gulf	0	Dadar et al.
	Mammals	Portugal	<0.07 ^a	Méndez-Fernandez et al. b
Copper	Green algae	Egypt	2.5-6	El-Din et al.
		Gulf of California	1.7-3.2	Hernández-Almaraz et al.
	Brown algae	Egypt	4-4.5	El-Din et al.
		Gulf of California	1.5-4.5	Hernández-Almaraz et al.
	Red algae	Egypt	3-4.2	El-Din et al.
		Gulf of California	1.6-5.1	Hernández-Almaraz et al.
	Sea weed	Greenland	0.1-27.5 ^c	Søndergaard et al.
	Macroalgae	Antarctica	3.5	Majer et al.
	Sea grass	Adriatic Sea	0.3-16.5	Stanković et al.
	Nemerteans	Antarctic	18.5	Majer et al.
	Amphipods	Antarctic	41-58	Majer et al.
	Isopods	Antarctic	119	Majer et al.
	Decapods	Persian Gulf	0.06-4.14	Dadar et al.
	Pelecypods	Namibia	0.72-1.4	Dahms et al.
		China	0.15-0.38 ^c	Tu et al.
		So. Africa	2-110 ^a	Greenfield et al.
		Adriatic Sea	3.7-88.4	Stanković et al.
		Greenland	8.9-25.7 ^c	Søndergaard et al.
		Greenland	24-75.6 ^c	Søndergaard et al.
		Gulf of California	<0.07-1.3	Hernández-Almaraz et al.
	Echinoderms	Antarctic	3.4	Majer et al.
		Greece	2.1-11.8	Stronglylandi et al.
	Fish	Antarctic	3.6-5.2	Majer et al.
India		0.51-5.8	Velusang et al.	
Neafutou		Greece	0.1-10	Giannakopoulou &
		Adriatic Sea	1.8-1.97	Kozic et al.
		Kuwait	7-10	Bu-Olayan & Thomas
	Mammals	Portugal	2.7-8.7	Méndez-Fernandez et al. b
Iron	Green algae	Gulf of California	330-372	Hernández-Almaraz et al.
	Brown algae	Gulf of California	116-135	Hernández-Almaraz et al.
	Red algae	Gulf of California	98-197	Hernández-Almaraz et al.

	Sea weed	Greenland	85-132 ^c	Søndergaard et al.
	Plankton	India	1,380-51,118	Bhattacharya et al.
	Decapods	Persian Gulf	0.62-1.98	Dadar et al.
	Pelecypods	Namibia	0.89-3.3	Dahms et al.
		China	0.03-0.05 ^c	Tu et al.
		So. Africa	50-190 ^c	Greenfield et al.
		Adriatic Sea	53-719	Stanković et al.
	Gastropods	Greenland	267-1,292 ^c	Søndergaard et al.
		Greenland	296-617 ^c	Søndergaard et al.
		Gulf of California	36.4-117.5	Hernández-Almaraz et al.
	Echinoderms	Greece	44-355	Stroglyandi et al.
	Fish	India	32.1-240.5	Velusamy et al.
		Kuwait	12-14	Bu-Olayau & Thomas
	Mammals	Portugal	123-398 ^a	Méndez-Fernandez et al. b
Lead	Green algae	Egypt	2-20	El-Din et al.
		Gulf of California	<0.07-3.9	Hernández-Almaraz et al.
	Brown algae	Egypt	15-18	El-Din et al.
		Gulf of California	<0.07-2.8	Hernández-Almaraz et al.
	Red algae	Egypt	1-28	El-Din et al.
		Gulf of California	<0.07-2.4	Hernández-Almaraz et al.
	Sea weed	Greenland	0.03-4.3 ^c	Søndergaard et al.
	Macroalgae	Antarctica	4.5	Majer et al.
	Sea grass	Adriatic Sea	2.2-19.1	Stanković et al.
	Plankton	India	0.04-97.5	Bhattacharya et al.
	Nemertean	Antarctica	2.6	Majer et al.
	Polychaetes	Persian Gulf	0.09-24	Amoozadel et al.
		France	2-10 ^c	Dedeh et al.
	Barnacles	Persian Gulf	2.1-452	Amoozadeh et al.
	Amphipods	Antarctic	4.26-9.3	Majer et al.
	Isopods	Antarctica	8.7	Majer et al.
	Decapods	Persian Gulf	0.01-2.54	Dadar et al.
		Persian Gulf	0.01-6.9	Amoozadeh et al.
	Pelecypods	Persian Gulf	0.05-23.8	Amoozadeh et al.
		Namibia	1.55-2.35	Dahms et al.
		Portugal	0.65 ^a	Freitas et al.
		China	0.005-0.015 ^c	Tu et al.
		So. Africa	0.5-4 ^c	Greenfield et al.
		France	2-45 ^c	Dedeh et al.
		Adriatic Sea	1.15-7	Stanković
	Gastropods	Greenland	1.96-13 ^c	Søndergaard et al.
		Gulf of California	<0.02-3.7	Hernández-Almaraz et al.
		Antarctica	5.9	Majer et al.
	Echinoderms	Antarctica	4.3-8.9	Majer et al.
	Fish	India	0-0.24	Velusamy et al.
		Gulf of California	0.04-0.06	Ruelas-Inzunza et al.
		Adriatic Sea	0.1-0.65	Rozic et al.
		Kuwait	2-4	Buo-Olayau & Thomas
Portugal		<0.07 ^a	Méndez-Fernandez et al. b	
Birds	France	0.01-0.38	Lucia et al.	
Manganese	Plankton	India	122-1,066	Bhattacharya et al.
	Polychaetes	Greece	0.5-13	Kalantzi et al.
	Anthropods	Greece	0.4-2.1	Kalantzi et al.
Manganese (cont.)	Molluscs	Greece	0.1-0.4	Kalantzi
		Adriatic Sea	2-13	Stanković et al.
	Pelecypods	Namibia	0.64-1.98	Dahms et al.
		So. Africa	2-64 ^c	Greenfield et al.
	Echinoderms	Greece	0.6-2.4	Kalantzi et al.
	Fish	India	1.17-7.75	Velusamy et al.
Mammals	Portugal	<2.5-3.4 ^a	Méndez-Fernandez et al. b	

Mercury	Phytoplankton	Brazil	0.0017-0.00175	Seixas et al.
		Brazil	0.0013 ^a	Silva-Filho et al.
	Seaweed	Greenland	0.03 ^c	Søndergaard et al.
	Zooplankton	Brazil	0.025 ^a	Silva-Filho et al.
	Polychaetes	France	0.3-2 ^c	Dedeh et al.
		Florida	0.013 ^a	Thera & Rumbold
	Isopods	Florida	0.015 ^a	Thera & Rumbold
	Decapods	Florida	0.014-0.098 ^a	Thera & Rumbold
		Mediterranean Sea		1.8 Cresson et al.
	Pelecypods	France	0.1-2	Dedeh et al.
		Greenland	0.1-0.11 ^c	Søndergaard et al.
	Gastropods	China	27.7	Ho & Leung
		Greenland	0.04-0.05 ^c	Søndergaard et al.
		Florida	0.008-0.02 ^a	Thera & Rumbold
	Cephalopods	Florida	0.017 ^a	Thera & Rumbold
	Echinoderms	Florida	0.004-0.03 ^a	Thera & Rumbold
		Fish	Brazil	0.08-0.34
	Florida		0.00037	Rumbold et al.
	Florida		0.03-0.48	Huge et al.
	Florida		0.04-2.84 ^a	Thera & Rumbold
Maryland	4-58 ^{a,c}		Dutton & Fisher	
India	0.01-0.23		Velusamy et al.	
Sweden	0.52-1.63		Boalt et al.	
Adriatic Sea	0.07-4.4		Horvat et al.	
Mediterranean Sea			1.3-7.13 Cresson et al.	
Kuwait	1-2		Bu-Olaguo & Thomas	
Mammals	Brazil	0.046-1.8	Seixas	
	Portugal	1.6-31 ^a	Méndez-Fernandez et al. b	
Birds	France	0.15	Lucia et al.	
Nickel	Seaweed	Greenland	0.08-2.5 ^c	Søndergaard et al.
	Macroalgae	Antarctica	0.85	Majer et al.
	Nemertean	Antarctica	0.48	Majer et al.
	Amphipods	Antarctica	0.86-1.8	Majer et al.
	Isopods	Antarctica	0.78	Majer et al.
	Decapods	Persian Gulf	0.29-0.31	Dadar et al.
	Pelecypods	Namibia	0.35-1.95	Dahms et al.
		China	0.02-0.04 ^c	Tu et al.
		So. Africa	0-15 ^c	Greenfield et al.
		Adriatic Sea	0.8-5	Stanković et al.
	Gastropods	Greenland	2.2-9.4 ^c	Søndergaard et al.
		Greenland	2-3.08 ^c	Søndergaard et al.
		So. Africa	0.37	Majer et al.
	Echinoderms	Greece	2.4-26.7	Strogloudi et al.
		So. Africa	0.6-1.16	Majer et al.
	Fish	Kuwait	6-8	Bu-Olayou & Thomas
Mammals	Portugal	<0.2-0.8 ^a	Méndez-Fernandez et al. b	
Selenium	Fish	Adriatic Sea	0.87-1.78	Rozic et al.
	Mammals	Portugal	2.7-16.9 ^a	Méndez-Fernandez et al. b
Silver	Seaweed	Greenland	0.01-0.19 ^c	Søndergaard et al.
	Pelecypods	Greenland	0.04-0.1 ^c	Søndergaard et al.
	Gastropods	Greenland	1.08-1.35 ^c	Søndergaard et al.
	Mammals	Portugal	<0.07 ^a	Méndez-Fernandez et al. b
	Birds	France	0.01-1.7	Lucia et al.
Strontium	Pelecypods	So. Africa	5-120 ^c	Greenfield et al.
Tin	Gastropods	R. Cabo Verde	0.005-0.037	Lopes-dos-Santos et al.

	Fish	Brazil	0.032-0.33	dos Santos et al.
Vanadium	Polychaetes	Greece	3.4-53.8	Kalantzi et al.
	Arthropods	Greece	1.7-6.6	Kalantzi et al.
	Molluscans	Greece	0.4-3.3	Kalantzi et al.
	Pelecypods	China	0.015-0.03 ^c	Tu et al.
	Echinoderms	Greece	1.3-4.5	Kalantzi et al.
	Mammals	Portugal	<0.1 ^a	Méndez-Fernandez et al. b
Zinc	Green algae	Egypt	8-40	El-Din et al.
		Gulf of California	16-20.8	Hernández-Almaraz et al.
	Brown algae	Egypt	17-45	El-Ding
		Gulf of California	15.3-23	Hernández-Almaraz et al.
	Red algae	Egypt	10-43	El-Din
		Gulf of California	9.1-18.7	Hernández-Almaraz et al.
	Seaweed	Greenland	0.05-40 ^c	Sandergaard et al.
	Macroalgae	Antarctica	21.7	Majer et al.
	Plankton	India	725-1,670	Bhattacharya et al.
	Nemertean	Antarctica	158	Majer et al.
	Amphipods	Antarctica	52	Majer et al.
	Isopods	Antarctica	53	Majer et al.
	Decapods	Persian Gulf	0.34-0.38	Dadar et al.
	Pelecypods	Namibia	1.37-2.92	Dahms et al.
		So. Africa	50-220 ^c	Greenfield et al.
		Adriatic Sea	2-245	Stanković et al.
		Greenland	8.9-30.1 ^c	Søndergaard et al.
	Gastropods	Greenland	24-75 ^c	Søndergaard et al.
		Gulf of California	24.7-30	Hernández-Almaraz et al.
		Antarctica	23.7	Majer et al.
	Echinoderms	Antarctica	58-353	Majer et al.
		Greece	19-1,024	Stroglyocede et al.
	Fish	India	12.77-60.75	Velusamy et al.
		Adriatic Sea	23.5-136	Kozic et al.
		Kuwait	10-25	Bu-Olayou & Thomas
		Greece	5-90	Giannakopoulou & Neofitou
	Mammals	Portugal	18.8-53 ^a	Méndez-Fernandez et al. b
Biphenols	Phytoplankton	Baltic Sea	0.0057-0.24	Staniszewska et al.
	Zooplankton	Baltic Sea	0-0.77	Staniszewska et al.
	Pelecypods	Baltic Sea	0-0.197	Staniszewska et al.
Chlorinated Paraffins	Decapods	China	0.003-1.05 ^a	Ma et al. a
		China	4.77-32.7	Ma et al. b
	Gastropods	China	0.0037-2.2 ^a	Ma et al. a
		China	0.0026-1.05 ^a	Ma et al. a
		China	0.003-2.9 ^a	Ma et al. a
Chlorodane	Fish	California	0.008-2.8	Lyons et al.
ΣDDT	Pelecypods	Korea	0.0001-0.003	Choi, Yang, Hong and Chin
		Massachusetts	0.78 ^b	Subedi, et al.
	Fish	China	0.008-0.04 ^b	Hao et al.
		China	0.097-5.3 ^b	Sun et al.
		Poland	0.006-0.035 ^a	Waszak et al.
		Portugal	0.0007-0.0024	Waszak et al.
	Mammals	California	0.004-2.4 ^b	Lyons et al.
		Brazil	0.000003-0.005 ^b	Santos-Neto et al.
Heptachlor	Fish	Poland	0.00006-0.0001 ^a	Waszak et al.
		Portugal	0.00002-0.00004 ^a	Waszak et al.

Σ HCBD	Fish	Poland Portugal	0.00008-0.0005 ^a 0.00002-0.0001 ^a	Waszak et al. Waszak et al.
Σ HCHe	Pelecypods Mammals	Korea Brazil	0.00009-0.0003 0.000005-0.00016 ^b	Choi,Yang, Hong and Chin Santos-Neto et al.
Methyl siloxanes	Fish	China	5.1 ^a	Hong et al.b
Mirex	Decapods Pelecypods Fish Mammals Birds	China China China Brazil China	0.00064-0.0015 ^a 0.001-0.02 ^a 0.006-0.098 ^a 0.0002-0.024 ^b 0.3-2.3 ^a	Peng et al. Peng et al. Peng et al. Santos-Neto et al. Peng et al.
PAHs	Brown algae Corals	Aegean Sea Taiwan	0.75-5 `0.14-1.7	Apostolopoulou et al. Ko, Chang and Cheng
PAHs cont.	Pelecypods Starfish Fish	Turkey Korea Chukchi Sea China Spain	0.4-589 1.2-1.49 0.65-5.2 0.01-0.94 0.003-0.04	Balcioglu et al. Kim, Hong, Kim and Yang Harvey et al. Li, Lu, Huang, Yang and Ran León et al.
PBDEs	Pelecypods California Fish	Massachusetts 0.001-0.1 China China Poland Portugal Germany Canada China China	277 ^b Dodder et al. 0.0029-0.0078 ^b 0.002-0.12 ^b 0.0002-0.002 ^a 0.0002-0.0007 ^a 0.002-0.0084 ^a 0.004-0.077 ^a 0.0013-0.049 ^b 0.0027-0.94 ^b	Subedi et al. Hao et al. Sun et al. Waszak et al. Waszak et al. Sühring et al. Sühring et al. Zhu et al. Ko, We and Chou
Σ PCBs	Pelecypods Fish Polar Bear Mar. Mammals	Korea Massachusetts China China Poland Portugal California Arctic Ocean Portugal Brazil Arctic Ocean Canada	0.001-0.0035 182-942 ^b 0.014-0.048 ^b 0.006-0.2 ^b 0.0049-0.016 ^a 0.0027-0.0087 ^a 0.22-48.2 ^b 1-8 ^b 2-61.2 0.00002-0.017 ^b 0.1-1.2 ^b 0.001-0.065 ^b	Choi,Yang, Hong and Chin Sibedi et al. Hao et al. Sun et al. Waszak et al. Waszak et al. Lyons et al. Binnington & Wania Méndez-Fernandez et al. a Santos-Neto et al. Binnington & Wania Binnington & Wania
Perfluorinated compounds	Pelecypods	California	0-0.009	Dodder et al.
Pharmaceuticals	Pelecypods	California UK	0.001-0.15 0.004-0.029 ^c	Dodder et al. McEnuff et al.

a=Wet weight

b=Experimental study

c=Lipid weight

d=Polycyclic aromatic hydrocarbons

e=Polybrominated diphenyl ethers